



Elecsys Anti-HEV IgG

cobas®

REF			SYSTEM
09044647190	09044647500	300	cobas e 402 cobas e 801

English

System information

Short name	ACN (application code number)
AHEVIGG	10222

Intended use

Immunoassay for the in vitro quantitative determination of IgG antibodies to the hepatitis E virus (HEV) in human serum and plasma. The Elecsys Anti-HEV IgG assay is used as an aid to detect a recent or past HEV infection.

The electrochemiluminescence immunoassay "ECLIA" is intended for use on **cobas e** immunoassay analyzers.

Summary

HEV is the etiological agent of hepatitis E and is regarded as an emerging pathogen of global public health concern.^{1,2} HEV infection is an important cause of both acute and chronic hepatitis, being responsible for more than 50 % of the cases of acute hepatitis in endemic countries.^{3,4,5} A global burden of disease study estimated that HEV genotypes 1 and 2 account for approximately 20.1 million HEV infections, 3.4 million symptomatic cases, 70000 deaths, and 3000 stillbirths annually.^{6,7}

HEV is an icosahedral, non-enveloped, positive-sense, single-stranded RNA virus with a diameter of 27-34 nm. The RNA genome of 7.2 kb has 3 open reading frames (ORFs): ORF1, encoding non-structural proteins involved in viral replication; ORF2, encoding a viral capsid protein important for virion assembly and immunogenicity; and ORF3, encoding a protein essential for virus release.^{1,4,5,8,9,10,11}

HEV, which is classified as species *Paslahepevirus balayani* in the family *Hepeviridae*, genus *Paslahepevirus*, includes 8 genotypes, of which HEV 1-4 are the most frequently detected globally. HEV-1 and HEV-2 infect only humans, whereas HEV-3 and HEV-4 infect humans but also several animal species such as pig, boar, rabbit, and deer.^{5,12,13,14,15,16,17} HEV-1 and HEV-2 are predominant in Africa and Asia,^{1,12,18} while HEV-3 and HEV-4 are seen largely in the developed world.^{1,12,18,19}

HEV-1 and HEV-2 are mainly transmitted by the fecal-oral route, principally via contaminated water, in areas of poor sanitation,²⁰ while HEV-3 and HEV-4 are transmitted zoonotically by direct contact with infected animals^{1,8,20} or by consumption of contaminated food, primarily undercooked pork meat.^{8,11,19,20} Vertical transmission from mother to fetus is one of the main transmission routes for HEV that causes premature birth and perinatal mortality.^{21,22,23,24} In several countries, occasional transmission of HEV infection (HEV-3) through blood transfusion has been reported.^{1,3,4}

HEV infection usually causes a mild or subclinical infection with a self-limiting illness that lasts from 2 to 6 weeks.^{19,25} Symptomatic hepatitis E is similar to other acute hepatitis infections (non-specific prodromal symptoms of fatigue, nausea, vomiting as well as jaundice and elevated liver enzymes).¹⁹ Testing for hepatitis E is recommended in all patients presenting with symptoms consistent with acute hepatitis, patients with unexplained flares of chronic hepatitis as well as in all immunocompromised patients with unexplained abnormal liver function tests.^{11,26} An important differential diagnosis of acute hepatitis E is drug-induced liver injury.²⁶

Mainly in immunocompromised patients, acute hepatitis can progress to chronic hepatitis (approximately 1-2 % of cases), cirrhosis, liver failure and acute-on-chronic liver failure.^{11,19,27,28} Pregnant women, particularly during the third trimester of pregnancy, are more vulnerable to HEV infections leading to severe clinical outcomes such as fulminant hepatic failure, birth complications, and a significant mortality rate as high as 10-30 %.^{9,13,19,24,29,30} Therefore, antenatal screening for HEV antibodies should be considered.³¹ Other high-risk populations are immunocompromised patients (especially transplant organ recipients), patients with underlying liver conditions, and elderly people.^{4,5,9,29,30,32} Acute and chronic HEV infection have also been associated with extra-hepatic manifestations, especially neurological and renal disorders.^{11,19,26,32} In the aforementioned clinical settings, it is imperative to test for HEV infection upon suspicion of viral hepatitis.^{33,34}

Around 3 weeks post-infection, HEV RNA becomes detectable in blood and stool, with viremia lasting approximately 3-6 weeks, and shedding of virus in stool for approximately 4-6 weeks.²⁶ IgM antibodies against the HEV capsid protein are detectable in serum after 1-4 weeks for up to 6-9 months post-infection, and are a key marker of recent or current infection.^{4,5,11,26,30,32,33}

Anti-HEV IgG antibodies appear around the same time as or soon after anti-HEV IgM antibodies. They are an indicator of recent or past infection and usually persist for several years.^{4,15,26,30,32,35}

Acute HEV infection can be diagnosed by the detection of anti-HEV antibodies (IgG, IgM, or both) in serum or plasma, in combination with testing for HEV RNA. Serological testing alone relies upon the combined detection of anti-HEV IgM and rising anti-HEV IgG titers.²⁶ Measuring rising anti-HEV IgG titers may help in diagnosis of HEV infection in situations with poor anti-HEV IgM response.⁹ The presence of anti-HEV IgG determines past infection.^{19,26,36,37} In addition, anti-HEV IgG testing is considered useful in seroprevalence studies. Moreover, in the future, testing for anti-HEV IgG titers may become useful for determining the effectiveness of HEV vaccines.¹

Acute HEV infection does not usually require antiviral therapy.^{26,38} Supportive care is the mainstay of treatment.³⁹ However, hospitalization is required for people with fulminant hepatitis and should also be considered for symptomatic pregnant women,^{20,38} and ribavirin treatment may be considered in cases of severe acute hepatitis E, acute-on-chronic liver failure, or immunocompromised patients with chronic hepatitis E.^{20,26} In certain situations, interferon or a reduction of immunosuppressive therapy have also been used successfully.^{20,26}

The Elecsys Anti-HEV IgG assay uses recombinant proteins based on structural domains of HEV ORF2 (genotype 1 and 3) as antigens in a double-antigen sandwich assay format for the quantitative determination of IgG antibodies to HEV. The quantitative assay result is also qualitatively interpreted for the detection of IgG antibodies to HEV. Quantitative measurement of IgG antibodies to HEV is intended as an aid, in conjunction with other laboratory results and clinical information, in the diagnosis of acute HEV infection (e.g. by determining rising IgG titers during acute infection, in combination with detection of IgM antibodies to HEV or HEV RNA), as part of the differential diagnosis of acute hepatitis to enable timely initiation of medical interventions; in assessing the immune status to HEV; in estimating the risk of HEV reinfection; or in detecting past HEV infections in seroepidemiological studies. Testing for HEV infection, including anti-HEV IgG antibodies, is also indicated in pregnant women.

Elecsys Anti-HEV IgG is not intended for use as a first-line screening test for donors of blood or blood products or human cells, tissues, and cellular and tissue-based products (HCT/PS).

Test principle

Double-antigen sandwich principle. Total duration of assay: 18 minutes.

- 1st incubation: 12 µL of sample, biotinylated recombinant HEV antigens, and HEV recombinant antigens labeled with a ruthenium complex^{a)} form a sandwich complex.
- 2nd incubation: After addition of streptavidin-coated microparticles, the complex becomes bound to the solid phase via interaction of biotin and streptavidin.
- The reaction mixture is aspirated into the measuring cell where the microparticles are magnetically captured onto the surface of the electrode. Unbound substances are then removed with ProCell II M. Application of a voltage to the electrode then induces chemiluminescent emission which is measured by a photomultiplier.
- Results are determined via a calibration curve which is instrument-specifically generated by 2-point calibration and a master curve provided via the **cobas** link.

a) Tris(2,2'-bipyridyl)ruthenium(II)-complex (Ru(bpy)₃²⁺)

Reagents - working solutions

The **cobas e** pack (M, R1, R2) is labeled as AHEVIGG.

- M Streptavidin-coated microparticles, 1 bottle, 16.0 mL:
Streptavidin-coated microparticles 0.72 mg/mL; preservative.

Elecsys Anti-HEV IgG

- R1 HEVAg (genotype 1, genotype 3)~biotin, 1 bottle, 16.7 mL:
Recombinant biotinylated HEVAg (genotype 1, genotype 3)
≥ 0.14 mg/L; TRIS buffer 50 mmol/L, pH 7.2; preservative.
- R2 HEVAg (genotype 1, genotype 3)~Ru(bpy)₃²⁺, 1 bottle, 16.7 mL:
Recombinant HEVAg (genotype 1, genotype 3), labeled with
ruthenium complex, ≥ 0.14 mg/L; TRIS buffer 50 mmol/L, pH 7.2;
preservative.
- AHEVIGG Cal1 Negative calibrator 1, 1 bottle of 1.0 mL:
Human serum; preservative.
- AHEVIGG Cal2 Positive calibrator 2, 1 bottle of 1.0 mL:
Anti-HEV IgG (human) approximately 5 U/mL in human
serum; preservative.

Precautions and warnings

For in vitro diagnostic use for health care professionals. Exercise the normal precautions required for handling all laboratory reagents.

Infectious or microbial waste:

Warning: handle waste as potentially biohazardous material. Dispose of waste according to accepted laboratory instructions and procedures.

Environmental hazards:

Apply all relevant local disposal regulations to determine the safe disposal.

Safety data sheet available for professional user on request.

This kit contains components classified as follows in accordance with the Regulation (EC) No. 1272/2008:



Warning

- H317 May cause an allergic skin reaction.

Prevention:

- P261 Avoid breathing mist or vapours.
- P272 Contaminated work clothing should not be allowed out of the workplace.
- P280 Wear protective gloves.

Response:

- P333 + P313 If skin irritation or rash occurs: Get medical advice/attention.
- P362 + P364 Take off contaminated clothing and wash it before reuse.

Disposal:

- P501 Dispose of contents/container to an approved waste disposal plant.

Product safety labeling follows EU GHS guidance.

Contact phone: all countries: +49-621-7590

All human material should be considered potentially infectious. All products derived from human blood are prepared exclusively from the blood of donors tested individually and shown to be free from HBsAg and antibodies to HCV and HIV. The testing methods use assays that have been approved by the FDA or that are in compliance with the legal rules applicable to placing in vitro diagnostic medical devices for human use on the market in the European Union.

However, as no testing method can rule out the potential risk of infection with absolute certainty, the material should be handled with the same level of care as a patient specimen. In the event of exposure, the directives of the responsible health authorities should be followed.^{40,41}

Avoid foam formation in all reagents and sample types (specimens, calibrators and controls).

Reagent handling

The reagents (M, R1, R2) in the kit are ready-for-use and are supplied in **cobas e** packs.

Calibrators:

The calibrators are supplied ready-for-use in bottles compatible with the system.

Unless the entire volume is necessary for calibration on the analyzer, transfer aliquots of the ready-for-use calibrators into empty snap-cap bottles (CalSet Vials). Attach the supplied labels to these additional bottles. Store the aliquots at 2-8 °C for later use.

Perform **only one** calibration procedure per aliquot.

All information required for correct operation is available via the **cobas** link.

Storage and stability

Store at 2-8 °C.

Do not freeze.

Store the **cobas e** pack **upright** in order to ensure complete availability of the microparticles during automatic mixing prior to use.

Stability of the cobas e pack:	
unopened at 2-8 °C	up to the stated expiration date
on the analyzers	16 weeks

Stability of the calibrators:	
unopened at 2-8 °C	up to the stated expiration date
after opening at 2-8 °C	16 weeks
on the analyzers at 20-25 °C	use only once

Store calibrators **upright** in order to prevent the calibrator solution from adhering to the snap-cap.

Specimen collection and preparation

Only the specimens listed below were tested and found acceptable.

Serum collected using standard sampling tubes or tubes containing separating gel.

Li-heparin, Na-heparin, K₂-EDTA, K₃-EDTA and Na-citrate plasma.

Criterion for Li-heparin, Na-heparin, K₂-EDTA and K₃-EDTA plasma: slope 1.00 ± 0.10 + bias at 0.150 U/mL ± 15 %.

For native samples collected in sodium citrated plasma: slope 0.84 ± 0.1.

Sampling devices containing liquid anticoagulants have a dilution effect resulting in lower values (U/mL) for individual patient specimens. In order to minimize dilution effects it is essential that respective sampling devices are filled completely according to manufacturer's instructions.

Stable for 7 days at 20-25 °C, 14 days at 2-8 °C, 3 months at -20 °C (± 5°C). The samples may be frozen 5 times.

The sample types listed were tested with a selection of sample collection tubes or systems that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube/collection system manufacturer.

Specimens should not be subsequently altered with additives (e.g. biocides, anti-oxidants or substances that could possibly change the pH or ionic strength of the sample) in order to avoid erroneous findings.

Centrifuge samples containing precipitates and thawed samples before performing the assay.

Do not use heat-inactivated samples.

Ensure the samples and calibrators are at 20-25 °C prior to measurement.

Due to possible evaporation effects, samples and calibrators on the analyzers should be analyzed/measured within 2 hours.

The performance of the Elecsys Anti-HEV IgG assay has not been established with body fluids other than serum and plasma.

Materials provided

See "Reagents – working solutions" section for reagents.

- 2 x 4 bottle labels

Elecsys Anti-HEV IgG



Materials required (but not provided)

- REF 09044655190, PreciControl Anti-HEV IgG, 16 x 1.3 mL
- REF 11776576322, CalSet Vials, 2 x 56 empty snap-cap bottles
- REF 07299001190, Diluent Universal, 36 mL sample diluent
- General laboratory equipment

cobas e analyzer

Additional materials for **cobas e 402** and **cobas e 801** analyzers:

- REF 06908799190, ProCell II M, 2 x 2 L system solution
- REF 04880293190, CleanCell M, 2 x 2 L measuring cell cleaning solution
- REF 07485409001, Reservoir Cup, 8 cups to supply ProCell II M and CleanCell M
- REF 06908853190, PreClean II M, 2 x 2 L wash solution
- REF 05694302001, Assay Tip/Assay Cup tray, 6 magazines x 6 magazine stacks x 105 assay tips and 105 assay cups, 3 wasteliners
- REF 07485425001, Liquid Flow Cleaning Cup, 2 adaptor cups to supply ISE Cleaning Solution/Elecsys SysClean for Liquid Flow Cleaning Detection Unit
- REF 07485433001, PreWash Liquid Flow Cleaning Cup, 1 adaptor cup to supply ISE Cleaning Solution/Elecsys SysClean for Liquid Flow Cleaning PreWash Unit
- REF 11298500316, ISE Cleaning Solution/Elecsys SysClean, 5 x 100 mL system cleaning solution

Assay

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.

Resuspension of the microparticles takes place automatically prior to use.

Place the cooled (stored at 2-8 °C) **cobas e** pack on the reagent manager. Avoid foam formation. The system automatically regulates the temperature of the reagents and the opening/closing of the **cobas e** pack.

Calibrators:

Place the calibrators in the sample zone.

Read in all the information necessary for calibrating the assay.

Calibration

Traceability: This method has been standardized against the WHO Reference Reagent for Hepatitis E Virus Antibody (NIBSC code: 95/584).

The predefined master curve is adapted to the analyzer using AHEVIGG Cal1 and AHEVIGG Cal2.

Calibration frequency: Calibration must be performed once per reagent lot using AHEVIGG Cal1, AHEVIGG Cal2 and fresh reagent (i.e. not more than 24 hours since the **cobas e** pack was registered on the analyzer).

Calibration interval may be extended based on acceptable verification of calibration by the laboratory.

Renewed calibration is recommended as follows:

- after 12 weeks when using the same reagent lot
- after 28 days when using the same **cobas e** pack on the analyzer
- as required: e.g. quality control findings outside the defined limits

Quality control

For quality control, use PreciControl Anti-HEV IgG.

Controls for the various concentration ranges should be run individually at least once every 24 hours when the test is in use, once per **cobas e** pack, and following each calibration.

The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

If necessary, repeat the measurement of the samples concerned.

Follow the applicable government regulations and local guidelines for quality control.

Calculation

The analyzer automatically calculates the analyte concentration of each sample in U/mL.

Interpretation of the results

Numeric result	Result message ^{b)}	Interpretation
< 0.15 U/mL	Non-reactive	Negative for anti-HEV IgG
≥ 0.15 U/mL	Reactive	Positive for anti-HEV IgG

b) A qualitative result message is displayed for results within the measuring range only.

Note: Due to the diversity of the antibodies, the measured anti-HEV IgG value can vary depending on the testing procedure used. Results obtained from a single sample using tests from different manufacturers can therefore differ.

Limitations - interference

The effect of the following endogenous substances and pharmaceutical compounds on assay performance was tested. Interferences were tested up to the listed concentrations and no impact on results was observed.

Endogenous substances

Compound	Concentration tested
Bilirubin	≤ 1129 μmol/L or ≤ 66 mg/dL
Hemoglobin	≤ 0.621 mmol/L or ≤ 1000 mg/dL
Intralipid	≤ 2000 mg/dL
Biotin	≤ 4912 nmol/L or ≤ 1200 ng/mL
Rheumatoid factors	≤ 1200 IU/mL
Albumin	≤ 70 g/L
IgG	≤ 70 g/L
IgA	≤ 16 g/L
IgM	≤ 10 g/L

Criterion: For concentrations of 0.05-0.2 U/mL, the deviation is ≤ 0.02 U/mL. For concentrations > 0.2-25 U/mL, the deviation is ≤ 10 %.

No false negative results due to a high-dose hook effect were found with the Elecsys Anti-HEV IgG assay but occurrence of high-dose hook effect cannot be completely excluded.

Pharmaceutical substances

In vitro tests were performed on 17 commonly used pharmaceuticals. No interference with the assay was found.

In addition, the following special drugs used in hepatitis E therapy were tested. No interference with the assay was found.

Special drugs

Drug	Concentration tested mg/L
Peginterferon alfa-2a	≤ 0.108
Ribavirin	≤ 720
Sofosbuvir	≤ 240

Drug interferences are measured based on recommendations given in CLSI guidelines EP07 and EP37 and other published literature. Effects of concentrations exceeding these recommendations have not been characterized.

In rare cases, interference due to extremely high titers of antibodies to immunological components, streptavidin or ruthenium can occur. These effects are minimized by suitable test design.

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

Results may remain negative during and after HEV infection in immunocompromised patients.

Limits and ranges

Measuring range

Elecsys Anti-HEV IgG



0.05-25 U/mL (defined by the Limit of Quantitation and the maximum of the master curve). Values below the Limit of Quantitation are reported as < 0.05 U/mL. Values above the measuring range are reported as > 25 U/mL (or up to 2500 U/mL for 100-fold diluted samples).

Lower limits of measurement

Limit of Blank, Limit of Detection and Limit of Quantitation

Limit of Blank = 0.01 U/mL

Limit of Detection = 0.025 U/mL

Limit of Quantitation = 0.05 U/mL

The Limit of Blank, Limit of Detection and Limit of Quantitation were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A2 requirements.

The Limit of Blank is the 95th percentile value from $n \geq 60$ measurements of analyte-free samples over several independent series. The Limit of Blank corresponds to the concentration below which analyte-free samples are found with a probability of 95 %.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low concentration samples. The Limit of Detection corresponds to the lowest analyte concentration which can be detected (value above the Limit of Blank with a probability of 95 %).

The Limit of Quantitation is defined as the lowest amount of analyte in a sample that can be accurately quantified with a CV of ≤ 20 %. It has been determined using samples with low concentration of anti-HEV IgG.

Dilution

Samples with anti-HEV IgG concentrations above the measuring range can be diluted with Diluent Universal. The recommended dilution is 1:100 (either automatically by the analyzers or manually). The concentration of the diluted sample must be ≥ 0.150 U/mL.

After manual dilution, multiply the result by the dilution factor.

After dilution by the analyzers, the software automatically takes the dilution into account when calculating the sample concentration.

Note: Antibodies to HEV are heterogeneous. In some isolated cases, this may lead to non-linear dilution behavior.

Specific performance data

Representative performance data on the analyzers are given below. Results obtained in individual laboratories may differ.

Precision

Precision was determined using Elecsys reagents, samples and controls in a protocol (EP05-A3) of the CLSI (Clinical and Laboratory Standards Institute): 2 runs per day in duplicate each for 21 days ($n = 84$). The following results were obtained:

cobas e 402 and cobas e 801 analyzers					
		Repeatability		Intermediate precision	
Sample	Mean U/mL	SD U/mL	CV %	SD U/mL	CV %
HS ^{c)} 1	0.0690	0.000895	1.3	0.000979	1.4
HS 2	0.138	0.00125	0.9	0.00144	1.0
HS 3	0.155	0.00162	1.0	0.00207	1.3
HS 4	1.43	0.0152	1.1	0.0178	1.2
HS 5	12.6	0.126	1.0	0.140	1.1
HS 6	21.3	0.212	1.0	0.263	1.2
PC ^{d)} Anti-HEV IgG 1	0.104	0.00101	1.0	0.00123	1.2
PC Anti-HEV IgG 2	0.355	0.00267	0.8	0.00423	1.2

c) HS = human sample (serum/plasma)

d) PC = PreciControl

Analytical specificity

No cross-reactions with samples containing antibodies against HAV, HBV, HCV, HIV, EBV, HSV, Rubella or CMV, or samples from autoimmune diseases (AMA and ANA) were observed. Measurements were performed

at each of the disease states listed above using ≥ 5 serum or plasma samples.

Seroconversion sensitivity and titer development

Seroconversion sensitivity of the Elecsys Anti-HEV IgG assay was shown by testing 9 commercial seroconversion panels in comparison to 5 other registered anti-HEV IgG assays. The Elecsys Anti-HEV IgG assay detected anti-HEV IgG in 84 out of a total of 112 panel members,⁹⁾ while the comparison assays detected 76 (+ 1 borderline, assay A), 80 (assay B), 79 (+ 1 borderline, assay C), 81 (assay D), and 80 (assay E) panel members, respectively.

In addition, anti-HEV IgG titer assessment indicated rising antibody titers in the seroconversion panels over time. The table below shows the titer development over sequential bleeds from the last bleed before the first Elecsys Anti-HEV IgG reactive result and up to 4 bleeds after.

Panel	AHEVIGG, U/mL (days since 1 st bleed)					
	Last AHEV-IGG non-reactive	AHEVIGG-reactive bleed				
		1 st	2 nd	3 rd	4 th	5 th
Panel 1 SCP-HEV-001b	0.103 (38 d)	1.89 (41 d)	5.39 (46 d)	8.97 (49 d)	16.2 (53 d)	58.2 (77 d)
Panel 2 SCP-HEV-002b	0.000173 (25 d)	0.262 (35 d)	6.00 (42 d)	31.4 (68 d)	60.4 (84 d)	n.a. ^{e)}
Panel 3 SCP-HEV-003a	n.a.	0.315 (0 d)	0.291 (3 d)	7.15 (18 d)	13.0 (25 d)	101 (39 d)
Panel 4 SCP-HEV-004a	n.a.	4.80 (0 d)	18.9 (11 d)	15.7 (14 d)	17.5 (18 d)	22.2 (22 d)
Panel 5 SCP-HEV-005b	0.0179 (21 d)	3.61 (28 d)	4.10 (35 d)	7.40 (42 d)	13.0 (49 d)	55.9 (63 d)
Panel 6 SCP-HEV-006b	0.0643 (46 d)	1.25 (50 d)	0.907 (56 d)	23.7 (105 d)	24.1 (108 d)	26.7 (112 d)
Panel 7 SCP-HEV-007a	0.0579 (28 d)	0.298 (32 d)	0.604 (39 d)	2.71 (46 d)	4.29 (52 d)	6.86 (59 d)
Panel 8 SCP-HEV-008a	0.00130 (0 d)	15.8 (46 d)	19.0 (49 d)	22.8 (54 d)	33.9 (61 d)	33.2 (64 d)
Panel 9 SCP-HEV-009a	0.000430 (21 d)	27.2 (59 d)	24.4 (63 d)	46.5 (80 d)	38.7 (84 d)	34.8 (105 d)

e) n.a. = not applicable

f) Samples not tested according to the vendor data sheet were excluded.

Relative sensitivity

Performance of the Elecsys Anti-HEV IgG assay was assessed by testing a total of 596 samples at 3 different study sites in Europe. 440 samples from patients with presumed acute HEV infection and 156 samples from patients recovered from a hepatitis E infection were measured with the Elecsys Anti-HEV IgG assay and 3 commercially available anti-HEV IgG assays. Samples from patients with presumed acute hepatitis E infection included 252 samples from Europe (endemic for HEV genotype 3) and 188 samples from Vietnam and Bangladesh (endemic for HEV genotype 1).

Additionally, 50 samples were measured at 1 study site in China (confirmed genotype 4) with the Elecsys Anti-HEV IgG assay and 3 anti-HEV IgG assays available in China.

Samples were considered positive if the result was reactive in all of the comparator assays.

Cohort	N	Elecsys Anti-HEV IgG assay reactive	Confirmed anti-HEV IgG positive samples	Sensitivity, % (95 % CI ⁹⁾)
Recovered from a HEV infection	156	141	141	100 (97.4-100)

Elecsys Anti-HEV IgG



Cohort	N	Elecsys Anti-HEV IgG assay reactive	Confirmed anti-HEV IgG positive samples	Sensitivity, % (95 % CI ^{g)})
Presumed acute HEV infection	440	375	380	98.7 (97.0-99.6)
Presumed acute HEV infection (China)	50	48	48	100 (92.6-100)

g) CI = confidence interval, 2-sided

Relative specificity

A total of 8011 samples from blood donors (n = 5040), diagnostic routine (n = 2427) and pregnant women (n = 544) were tested at 4 centers in Europe with the Elecsys Anti-HEV IgG assay and 3 commercially available anti-HEV IgG assays. Samples were considered negative for anti-HEV IgG if they were non-reactive in 2 out of 3 comparator assays. Discrepant Elecsys Anti-HEV IgG reactive samples were resolved using an in-house neutralization assay based on an alternative recombinant HEV antigen (HyTest Ltd). Samples were considered positive if, in presence of the neutralizing HEV antigen, they showed a significantly lower anti-HEV IgG concentration in the Elecsys Anti-HEV IgG assay.

Cohort	Elecsys Anti-HEV IgG assay non-reactive	Anti-HEV IgG comparator assays non-reactive	Discrepant positive samples (confirmed reactive after neutralization / discrepant reactive samples resolved using neutralization)	Confirmed anti-HEV IgG negative samples	Specificity, % (95 % CI)
Blood donors (n = 5040)	4051	4397	4 (340/344) ^{h)}	4055	99.9 (99.7-100)
Diagnostic routine ⁱ⁾ (n = 2427)	1425	1754	9 (320/329)	1434	99.4 (98.8-99.7)
Pregnant women (n = 544)	434	455	1 (20/21)	435	99.8 (98.7-100)

h) 2 discrepant positive samples were excluded because of missing neutralization results.

i) suspected for viral hepatitis

References

- Khuroo MS, Khuroo MS, Khuroo NS. Hepatitis E: Discovery, global impact, control and cure. *World J Gastroenterol* 2016;22(31):7030-7045.
- Webb GW, Dalton HR. Hepatitis E: an underestimated emerging threat. *Ther Adv Infect Dis* 2019;6:2049936119837162.
- Pérez-Gracia MT, Suay B, Mateos-Lindemann ML. Hepatitis E: an emerging disease. *Infect Genet Evol* 2014;22:40-59.
- Al-Sadeq DW, Majdalawieh AF, Mesleh AG, et al. Laboratory challenges in the diagnosis of hepatitis E virus. *J Med Microbiol* 2018;67(4):466-480.
- Li P, Liu J, Li Y, et al. The global epidemiology of hepatitis E virus infection: A systematic review and meta-analysis. *Liver Int* 2020;40(7):1516-1528.
- World Health Organization (WHO). Hepatitis E vaccine: WHO position paper. *Wkly Epidemiol Rec* 2015;18:185-200.
- Rein DB, Stevens GA, Theaker J, et al. The global burden of hepatitis E genotypes 1 and 2 in 2005. *Hepatology* 2012;55(4):988-997.
- Izopet J, Tremieux P, Marion O, et al. Hepatitis E virus infections in Europe. *J Clin Virol* 2019;120:20-26.
- Aggarwal R. Diagnosis of hepatitis E. *Nat Rev Gastroenterol Hepatol* 2013;10(1):24-33.
- Blasco-Perrin H, Abravanel F, Blasco-Baque V, et al. Hepatitis E, the neglected one. *Liver Int* 2016;36 Suppl 1:130-134.
- Lhomme S, Legrand-Abravanel F, Kamar N, et al. Screening, diagnosis and risks associated with Hepatitis E virus infection. *Expert Rev Anti Infect Ther* 2019;17(6):403-418.
- Goel A, Aggarwal R. Hepatitis E: Epidemiology, Clinical Course, Prevention, and Treatment. *Gastroenterol Clin North Am* 2020;49(2):315-330.
- Raji YE, Toung OP, Taib NM, et al. Hepatitis E Virus: An emerging enigmatic and underestimated pathogen. *Saudi J Biol Sci* 2022;29(1):499-512.
- Kamar N, Bendall R, Legrand-Abravanel F, et al. Hepatitis E. *Lancet* (London, England) 2012;379(9835):2477-2488.
- Aggarwal R, Goel A. Natural History, Clinical Manifestations, and Pathogenesis of Hepatitis E Virus Genotype 1 and 2 Infections. *Cold Spring Harb Perspect Med* 2019;9(7).
- Smith DB, Izopet J, Nicot F, et al. Update: proposed reference sequences for subtypes of hepatitis E virus (species Orthohepevirus A). *J Gen Virol* 2020;101(7):692-698.
- Purdy MA, Drexler JF, Meng XJ, et al. ICTV Virus Taxonomy Profile: Hepeviridae 2022. *J Gen Virol* 2022;103(9).
- Ruggeri FM, Di Bartolo I, Ponterio E, et al. Zoonotic transmission of hepatitis E virus in industrialized countries. *New Microbiol* 2013;36(4):331-44.
- Kamar N, Izopet J, Pavio N, et al. Hepatitis E virus infection. *Nat Rev Dis Primers* 2017;3:17086.
- World Health Organisation (WHO) Factsheet. Hepatitis E. Last updated June 2022. Available at: <https://www.who.int/news-room/factsheets/detail/hepatitis-e>. Accessed 31 October 2022.
- Jilani N, Das BC, Husain SA, et al. Hepatitis E virus infection and fulminant hepatic failure during pregnancy. *J Gastroenterol Hepatol* 2007;22(5):676-682.
- Khuroo MS, Kamili S, Khuroo MS. Clinical course and duration of viremia in vertically transmitted hepatitis E virus (HEV) infection in babies born to HEV-infected mothers. *J Viral Hepat* 2009;16(7):519-523.
- Li M, Bu Q, Gong W, et al. Hepatitis E virus infection and its associated adverse fetal-maternal outcomes among pregnant women in Qinhuangdao, China. *J Matern Fetal Neonatal Med* 2020;33(21):3647-3651.
- Wu C, Wu X, Xia J. Hepatitis E virus infection during pregnancy. *Virol J* 2020;17(1):73.
- Lhomme S, Marion O, Abravanel F, et al. Hepatitis E Pathogenesis. *Viruses* 2016;8(8).
- European Association of the Study of the Liver (EASL). Clinical Practice Guidelines on hepatitis E virus infection. *J Hepatol* 2018;68(6):1256-1271.
- Larrue H, Abravanel F, Péron JM. Hepatitis E, what's the real issue? *Liver Int* 2020;40 Suppl 1:43-47.
- Lhomme S, Dubois M, Abravanel F, et al. Risk of zoonotic transmission of HEV from rabbits. *J Clin Virol* 2013;58(2):357-362.
- Zhao Q, Zhang J, Wu T, et al. Antigenic determinants of hepatitis E virus and vaccine-induced immunogenicity and efficacy. *J Gastroenterol* 2013;48(2):159-168.
- Aggarwal R. Hepatitis E: clinical presentation in disease-endemic areas and diagnosis. *Semin Liver Dis* 2013;33(1):30-40.
- Sultana R, Islam MT, Bhuyan GS, et al. IgG antibody response demonstrates inverse correlation with viral load in Bangladeshi women with acute hepatitis E virus genotype 1 infection. *Int J Infect Dis* 2021;104:482-490.
- Velavan TP, Pallerla SR, John R, et al. Hepatitis E: An update on One Health and clinical medicine. *Liver Int* 2021;41(7):1462-1473.
- Kar P, Karna R. A Review of the Diagnosis and Management of Hepatitis E. *Curr Treat Options Infect Dis* 2020;12(3):310-320.
- Pawlotsky JM. Hepatitis E screening for blood donations: an urgent need? *Lancet* 2014;384(9956):1729-1730.

Elecsys Anti-HEV IgG



- 35 Zhou Y. Immunobiology and Host Response to HEV. Adv Exp Med Biol 2016;948:113-141.
- 36 Bendall R, Ellis V, Ijaz S, et al. A comparison of two commercially available anti-HEV IgG kits and a re-evaluation of anti-HEV IgG seroprevalence data in developed countries. J Med Virol 2010;82(5):799-805.
- 37 European Centre for Disease Prevention and Control (ECDC). Options for national testing and surveillance for hepatitis E virus in the EU/EEA - operational guidance. 2019. Available at: <https://www.ecdc.europa.eu/en/publications-data/options-national-testing-and-surveillance-hepatitis-e-virus-eueea-operational>. Accessed 28 October 2022.
- 38 Centers for Disease Control and Prevention (CDC). Hepatitis E Questions and Answers for Health Professionals. 2020. Available at: <https://www.cdc.gov/hepatitis/hev/hevfaq.htm>. Accessed 26 January 2023.
- 39 Seth A, Sherman KE. Hepatitis E: What We Think We Know. Clin Liver Dis 2020;15(Suppl 1):S37-S44.
- 40 Occupational Safety and Health Standards: Bloodborne pathogens. (29 CFR Part 1910.1030). Fed. Register.
- 41 Directive 2000/54/EC of the European Parliament and Council of 18 September 2000 on the protection of workers from risks related to exposure to biological agents at work.

For further information, please refer to the appropriate operator's manual for the analyzer concerned, the respective application sheets and the Method Sheets of all necessary components (if available in your country).

A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Any serious incident that has occurred in relation to the device shall be reported to the manufacturer and the competent authority of the Member State in which the user and/or the patient is established.

The Summary of Safety & Performance Report can be found here: <https://ec.europa.eu/tools/eudamed>

Symbols

Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard (for USA: see [dialog.roche.com](https://www.dialog.roche.com) for definition of symbols used):

	Contents of kit
	Analyzers/Instruments on which reagents can be used
	Reagent
	Calibrator
	Volume for reconstitution
	Global Trade Item Number

COBAS, COBAS E, ELECSYS and PRECICONTROL are trademarks of Roche. INTRALIPID is a trademark of Fresenius Kabi AB.

All other product names and trademarks are the property of their respective owners.

Additions, deletions or changes are indicated by a change bar in the margin.

© 2023, Roche Diagnostics



Roche Diagnostics GmbH, Sandhofer Strasse 116, D-68305 Mannheim
www.roche.com

+800 5505 6606

