The Competition test a) the with indications binding plasma Estrogens hormone the female Summary on human Immunoassay Intended Number 223 For System English

Elecsys Estradiol III

<table>
<thead>
<tr>
<th>REF</th>
<th>06656021190*</th>
<th>06656021214*</th>
<th>06656021500</th>
</tr>
</thead>
</table>

* Some kits shown may not be available in all countries.

English

System information

For cobas e 411 analyzer: test number 1370
For cobas e 601 and cobas e 602 analyzers: Application Code Number 223

Intended use

Immunoassay for the in vitro quantitative determination of estradiol in human serum and plasma. The electrochemiluminescence immunoassay “ECLIA” is intended for use on Elecsys and cobas e immunoassay analyzers.

Summary

Estrogens are responsible for the development of the secondary female sex characteristics. Together with gestagens they control all the important female reproductive processes.

The biologically most active estrogen is 17β-estradiol. This is a steroid hormone having a molecular weight of 272 daltons.

Estrogens are produced primarily in the ovary (follicle, corpus luteum), but small quantities are also formed in the testes and in the adrenal cortex. During pregnancy, estrogens are mainly formed in the placenta. In human plasma the bulk of estradiol is bound specifically to SHBG (= sex hormone binding globulin) and non-specifically to human serum albumin.

Estrone secretion is biphasic during the menstrual cycle. The determination of estradiol is utilized clinically in the elucidation of fertility disorders in the hypothalamus-pituitary-gonad axis, gynecomastia, estrogen-producing ovarian and testicular tumors. Further clinical indications are the monitoring of fertility therapy and determining the time of ovulation within the framework of in vitro fertilization (IVF).

The Elecsys Estradiol III assay employs a competitive test principle using two monoclonal antibodies specifically directed against 17β-estradiol. Endogenous estradiol released from the sample by mesterolone competes with the added estradiol derivative labeled with a ruthenium complex for the binding sites on the biotinylated antibody.

Test principle

Competition principle. Total duration of assay: 18 minutes.

- 1st incubation: By incubating the sample (25 μL) with two estradiol-specific biotinylated antibodies, immunocomplexes are formed, the amount of which is dependent upon the analyte concentration in the sample.
- 2nd incubation: After addition of streptavidin-coated microparticles and an estradiol derivative labeled with a ruthenium complex, the still-vacant sites of the biotinylated antibodies become occupied, with formation of an antibody-hapten complex. The entire complex becomes bound to the solid phase via interaction of streptavidin.
- The reaction mixture is aspirated into the measuring cell where the microparticles are magnetically captured onto the surface of the electrode. Unbound substances are then removed with ProCell/ProCell M. Application of a voltage to the electrode then induces chemiluminescent emission which is measured by a photomultiplier.
- Results are determined via a calibration curve which is instrument-specifically generated by 2-point calibration and a master curve provided via the reagent barcode or e-barcode.

Reagents - working solutions

The reagent rackpack is labeled as E2 III.

M Streptavidin-coated microparticles (transparent cap), 1 bottle, 6.5 mL:
  Streptavidin-coated microparticles 0.72 mg/mL; preservative.

R1 Anti-estradiol-Ab–biotin (gray cap), 1 bottle, 9 mL:
  Two biotinylated monoclonal anti-estradiol antibodies (rabbit)
  2.5 ng/mL and 4.5 ng/mL; mesterolone 130 ng/mL; MES buffer 50 mmol/L, pH 6.0; preservative.

R2 Estradiol-ester–Ru(bpy)32+ (black cap), 1 bottle, 9 mL:
  Estradiol derivative, labeled with ruthenium complex 4.5 ng/mL; MES buffer 50 mmol/L, pH 6.0; preservative.

b) MES = 2-morpholino-ethane sulfonic acid

Precautions and warnings

For in vitro diagnostic use for health care professionals. Exercise the normal precautions required for handling all laboratory reagents.

Infectious or microbial waste:

Warning: handle waste as potentially biohazardous material. Dispose of waste according to accepted laboratory instructions and procedures.

Environmental hazards:

Apply all relevant local disposal regulations to determine the safe disposal.

Safety data sheet available for professional user on request.

For USA: Caution: Federal law restricts this device to sale by or on the order of a physician.

This kit contains components classified as follows in accordance with the Regulation (EC) No. 1272/2008:

Warning

H317 May cause an allergic skin reaction.

Prevention:

P261 Avoid breathing dust/fume/gas/mist/vapours/spray.

P272 Contaminated work clothing should not be allowed out of the workplace.

P280 Wear protective gloves.

Response:

P333 + P313 If skin irritation or rash occurs: Get medical advice/attention.

P362 + P364 Take off contaminated clothing and wash it before reuse.

Disposal:

P501 Dispose of contents/container to an approved waste disposal plant.

Product safety labeling follows EU GHS guidance.

Contact phone: all countries: +49-621-7590, USA: 1-800-428-2336
Avoid foam formation in all reagents and sample types (specimens, calibrators and controls).

Reagent handling

The reagents in the kit have been assembled into a ready-for-use unit that cannot be separated.

All information required for correct operation read in from the respective reagent barcodes.
Elecsys Estradiol III

Storage and stability
Store at 2-8 °C.
Do not freeze.
Store the Elecsys reagent kit upright in order to ensure complete availability of the microparticles during automatic mixing prior to use.

Stability:

<table>
<thead>
<tr>
<th>Condition</th>
<th>Stability</th>
</tr>
</thead>
<tbody>
<tr>
<td>unopened at 2-8 °C</td>
<td>up to the stated expiration date</td>
</tr>
<tr>
<td>after opening at 2-8 °C</td>
<td>12 weeks</td>
</tr>
<tr>
<td>on the analyzers</td>
<td>8 weeks</td>
</tr>
</tbody>
</table>

Specimen collection and preparation
Only the specimens listed below were tested and found acceptable.
Serum collected using standard sampling tubes or tubes containing separating gel.
Li-heparin, K₂-EDTA and K₂-EDTA plasma as well as plasma separation tubes.

Criterion: Slope 0.9-1.1 + intercept within ± 10 pg/mL + coefficient of correlation ≥ 0.95.
For individual samples recovery within 70-130 % of serum value > 100 pg/mL, recovery of ± 20 pg/mL of serum value ≤ 100 pg/mL.
Stable for 24 hours at 20-25 °C, 2 days at 2-8 °C, 6 months at -20 °C (± 5 °C). Freeze only once.
The samples tested were a selection of samples from various manufacturers, testers may contain differing materials which could affect test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube manufacturer.
Centrifuge samples containing precipitates and frozen samples before performing the assay.
Do not use heat-inactivated samples.
Do not use samples and controls sterilized with azide.

Due to possible evaporation effects, samples, calibrators and controls on the analyzers should be analyzed/measured within 24 hours.
Sample stability claims were established by experimental data by the manufacturer or based on reference literature and only for the temperatures/time frames as stated in the method sheet. It is the responsibility of the individual laboratory to use all available references and/or its own studies to determine specific stability criteria for its laboratory.

Materials provided
See “Reagents – working solutions” section for reagents.

Materials required (but not provided)
- 06656048190, Estradiol III CalSet, for 4 x 1.0 mL
- 11731416190, PreciControl Universal, for 4 x 3.0 mL
- 11731416160, PreciControl Universal, for 4 x 3.0 mL (for USA)
- 03069887190, Diluent MultiAssay, 2 x 16 mL sample diluent
- General laboratory equipment
- cobas e analyzer

Additional materials for cobas e 411 analyzer:
- 11662988122, ProCell, 6 x 380 mL system buffer
- 11662970122, CleanCell, 6 x 380 mL measuring cell cleaning solution
- 11930346122, Elecsys SysWash, 1 x 500 mL washer additive
- 11933159001, Adapter for SysClean
- 11706802001, AssayCup, 60 x 60 reaction cups
- 11706799001, AssayTip, 30 x 120 pipette tips
- 11800507001, Clean-Liner

Additional materials for cobas e 601 and cobas e 602 analyzers:
- 04880340190, ProCell M, 2 x 2 L system buffer
- 04880293190, CleanCell M, 2 x 2 L measuring cell cleaning solution
- 03023141001, PC/CC-Cups, 12 cups to prewarm ProCell M and CleanCell M before use
- 03005712190, ProbeWash M, 12 x 70 mL cleaning solution for run finalization and rinsing during reagent change
- 03004899190, PreClean M, 5 x 600 mL detection cleaning solution
- 12102137001, AssayTip/AssayCup, 48 magazines x 84 reaction cups or pipette tips, waste bags
- 03023150001, WasteLiner, waste bags
- 03027651001, SysClean Adapter M

Additional materials for all analyzers:
- 11298500316, IS8 Cleaning Solution/Elecsys SysClean, 5 x 100 mL system cleaning solution
- 11298500160, IS8 Cleaning Solution/Elecsys SysClean, 5 x 100 mL system cleaning solution (for USA)

Assay
For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator’s manual for analyzer-specific assay instructions.
Resuspension of the microparticles takes place automatically prior to use. Read in the test-specific parameters via the reagent barcode. If in exceptional cases the barcode cannot be read, enter the 15-digit sequence of numbers.

cobas e 601 and cobas e 602 analyzers: PreClean M solution is necessary.

Bring the cooled reagents to approximately 20 °C and place them in the reagent rack (20 °C) of the analyzer. Avoid foam formation. The system automatically regulates the temperature of the reagents and the opening/closing of the bottles.

Calibration
Traceability: This method has been standardized against CRM 6004a via ID-GCMS (isotope dilution-gas chromatography/mass spectrometry).
Every Elecsys reagent set has a barcoded label containing specific information for calibration of the particular reagent lot. The predefined master curve is adapted to the analyzer using the relevant CalSet.
Calibration frequency: Calibration must be performed once per reagent lot using fresh reagent (i.e. not more than 24 hours since the reagent kit was registered on the analyzer).
Calibration interval may be extended based on acceptable verification of calibration by the laboratory.

Renewed calibration is recommended as follows:
- after 1 month (28 days) when using the same reagent lot
- after 7 days (when using the same reagent kit on the analyzer)
- as required: e.g. quality control findings outside the defined limits

Quality control
For quality control, use PreciControl Universal.
In addition, other suitable control material can be used.
Controls for the various concentration ranges should be run individually at least once every 24 hours when the test is in use, once per reagent kit, and following each calibration.
The control intervals and limits should be adapted to each laboratory’s individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.
If necessary, repeat the measurement of the samples concerned.
Follow the applicable government regulations and local guidelines for quality control.
Elecys Estradiol III

Calculation
The analyzer automatically calculates the analyte concentration of each sample (either in pmol/L, pg/mL, ng/L or additionally in nmol/L with cobas e 601 and cobas e 602 analyzers).

Conversion factors:
- pmol/L x 0.272 = pg/mL (ng/L)
- pmol/L x 3.67 = pmol/L
- pmol/L x 0.00367 = nmol/L

Limitations - interference
The assay is unaffected by icterus (bilirubin ≤ 1129 μmol/L or ≤ 66 mg/dL), hemolysis (Hb ≤ 0.621 mmol/L or ≤ 1.0 g/dL), lipemia (triglycerides ≤ 1000 mg/dL) and biotin (≤ 147 nmol/L or ≤ 36 ng/mL).

Criterion: Recovery within ± 10 % of initial value.

Samples should not be taken from patients receiving therapy with high biotin doses (i.e. > 5 mg/day) until at least 8 hours following the last biotin administration.

No interference was observed from rheumatoid factors up to a concentration of 1200 IU/mL.

In vitro tests were performed on 17 commonly used pharmaceuticals. No interference with the assay was found.

Erroneous test results may be obtained from samples taken from patients who have been exposed to vaccines containing rabbit serum or when keeping rabbits as pets animals.

Due to the risk of cross-reactivity, this assay should not be used when monitoring Estradiol levels in patients being treated with Fulvestrant.

Steroid drugs may interfere with this test.

In rare cases, interference due to extremely high titers of antibodies to analyte-specific antibodies, streptavidin or ruthenium can occur. These effects are minimized by suitable test design.

For diagnostic purposes, the results should always be assessed in conjunction with the patient’s medical history, clinical examination and other findings.

Limits and ranges

Limit of Blank, Limit of Detection and Limit of Quantitation

Limit of Blank = 11 pmol/L (3 μg/L)

Limit of Detection = 18.4 pmol/L (5 μg/L)

Limit of Quantitation = 91.8 pmol/L (25 pg/mL) with a total allowable error of ≤ 30 %

A study was performed for Limit of Quantitation using human serum samples diluted and measured in 6 runs over ≥ 3 days on 2 analyzers. At a total allowable error of ≤ 30 % Limit of Quantitation was 61.3 pmol/L (16.7 pg/mL).

The Limit of Blank, Limit of Detection and Limit of Quantitation were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A requirements.

The Limit of Blank is the 95th percentile value from n ≥ 60 measurements of analyte-free samples over several independent series. The Limit of Blank corresponds to the concentration below which analyte-free samples are found with a probability of 95 %.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low concentration samples. The Limit of Detection corresponds to the lowest analyte concentration which can be detected (value above the Limit of Blank with a probability of 95 %).

The Limit of Quantitation is defined as the lowest amount of analyte in a sample that can be accurately quantitated with a total allowable error of ≤ 30 %.

Dilution
Samples with estradiol concentrations above the measuring range can be diluted with Diluent MultiAssay. The recommended dilution is 1:10 (automatically by the analyzers). The concentration of the diluted sample must be > 881 pmol/L (> 240 pg/mL).

After dilution by the analyzers, the software automatically takes the dilution into account when calculating the sample concentration.

The endogenous analyte concentration of the diluted (< 220 pmol/L or < 60.0 pg/mL) is not taken into account for dilutions above the measuring range.

Expected values
The expected ranges were determined by testing specimens drawn from 150 apparently healthy males, 142 apparently healthy, post-menopausal women over the age of 50, and from 412 apparently healthy pregnant women between the ages of 18 and 50 (136 in the first trimester, 140 in the second trimester, and 136 in the third trimester). The expected range for healthy women was determined by collecting blood at multiple time points of one menstrual cycle from 85 apparently healthy subjects with a natural menstrual cycle that were not taking any hormonal contraceptives. A menstrual cycle was defined as the phase between two subsequent menstrual bleedings. Cycle length (29 days) and day of ovulation (day 15) were standardized to account for variation in cycle length within the study population and to enable determination of expected values for further sub-phases. Only ovulatory menstrual cycles were used for value analysis. The following ranges were obtained:

Test subjects N 2.5th percentile Median 97.5th percentile

<table>
<thead>
<tr>
<th>Analyte</th>
<th>pmol/L</th>
<th>pmol/L</th>
<th>pmol/L</th>
</tr>
</thead>
<tbody>
<tr>
<td>Healthy men</td>
<td>150</td>
<td>13.4 (90 % CI)</td>
<td>30.9 (90 % CI)</td>
</tr>
<tr>
<td>Healthy postmenopausal women</td>
<td>142</td>
<td>&lt; 18.4 (18.4-19.2)</td>
<td>18.4 (18.4-19.2)</td>
</tr>
<tr>
<td>Healthy pregnant women</td>
<td>136</td>
<td>3.2 (19.1-135)</td>
<td>3.2 (19.1-135)</td>
</tr>
</tbody>
</table>

* CI = confidence interval

Healthy women

<table>
<thead>
<tr>
<th>Cycle Phase</th>
<th>N</th>
<th>5th percentile</th>
<th>Median</th>
<th>95th percentile</th>
</tr>
</thead>
<tbody>
<tr>
<td>Follicular</td>
<td>85</td>
<td>91 (90 % CI)</td>
<td>166 (90 % CI)</td>
<td>332 (90 % CI)</td>
</tr>
<tr>
<td>Ovulation</td>
<td>81</td>
<td>222 (19.5-283)</td>
<td>757 (667-944)</td>
<td>1959 (1588-3338)</td>
</tr>
<tr>
<td>Luteal</td>
<td>85</td>
<td>222 (19.5-283)</td>
<td>412 (300-488)</td>
<td>854 (760-1334)</td>
</tr>
</tbody>
</table>

Healthy women (sub-phase regulation)

<table>
<thead>
<tr>
<th>Sub-phase</th>
<th>N</th>
<th>5th percentile</th>
<th>Median</th>
<th>95th percentile</th>
</tr>
</thead>
<tbody>
<tr>
<td>Early follicular</td>
<td>78</td>
<td>75.5 (18.4-78.5)</td>
<td>126 (120-135)</td>
<td>291 (192-283)</td>
</tr>
<tr>
<td>Intermediate follicular</td>
<td>83</td>
<td>95.6 (19.1-114)</td>
<td>172 (159-180)</td>
<td>294 (262-689)</td>
</tr>
<tr>
<td>Late follicular</td>
<td>84</td>
<td>182 (84-215)</td>
<td>464 (424-519)</td>
<td>858 (71-1337)</td>
</tr>
</tbody>
</table>

* N = number of patients contributing to the data in this menstrual cycle phase (not number of samples); differences in N per phase are due to cycle standardization procedure

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Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

Specific performance data
Representative performance data on the analyzers are given below. Results obtained in individual laboratories may differ.

Precision
Precision was determined using Elecsys reagents, samples and controls in a protocol (EPS-A2) of the CLSI (Clinical and Laboratory Standards Institute): 2 runs per day in duplicate each for 21 days (n = 84). The following results were obtained:

<table>
<thead>
<tr>
<th>Test</th>
<th>cobas e 411 analyzer</th>
<th>cobas e 601 and cobas e 602 analyzers</th>
<th>cobas e 411 analyzer</th>
</tr>
</thead>
<tbody>
<tr>
<td>50%</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Luteal</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Postmenopausal women</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Healthy men</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Healthy pregnant women</td>
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<td></td>
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<tr>
<td>Early follicular</td>
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<td></td>
<td></td>
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<tr>
<td>Intermediate follicular</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>Late follicular</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ovulation</td>
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<tr>
<td>Early luteal</td>
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<td></td>
</tr>
<tr>
<td>Intermediate luteal</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>Late luteal</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>Cycle Phase</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cycle Sub-Phase</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Follicular</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Luteal</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Healthy women</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Elecsys Estradiol III

<table>
<thead>
<tr>
<th>Substance</th>
<th>n. d.</th>
</tr>
</thead>
<tbody>
<tr>
<td>17-Hydroxyprogesterone</td>
<td></td>
</tr>
<tr>
<td>d)</td>
<td>Substance added in a concentration of 0.2 μg/mL:</td>
</tr>
<tr>
<td>17α-Ethinylestradiol</td>
<td>0.334</td>
</tr>
<tr>
<td>Cortisol</td>
<td>0.003</td>
</tr>
<tr>
<td>Cortisone</td>
<td>0.001</td>
</tr>
<tr>
<td>Tamoxifen</td>
<td>0.001</td>
</tr>
<tr>
<td>e)</td>
<td>Substance added in a concentration of 0.25 μg/mL:</td>
</tr>
<tr>
<td>Chlormephan</td>
<td>0.001</td>
</tr>
<tr>
<td>f)</td>
<td>Substance added in a concentration of 1.0 μg/mL:</td>
</tr>
<tr>
<td>Prednisolone</td>
<td>n. d.</td>
</tr>
<tr>
<td>g)</td>
<td>Substance added in a concentration of 10 μg/mL:</td>
</tr>
<tr>
<td>Danazol</td>
<td>0.001</td>
</tr>
<tr>
<td>DHEA-S</td>
<td>n. d.</td>
</tr>
<tr>
<td>Mesterolone</td>
<td>n. d.</td>
</tr>
<tr>
<td>Testosterone</td>
<td>n. d.</td>
</tr>
<tr>
<td>5α-Dihydrotestosterone (DHT)</td>
<td>n. d.</td>
</tr>
<tr>
<td>5-Androsten-3β,17β-diol</td>
<td>n. d.</td>
</tr>
</tbody>
</table>

Method comparison
A comparison of the Elecsys Estradiol III assay (y) with ID-GC/MS (x) using clinical samples gave the following correlations (pg/mL):

Number of samples measured: 25

Passing/Bablok

y = 0.993x + 1.26
y = 1.00x + 2.07
τ = 0.987
r = 0.999

The sample concentrations were between 37.4 and 10768 pmol/L (10.2 and 2934 pg/mL) (ID-GC-MS concentrations).

Analytical specificity
For the Elecsys Estradiol III assay, the following cross-reactivities were found (in %):

a) Substance added in a concentration of 0.001 µg/mL:
- α-Hydroxy-Estradiol: 74.1

b) Substance added in a concentration of 0.01 µg/mL:
- 4-Hydroxyestradiol: 0.754

References

For further information, please refer to the appropriate operator's manual for the analyzer concerned, the respective application sheets and the Method Sheets of all necessary components (if available in your country).

A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Any serious incident that has occurred in relation to the device shall be reported to the manufacturer and the competent authority of the Member State in which the user and/or the patient is established.

Symbols
Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard (for USA: see dialog.roche.com for definition of symbols used):

- CONTENT: Contents of kit
- SYSTEM: Analyzers/Instruments on which reagents can be used
- REAGENT: Reagent
- CALIBRATOR: Calibrator
- VOLUME: Volume for reconstitution
- GTIN: Global Trade Item Number
FOR US CUSTOMERS ONLY: LIMITED WARRANTY

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