Rx Only

cobas® CT/NG

Qualitative nucleic acid test
for use on the cobas® 5800/6800/8800 Systems

For in vitro diagnostic use

cobas® CT/NG	P/N: 09040501190
For use on the cobas® 5800 System	
cobas® CT/NG Positive Control Kit	P/N: 09040510190
cobas® Buffer Negative Control Kit	P/N: 09051953190
For use on the cobas® 6800/8800 Systems	
cobas® CT/NG Positive Control Kit	P/N: 07460082190 or P/N: 09040510190
cobas® Buffer Negative Control Kit	P/N: 07002238190 or P/N: 09051953190

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Intended use

cobas® CT/NG for use on the **cobas**® 5800/6800/8800 Systems is an automated, qualitative in vitro diagnostic test, that utilizes real-time polymerase chain reaction (PCR), for the direct detection of *Chlamydia trachomatis* (CT) and/or *Neisseria gonorrhoeae* (NG) DNA in male and female urine, clinician-instructed self-collected vaginal swab specimens, clinician-collected vaginal swab specimens, endocervical swab specimens, oropharyngeal (throat) swab specimens, and anorectal swab specimens, all collected in **cobas**® PCR Media (Roche Molecular Systems, Inc.), and cervical specimens collected in PreservCyt® Solution. This test is intended as an aid in the diagnosis of chlamydial and gonococcal disease in both symptomatic and asymptomatic individuals.

Summary and explanation of the test

Background

Infection with CT is the leading bacterial cause of sexually transmitted diseases worldwide, with approximately 89.1 million cases occurring annually.[1] *C. trachomatis* is the most frequently reported bacterial sexually transmitted disease (STD) in the United States[1,2] and prevalence is highest in persons aged ≤ 24 years.[3] In 2013, a total of 1,401,906 cases of *C. trachomatis* infection were reported to the CDC corresponding to a rate of 446.6 cases per 100,000 population.[3]

CT is a gram-negative, nonmotile, obligate intracellular bacterium with a unique biphasic lifecycle.[1] CT causes a variety of infections including urethritis, cervicitis, proctitis, conjunctivitis, endometritis, and salpingitis; if left untreated, the infection may ascend to the uterus, fallopian tubes, and ovaries causing pelvic inflammatory syndrome, ectopic pregnancy, and tubal factor infertility. Reiter's syndrome (urethritis, conjunctivitis, arthritis, and mucocutaneous lesions) has also been associated with genital CT infection.[1] Many infections remain asymptomatic, and high numbers of infected patients may not seek care.[4] Patients often become re-infected if their sexual partners are not treated. Infants born to infected mothers can develop conjunctivitis, pharyngitis, and pneumonia. The predominant symptoms in men and women are increased discharge and dysuria; women may also present with irregular uterine bleeding.[1]

The diagnosis of *C. trachomatis* urogenital infection in women is made by testing first-catch urine or collecting swab specimens from the endocervix or vagina. Diagnosis of *C. trachomatis* urethral infection in men can be made by testing a urethral swab or first-catch urine specimen. Nucleic acid amplification tests (NAATs) are the most sensitive tests for these specimens and therefore are recommended for detecting *C. trachomatis* infection.[5] Anorectal and oropharyngeal *C. trachomatis* infection in persons engaging in receptive anal or oral intercourse can be diagnosed by testing at the anatomic site of exposure.

Annual screening for CT of all sexually active women aged < 25 years is recommended and screening of older women is recommended in the presence of increased risk for infection (e.g., those who have a new sex partner, more than one sex partner, a sex partner with concurrent partners, or a sex partner who has a sexually transmitted infection).[6] Chlamydia screening programs have been demonstrated to reduce the rates of PID in women.[7,8] Although the evidence to support routine screening for CT in sexually active young men is insufficient, due to the relative lack of feasibility, efficacy, and cost-effectiveness studies, the screening of sexually active young men should be considered in clinical settings with a high prevalence of chlamydia (e.g., adolescent clinics, correctional facilities, and STD clinics) or in populations with high burden of infection (e.g., MSM).[2,6] The primary focus of chlamydia screening efforts among women should be to detect chlamydia, prevent complications, and test and treat their partners, whereas targeted chlamydia screening in men should only be considered when resources permit, prevalence is high, and such screening does not hinder chlamydia screening efforts in women.[9,10] More frequent screening for some women (e.g., adolescents) or certain men (e.g., MSM) might be indicated.[2]

NG is the etiologic agent of gonorrhea. NG are cytochrome oxidase-positive, non-motile, non-spore forming gram-negative diplococci. In the United States, an estimated 820,000 new *N. gonorrhoeae* infections occur each year.[11] Gonorrhea is the second most commonly reported communicable disease.[3] Clinical manifestations of NG infections are numerous.[4] In men, acute urethritis presents itself after a 1-10 day incubation period with urethral discharge and dysuria. Only a small proportion of men remain asymptomatic without signs of urethritis.[12] Acute epididymitis is the most common complication, especially in young men. In women, the primary site of infection is the endocervix. There is a high prevalence of coalescence of symptoms with CT, *Trichomonas vaginalis*, and vaginosis; many women remain asymptomatic and therefore do not seek medical care. In symptomatic women increased discharge, dysuria, and intermenstrual bleeding may be observed.[13] Pelvic inflammatory disease (PID) can occur in 10%-20% of women, combined with endometritis, salpingitis, tubo-ovarian abscess, pelvic peritonitis, and perihepatitis.[14] PID can result in tubal scarring that can lead to infertility and ectopic pregnancy. Other gonococcal infected sites in men and women are the rectum, pharynx, conjunctiva, and to a lesser degree the disease presents itself as disseminated gonococcal infection. Infants from infected mothers can develop conjunctivitis.

Annual screening for *N. gonorrhoeae* infection is recommended for all sexually active women aged < 25 years and for older women at increased risk for infection (e.g., those who have a new sex partner, more than one sex partner, a sex partner with concurrent partners, or a sex partner who has an STI).[6] Additional risk factors include inconsistent condom use among persons with multiple sex partners, previous or coexisting sexually transmitted infections, and exchanging sex for money or drugs.[2] In addition to urethral infections, the CDC also recommends the use of NAATs for routine annual screening for men who have sex with men (MSM) for anorectal or oral infection.[5]

Rationale for CT/NG testing

NAATs are the recommended method for CT and NG screening.[15] For women, a vaginal swab is the recommended sample type and first catch urine is recommended for men. Alternative acceptable sample types for women include an endocervical swab when a pelvic examination is indicated or a first catch urine sample, but a urine sample may detect up to 10% fewer infections when compared with vaginal and endocervical swabs. In addition to urine for men, a urethral swab is also acceptable. In addition, the CDC recommends at least annual screening for CT from urethral or anorectal specimens and for NG from urethral, anorectal or oral specimens in MSM.[2]

cobas® CT/NG for use on the **cobas®** 5800 System and **cobas®** 6800/8800 Systems (referred to as **cobas®** CT/NG throughout the remainder of this document) is an automated, qualitative real-time PCR test designed to detect CT and NG DNA in urogenital, oropharyngeal and anorectal specimens from male and female patients and thus fulfills the medical need for a rapid, high throughput molecular screening test for use as an aid in the diagnosis of chlamydial and gonococcal disease in both symptomatic and asymptomatic individuals.

Explanation of the test

cobas® CT/NG is a qualitative test performed on the **cobas®** 5800 System, **cobas®** 6800 System or **cobas®** 8800 System. **cobas®** CT/NG enables the detection of CT/NG DNA in endocervical, vaginal, oropharyngeal, anorectal, urine and cervical specimens of infected female patients and oropharyngeal, anorectal and urine specimens in male patients. Target-specific primers and two probes are used to detect but not discriminate between the CT cryptic plasmid and the ompA gene. Additionally, target-specific primers and two probes are used to detect but not discriminate between two conserved sequences in the NG DR-9 region. The DNA Internal Control, used to monitor the entire sample preparation and PCR amplification process, is introduced into each specimen during sample processing. In addition, the test utilizes a low titer positive and a negative control.

Principles of the procedure

cobas® CT/NG is based on fully automated sample preparation (nucleic acid extraction and purification) followed by PCR amplification and detection. The **cobas**® 5800 System is designed as one integrated instrument. The **cobas**® 6800/8800 Systems consist of the sample supply module, the transfer module, the processing module, and the analytic module. Automated data management is performed by the **cobas**® 5800 System or **cobas**® 6800/8800 System software which assigns test results for all tests as positive, negative or invalid. Results can be reviewed directly on the system screen, exported, or printed as a report.

Nucleic acid from patient samples, external controls and added internal control DNA (DNA-IC) molecules is simultaneously extracted. In summary, bacterial nucleic acid is released by addition of proteinase and lysis reagent to the sample. The released nucleic acid binds to the silica surface of the added magnetic glass particles. Unbound substances and impurities, such as denatured protein, cellular debris and potential PCR inhibitors are removed with subsequent wash steps and purified nucleic acid is eluted from the magnetic glass particles with elution buffer at elevated temperature.

Selective amplification of target nucleic acid from the sample is achieved by the use of target-specific forward and reverse primers which are selected from highly conserved plasmid and genomic regions of CT and NG. A region on the CT cryptic plasmid and the ompA gene (dual target) and two conserved sequences of the NG DR-9 region are amplified by **cobas**® CT/NG. Selective amplification of DNA IC is achieved by the use of sequence-specific forward and reverse primers which are selected to have no homology with either the CT or NG target regions. A thermostable DNA polymerase enzyme is used for PCR amplification. The target and DNA-IC sequences are amplified simultaneously utilizing a universal PCR amplification profile with predefined temperature steps and number of cycles. The master mix includes deoxyuridine triphosphate (dUTP), instead of deoxythimidine triphosphate (dTTP), which is incorporated into the newly synthesized DNA (amplicon). Any contaminating amplicon from previous PCR runs are eliminated by the AmpErase enzyme, which is included in the PCR master mix, during the first thermal cycling step.[16] However, newly formed amplicon are not eliminated since the AmpErase enzyme is inactivated once exposed to temperatures above 55°C.

The **cobas**® CT/NG master mix contains two detection probes specific for the CT target sequences, two detection probes specific for the NG target sequences and one for the DNA-IC. The probes are labeled with target specific fluorescent reporter dyes allowing simultaneous detection of CT targets, NG targets and DNA-IC in three different target channels.[17,18] When not bound to the target sequence, the fluorescent signal of the intact probes is suppressed by a quencher dye. During the PCR amplification step, hybridization of the probes to the specific single-stranded DNA template results in cleavage of the probe by the 5' to 3' exonuclease activity of the DNA polymerase resulting in separation of the reporter and quencher dyes and the generation of a fluorescent signal. With each PCR cycle, increasing amounts of cleaved probes are generated and the cumulative signal of the reporter dye increases concomitantly. Real-time detection and discrimination of PCR products is accomplished by measuring the fluorescence of the released reporter dyes for the CT and NG targets and DNA-IC, respectively.

Reagents and materials

cobas® CT/NG reagents and controls

All unopened reagents and controls shall be stored as recommended in Table 1 to Table 4.

Table 1 cobas® CT/NG

(cobas® CT/NG)

Store at 2-8°C

480 test cassette (P/N 09040501190)

Kit components	Reagent ingredients	Quantity per kit 480 tests
Proteinase Solution (PASE)	Tris buffer, < 0.05% EDTA, Calcium chloride, Calcium acetate, 8% Proteinase EUH210: Safety data sheet available on request. EUH208: Contains Subtilisin from Bacillus subtilis. May produce an allergic reaction.	38 mL
DNA Internal Control (DNA-IC)	Tris buffer, < 0.05% EDTA, < 0.001% non-CT/NG related DNA construct containing primer and probe specific sequence regions, < 0.1% Sodium azide	38 mL
Elution Buffer (EB)	Tris buffer, 0.2% Methyl-4 hydroxibenzoate	38 mL
Master Mix Reagent 1 (MMX-R1)	Manganese acetate, Potassium hydroxide, < 0.1% Sodium azide	14.5 mL
CT/NG Master Mix Reagent 2 (CT/NG MMX-R2)	Tricine buffer, Potassium acetate, EDTA, Glycerol, < 18% Dimethyl sulfoxide, < 0.12% dATP, dCTP, dGTP, dUTPs, < 0.1% Tween 20, < 0.1% Sodium azide, < 0.1% Z05 DNA polymerase, < 0.10% AmpErase (uracil-N glycosylase) enzyme (microbial), < 0.01% Internal Control forward and reverse primers, < 0.01% Upstream and downstream CT/NG primers, < 0.01% Fluorescent-labeled oligonucleotide probes specific for CT, NG and the DNA Internal Control, < 0.01% Oligonucleotide aptamer	17.5 mL

Table 2 cobas® CT/NG Positive Control Kit

(cobas® CT/NG Positive Control Kit)

Store at 2–8°C

For use on the **cobas®** 5800 System (P/N 09040510190)

For use on the **cobas®** 6800/8800 Systems (P/N 07460082190 or P/N 09040510190)

Kit components	Reagent ingredients	Quantity per kit
CT/NG Positive Control (CT/NG (+) C)	Tris buffer, < 0.05% Sodium azide, < 0.005% EDTA, < 0.003% Poly rA, <0.01% Non-infectious plasmid DNA (microbial) containing *C. trachomatis*, <0.01% Non-infectious plasmid DNA (microbial) containing *N. gonorrhoeae*	16 mL (16 x 1mL)

Table 3 cobas® Buffer Negative Control Kit

(cobas® Buffer Negative Control Kit)

Store at 2-8°C

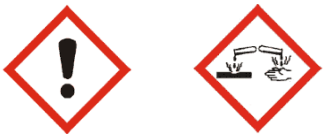
For use on the **cobas®** 5800 System (P/N 09051953190)

For use on the **cobas®** 6800/8800 Systems (P/N 07002238190 or P/N 09051953190)

Kit components	Reagent ingredients	Quantity per kit
cobas® Buffer Negative Control (BUF (-) C)	Tris buffer, < 0.1% sodium azide, EDTA, < 0.002% Poly rA RNA (synthetic)	16 mL (16 x 1mL)

cobas® omni reagents for sample preparation

Table 4 cobas® omni reagents for sample preparation*

Reagents	Reagent ingredients	Quantity per kit	Safety symbol and warning**
cobas® omni MGP Reagent (MGP) Store at 2–8°C (P/N 06997546190)	Magnetic glass particles, Tris buffer, 0.1% methyl-4 hydroxybenzoate, < 0.1% sodium azide	480 tests	Not applicable
cobas® omni Specimen Diluent (SPEC DIL) Store at 2–8°C (P/N 06997511190)	Tris buffer, 0.1% methyl-4 hydroxybenzoate, < 0.1% sodium azide	4 x 875 mL	Not applicable
cobas® omni Lysis Reagent (LYS) Store at 2–8°C (P/N 06997538190)	42.56% (w/w) guanidine thiocyanate***, 5% (w/v) polydocanol***, 2% (w/v) dithiothreitol***, dihydro sodium citrate	4 x 875 mL	 **DANGER** H302: Harmful if swallowed. H314: Causes severe skin burns and eye damage. H412: Harmful to aquatic life with long lasting effects. EUH032: Contact with acids liberates very toxic gas. EUH071: Corrosive to the respiratory tract. P273: Avoid release to the environment. P280: Wear protective gloves/protective clothing/eye protection/face protection/ hearing protection. P301 + P330 + P331: IF SWALLOWED: Rinse mouth. Do NOT induce vomiting. P303 + P361 + P353: IF ON SKIN (or hair): Take off immediately all contaminated clothing. Rinse skin with water. P304 + P340 + P310: IF INHALED: Remove person to fresh air and keep comfortable for breathing. Immediately call a POISON CENTER/doctor. P305 + P351 + P338 + P310: IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing. Immediately call a POISON CENTER/doctor. 593-84-0 Guanidinium thiocyanate 9002-92-0 Polidocanol 3483-12-3 (R*,R*)-1,4-dimercaptobutane-2,3-diol
cobas® omni Wash Reagent (WASH) Store at 15–30°C (P/N 06997503190)	Sodium citrate dihydrate, 0.1% methyl-4 hydroxybenzoate	4.2L	Not applicable

* These reagents are not included in the **cobas**® CT/NG kit. See listing of additional materials required for **cobas**® 5800 and **cobas**® 6800/8800 (Table 8 and Table 9).

** Product safety labeling primarily follows EU GHS guidance

***Hazardous substance

Reagent storage and handling requirements

Reagents shall be stored and will be handled as specified in Table 5, Table 6, and Table 7.

When reagents are not loaded on the **cobas**® 5800 System or **cobas**® 6800/8800 Systems, store them at the corresponding temperature specified in Table 5.

Table 5 Reagent storage (when reagent is not on the system)

Reagent	Storage temperature
cobas® CT/NG	2–8°C
cobas® CT/NG Positive Control Kit	2–8°C
cobas® Buffer Negative Control Kit	2–8°C
cobas® **omni** Lysis Reagent	2–8°C
cobas® **omni** MGP Reagent	2–8°C
cobas® **omni** Specimen Diluent	2–8°C
cobas® **omni** Wash Reagent	15–30°C

Reagent handling requirements for cobas® 5800 System

Reagents loaded onto the **cobas**® 5800 System are stored at appropriate temperatures and their expiration is monitored by the system. The **s**ystem allows reagents to be used only if all of the conditions shown in Table 6 are met. The system automatically prevents use of expired reagents. Table 6 allows the user to understand the reagent handling conditions enforced by the **cobas**® 5800 System.

Table 6 Reagent expiry conditions enforced by the **cobas**® 5800 System

Reagent	Kit expiration date	Open-kit stability	Number of runs for which this kit can be used	On-board stability
cobas® CT/NG	Date not passed	90 days from first usage	Max 40 runs	Max 36 days*
cobas® CT/NG Positive Control Kit	Date not passed	Not applicable**	Not applicable	Max 36 days*
cobas® Buffer Negative Control Kit	Date not passed	Not applicable**	Not applicable	Max 36 days*
cobas® **omni** Lysis Reagent	Date not passed	30 days from loading*	Not applicable	Not applicable
cobas® **omni** MGP Reagent	Date not passed	30 days from loading*	Not applicable	Not applicable
cobas® **omni** Specimen Diluent	Date not passed	30 days from loading*	Not applicable	Not applicable
cobas® **omni** Wash Reagent	Date not passed	30 days from loading*	Not applicable	Not applicable

* Time is measured from the first time that reagent is loaded onto the **cobas**® 5800 System.

** Single use reagents

Reagent handling requirements for cobas® 6800/8800 Systems

Reagents loaded onto the **cobas**® 6800/8800 Systems are stored at appropriate temperatures and their expiration is monitored by the system. The **cobas**® 6800/8800 Systems allow reagents to be used only if all of the conditions shown in Table 7 are met. The system automatically prevents use of expired reagents. Table 7 allows the user to understand the reagent handling conditions enforced by the **cobas**® 6800/8800 Systems.

Table 7 Reagent expiry conditions enforced by the **cobas**® 6800/8800 Systems

Reagent	Kit expiration date	Open-kit stability*	Number of runs for which this kit can be used	On-board stability (cumulative time on board outside refrigerator)
cobas® CT/NG	Date not passed	90 days from first usage	Max 20 runs	Max 20 hours
cobas® CT/NG Positive Control Kit	Date not passed	Not applicable**	Not applicable	Max 10 hours
cobas® Buffer Negative Control Kit	Date not passed	Not applicable**	Not applicable	Max 10 hours
cobas® **omni** Lysis Reagent	Date not passed	30 days from loading*	Not applicable	Not applicable
cobas® **omni** MGP Reagent	Date not passed	30 days from loading*	Not applicable	Not applicable
cobas® **omni** Specimen Diluent	Date not passed	30 days from loading*	Not applicable	Not applicable
cobas® **omni** Wash Reagent	Date not passed	30 days from loading*	Not applicable	Not applicable

* Time is measured from the first time that reagent is loaded onto the **cobas**® 6800/8800 Systems.

** Single use reagent

Additional materials required for cobas® 5800 System

Table 8 Material and consumables for use on **cobas®** 5800 System

Material	P/N
cobas® omni Processing Plate 24	08413975001
cobas® omni Liquid Waste Plate 24	08413983001
cobas® omni Amplification Plate 24	08499853001
Tip CORE TIPS with Filter, 1ml	04639642001
Tip CORE TIPS with Filter, 300uL	07345607001
cobas® omni Liquid Waste Container	07094388001
cobas® omni Lysis Reagent	06997538190
cobas® omni MGP Reagent	06997546190
cobas® omni Specimen Diluent	06997511190
cobas® omni Wash Reagent	06997503190
Solid Waste Bag or Solid Waste Bag With insert	07435967001 or 08030073001
16-position tube S-carrier complete	09224319001
5-position Rack Carrier	09224475001
Cell Collection Media Carrier	09224599001

Additional materials required for cobas® 6800/8800 Systems

Table 9 Materials and consumables for use on **cobas**® 6800/8800 Systems

Material	P/N
cobas® **omni** Processing Plate	05534917001
cobas® **omni** Amplification Plate	05534941001
cobas® **omni** Pipette Tips	05534925001
cobas® **omni** Liquid Waste Container	07094388001
cobas® **omni** Lysis Reagent	06997538190
cobas® **omni** MGP Reagent	06997546190
cobas® **omni** Specimen Diluent	06997511190
cobas® **omni** Wash Reagent	06997503190
Solid Waste Bag and Solid Waste Container or Solid Waste Bag With Insert and Kit Drawer	07435967001 and 07094361001 or 08030073001 and 08387281001
STD-Rack. re-run R001-R025 PINK	12025639001

Instrumentation and software required

The **cobas**® 5800 software and **cobas**® CT/NG analysis package (ASAP) for **cobas**® 5800 System shall be installed on the **cobas**® 5800 instrument. The Data Manager software and PC for **cobas**® 5800 System will be provided with the system.

The **cobas**® 6800/8800 System software and **cobas**® CT/NG analysis packages (ASAPs) for the **cobas**® 6800/8800 shall be installed on the instrument(s). The Instrument Gateway (IG) server will be provided with the system.

Table 10 Instrumentation

Equipment	P/N
cobas® 5800 System	08707464001
cobas® 6800 System (Option Moveable)	05524245001 and 06379672001
cobas® 6800 System (Fix)	05524245001 and 06379664001
cobas® 8800 System	05412722001
Sample Supply Module for **cobas**® 6800/8800 Systems	06301037001

Additional materials required for sample collection for cobas® CT/NG

Table 11 Specimen collection kits used with **cobas®** CT/NG

Collection Kit	P/N
cobas® PCR Media Kit	06466281190
cobas® PCR Urine Sample Kit	05170486190
cobas® PCR Media Uni Swab Sample Kit	07958030190
cobas® PCR Media Dual Swab Sample Kit	07958021190
ThinPrep Pap Test Physician's Kit (500 vials & Broom-like collection devices) ThinPrep Pap Test Physician's Kit (500 vials & Cytobrush/spatula collection devices)	Hologic: 70136-001 Hologic: 70136-002

cobas® CT/NG accepts the primary tube used for all **cobas®** PCR media swab and urine specimen types. Refer to the **cobas®** 5800 System or the **cobas®** 6800/8800 Systems User Assistance and/or User Guides for additional information for primary and secondary sample tubes accepted on the instruments.

Note: Contact your local Roche representative for a detailed order list for sample racks, racks for clotted tips and rack trays accepted on the instruments

Additional materials required for sample aliquoting and sample loading for cobas® CT/NG

Table 12 Specimen collection kits used with **cobas®** CT/NG

Material	P/N
cobas® PCR Media Secondary Tube Kit	07958048190
cobas® PCR Media Tube Replacement Cap Kit	07958056190
Replacement Caps for PreservCyt® Vials	08037230190
cobas® PCR Media Disposable Tube Stand (Optional)	07958064190
MPA RACK 16 MM LIGHT GREEN 7001-7050[a,b,c]	03143449001
RD5 RACK – RD Standard rack 0001-0050 LR[a,b,c]	11902997001

[a] RD5 or MPA racks are required in combination with the 5-position Rack Carrier on the **cobas®** 5800 System.

[b] MPA 16mm rack or 16-position tube carrier are the preferred racks for use with samples collected in **cobas®** PCR Media tubes.

[c] MPA or RD5 racks identified are example materials and part numbers. Please contact your local Roche representative for a detailed order list for sample racks and rack carriers accepted on the instruments.

Precautions and handling requirements

Warnings and precautions

As with any test procedure, good laboratory practice is essential to the proper performance of this assay. Due to the high sensitivity of this test, care should be taken to keep reagents and amplification mixtures free of contamination.

- For in vitro diagnostic use only.
- All patient samples should be handled as if infectious, using good laboratory procedures as outlined in Biosafety in Microbiological and Biomedical Laboratories and in the CLSI Document M29-A4.[19,20] Only personnel proficient in handling infectious materials and the use of **cobas**® CT/NG and **cobas**® 5800 System or **cobas**® 6800/8800 Systems should perform this procedure.
- All human-sourced materials should be considered potentially infectious and should be handled with universal precautions. If spillage occurs, immediately disinfect with a freshly prepared solution of 0.6% sodium or potassium hypochlorite in distilled or deionized water or follow appropriate site procedures.
- Do not freeze any samples.
- Use only supplied or specified required consumables to ensure established test performance.
- Safety Data Sheets (SDS) are available on request from your local Roche representative.
- Closely follow procedures and guidelines provided to ensure that the test is performed correctly. Any deviation from the procedures and guidelines may affect established test performance.
- False positive results may occur if carryover of samples is not adequately controlled during sample handling and processing.
- **cobas**® PCR Media (from primary specimen tube) contains guanidine hydrochloride. **Do not allow direct contact between guanidine hydrochloride and sodium hypochlorite (bleach) or other highly reactive reagents such as acids or bases. These mixtures can release a noxious gas**. If liquid containing guanidine hydrochloride is spilled, clean with suitable laboratory detergent and water. If the spilled liquid contains potentially infectious agents, **FIRST** clean the affected area with laboratory detergent and water, and then with 0.6% sodium or potassium hypochlorite.
- Inform your local competent authority about any serious incidents which may occur when using this assay.

Reagent handling

- Handle all reagents, controls, and samples according to good laboratory practice in order to prevent carryover of samples, reagents, or controls.
- Before use, visually inspect each reagent cassette, diluent, lysis reagent, and wash reagent to ensure that there are no signs of leakage. If there is any evidence of leakage, do not use that material for testing.
- **cobas**® **omni** Lysis Reagent contains guanidine thiocyanate, a potentially hazardous chemical. Avoid contact of reagents with the skin, eyes, or mucous membranes. If contact does occur, immediately wash with generous amounts of water; otherwise, burns can occur.
- Expended control kits contain pierced vials with residual reagent; special care should be taken during disposal to avoid spills and contact.
- **cobas**® CT/NG kit, **cobas**® CT/NG Positive Control kit, **cobas**® Buffer Negative Control kit, **cobas**® **omni** MGP Reagent, and **cobas**® **omni** Specimen Diluent contain sodium azide as a preservative. Avoid contact of reagents with the skin, eyes, or mucous membranes. If contact does occur, immediately wash with generous amounts of

water; otherwise, burns can occur. If these reagents are spilled, dilute with water before wiping dry.

- Do not allow **cobas**® **omni** Lysis Reagent, which contains guanidine thiocyanate, to contact sodium hypochlorite (bleach) solution. This mixture can produce a highly toxic gas.
- Dispose of all materials that have come in contact with samples and reagents in accordance with country, state, and local regulations.

Good laboratory practice

- Do not pipette by mouth.
- Do not eat, drink, or smoke in designated work areas.
- Wear laboratory gloves, laboratory coats, and eye protection when handling samples and reagents. Avoid contaminating gloves when handling samples and controls. Gloves must be changed between handling samples and **cobas**® CT/NG kit, **cobas**® CT/NG Positive Control kit, **cobas**® Buffer Negative Control kit**,** and **cobas**® **omni** reagents to prevent contamination.
- Wash hands thoroughly after handling samples and reagents, and after removing the gloves.
- Thoroughly clean and disinfect all laboratory work surfaces with a freshly prepared solution of 0.6% sodium or potassium hypochlorite in distilled or deionized water. Follow by wiping the surface with 70% ethanol.
- If spills occur on the **cobas**® 5800 instrument or **cobas**® 6800/8800 instruments, follow the instructions in the **cobas**® 5800 Systems or **cobas**® 6800/8800 Systems User Assistance and/or User Guides to properly clean and decontaminate the surface of instrument(s).

Specimen collection, transport, and storage

Note: Handle all samples and controls as if they are capable of transmitting infectious agents.

Specimen collection

Endocervical swab specimens collected with the **cobas**® PCR Media Dual Swab Sample Kit, vaginal swab specimens, anorectal swab specimens and oropharyngeal swab specimens collected with either the **cobas**® PCR Media Uni Swab Sample Kit or **cobas**® PCR Media Dual Swab Sample Kit, male and female urine collected with the **cobas**® PCR Urine Sample Kit and cervical specimens collected in PreservCyt® Solution have been validated for use with **cobas**® CT/NG (see Table 11 for a list of all collection kits). Follow the instructions for collecting all swab and urine specimens in their respective collection kit IFU. Follow the manufacturer's instructions for collecting cervical specimens into PreservCyt® Solution.

Specimen transport

All specimen types listed in Specimen Collection section above can be transported at 2-30°C. Transportation of CT/NG specimens in **cobas**® PCR Media and PreservCyt® Solution must comply with country, federal, state and local regulations for the transport of etiologic agents.[21]

Specimen storage

Table 13 Summary of acceptable specimen storage conditions prior to testing with **cobas**® CT/NG

Specimen Type		**2-30°C**
Samples in **cobas**® PCR Media		12 months
PreservCyt®	in collection device *or* PreservCyt® samples aliquoted to secondary tubes	12 months 31 days

Note: PreservCyt® and **cobas**® PCR Media specimens should not be frozen.

Male and female urine specimens

- Use only the **cobas**® PCR Urine Sample Kit or the **cobas**® PCR Media Kit to collect urine specimens for **cobas**® CT/NG. **cobas**® CT/NG has not been validated for use with other urine collection devices or media types. Using **cobas**® CT/NG with other urine collection devices or other media types may lead to false negative, false positive, and/or invalid results.
- To avoid cross contamination of processed specimens, additional caps for **cobas**® PCR Media tubes in an alternate color (neutral; see **Additional materials required for sample aliquoting and sample loading for cobas® CT/NG**) should be used to recap specimens after processing.
- Untested urine specimens must show the top of the liquid level between the two black lines on the **cobas**® PCR Media tube label window. If the liquid level is above or below these lines, the specimen has not been collected properly and cannot be used for testing.
- If additional testing is required, ensure that there is at least 1.2 mL of specimen remaining the in **cobas**® PCR Media tube.

Endocervical, vaginal, anorectal and oropharyngeal specimens

- The presence of mucus in endocervical and cervical specimens may cause processing delays due to clotting. Mucus free specimens are required for optimal test performance. Use the large woven polyester swab in the **cobas**® PCR Media Dual Swab Sample Kit or an equivalent device to remove cervical secretions and discharge before obtaining the endocervical or cervical specimen.
- Use only the flocked swab in the **cobas**® PCR Media Dual Swab Sample Kit to collect endocervical specimens. Use only the woven polyester swab in either the **cobas**® PCR Media Uni Swab Sample Kit or the **cobas**® PCR Media Dual Swab Sample Kit to collect vaginal, anorectal, and oropharyngeal swab specimens. **cobas**® CT/NG has not been validated for use with other swab collection devices or media types. Using **cobas**® CT/NG with other swab collection devices or media types may lead to false negative, false positive, and/or invalid results.
- To avoid cross contamination of processed specimens, additional caps for **cobas**® PCR Media tubes in an alternate color (neutral; see **Additional materials required for sample aliquoting and sample loading for cobas® CT/NG**) should be used to recap specimens after processing.
- All swab specimens containing a single swab in the **cobas**® PCR Media tube can be directly processed on the **cobas**® 5800 System or **cobas**® 6800/8800 Systems. If desired, the swab may be removed before the specimen tube is loaded onto the instrument, however utmost care must be exercised to avoid cross contamination.
- A properly collected swab specimen should have a single swab with the shaft broken at the score line. Swab shafts which are broken above the score line will appear longer than normal and may also be bent over to fit into the **cobas**® PCR Media tube. This can create an obstruction to the pipetting system which may cause the loss of sample, test results and/or mechanical damage to the instrument. In the event that a swab specimen has an improperly broken shaft, remove the swab prior to sample processing on the **cobas**® 5800 System or **cobas**® 6800/8800 Systems. Use caution when disposing of specimen swabs; avoid splashing or touching swabs to other surfaces during disposal to prevent contamination.
- Incoming primary swab specimen tubes with no swabs or with two swabs have not been collected according to the instructions in their respective collective kit IFU and should not be tested.
- Occasionally, incoming swab specimens contain excessive mucus which may induce a pipetting error (e.g., clot or other obstruction) on the **cobas**® 5800 System or **cobas**® 6800/8800 Systems. Prior to retesting of specimens that exhibited clots during initial processing, remove and discard the swab, then re-cap and vortex these specimens for 30 seconds to disperse the excess mucus.
- Swab specimens can be assayed twice on the **cobas**® 5800 System or **cobas**® 6800/8800 Systems while the swab is in the collection tube. If additional testing is required, or if the first test fails due to specimen pipetting error (e.g., clot or other obstruction), the swab must be removed prior to testing and the remaining fluid must have a minimum volume of 1.0 mL.

Cervical specimens in PreservCyt® Solution

cobas® CT/NG is validated for use with cervical specimens collected in PreservCyt® Solution. **cobas**® CT/NG has not been validated for use with cervical specimens obtained in other media types. Using **cobas**® CT/NG with other media types may lead to false negative, false positive, and/or invalid results.

cobas® 5800 System

- The **cobas**® 5800 System may process cervical specimens in PreservCyt® Solution directly out of their primary containers with a proper barcode or out of a properly barcoded cobas® PCR Media Secondary Tube (see **cobas**® 5800/6800/8800 System section below for optional aliquoting instructions for **cobas**® 5800 System).
    1. With clean gloved hands, vortex the capped primary vial for **10 seconds** immediately **prior** to loading.
    2. Uncap the primary vial and place on a Cell Collection Media Carrier.
- For primary vial loading, the minimum volume required in the PreservCyt® Solution primary containers is 3.0 mL.

cobas® 5800/6800/8800 Systems

- Cervical specimens in PreservCyt® Solution should be aliquoted into **cobas**® PCR Media Secondary Tubes as follows, for processing on the **cobas**® 5800 System or **cobas**® 6800/8800 Systems:
    1. Prepare a barcoded **cobas**® PCR Media Secondary Tube for each PreservCyt® specimen to be tested.
    2. With clean gloved hands, vortex each PreservCyt® primary specimen vial for 10 seconds immediately prior to transfer.
    3. Uncap a primary vial and transfer at least 1.0 mL but no more than 4.0 mL into the prepared barcoded secondary tube from step 1.
        - *Always use caution when transferring specimens from primary containers to secondary tubes.*
        - *Always use a new pipette tip for each specimen.*
        - *Always use pipettors with aerosol-barrier or positive-displacement tips to handle specimens.*
        - *To avoid cross contamination, additional caps for these tubes in an alternate color (neutral; see **Additional materials required for sample aliquoting and sample loading for cobas® CT/NG**) should be used to recap these specimens after processing.*
        - *Transfer tube to a rack if testing is to be performed shortly after or cap the secondary tube if testing will be performed at a future time.*
    4. Re-cap the primary vial with a replacement cap before moving to the next specimen. Store the primary vial upright.
    5. Only racks of uncapped tubes must be loaded on to the systems for CT/NG testing.
- Aliquots of the primary specimen must contain a minimum volume of 1.0 mL.

Instructions for use

Procedural notes

- Do not use **cobas**® CT/NG, **cobas**® CT/NG Positive Control Kit, **cobas**® Buffer Negative Control Kit, or **cobas**® **omni** reagents after their expiry dates.
- Do not reuse consumables. They are for one-time use only.

- Ensure that specimen barcode labels on sample tubes are visible through the openings on the side of sample carriers. Refer to the **cobas**® 5800 System or **cobas**® 6800/8800 Systems User Assistance and/or User Guide for proper barcode specifications and additional information on loading sample tubes.
- Refer to the **cobas**® 5800 System or **cobas**® 6800/8800 Systems User Assistance and/or User Guides for proper maintenance of instruments.

Running cobas® CT/NG on cobas® 5800 System

cobas® CT/NG can be run with a minimum required sample volume of 1.0 mL for swab and PreservCyt® specimens, 1.2 mL for urine specimens in **cobas**® PCR Media Tube, and 3.0 mL for PreservCyt® specimens in primary vial. The operation of the instrument is described in detail in the **cobas**® 5800 System User Assistance. Figure 1 below summarizes the procedure.

- Swab and Urine specimens should be uncapped and loaded directly onto racks for processing on the **cobas**® 5800 System.
- PreservCyt® specimens should be uncapped and run from primary vials.

Note: Use slow and steady movements when loading and unloading the Cell Collection Media Carrier (holding primary vials) to avoid splashing of specimens.

- Optionally, PreservCyt® specimens may be aliquoted into barcoded 13 mL round-bottom **cobas**® PCR Media Secondary tubes for processing on the **cobas**® 5800 System. Refer to the preparation instructions for cervical specimens found in section: "Cervical specimens in PreservCyt® Solution"
- A single run can have any combination of specimens (Swab, Urine, and PreservCyt®) and each specimen can be tested for either CT/NG, CT, or NG.
- Specimens collected in **cobas**® PCR Media or PreservCyt® Solution should be processed using the sample type selection in the user interface (UI) of the **cobas**® CT/NG as described in Table 14.

Table 14 Sample type selection in the user interface of the **cobas**® CT/NG

Specimen		**Collection kit type**	**Process as Sample Type**
Female	Vaginal swab	**cobas**® PCR Media Uni or Dual Swab Sample Kit	**Swab**
	Endocervical swab	**cobas**® PCR Media Dual Swab Sample Kit	**Swab**
	Oropharyngeal swab	**cobas**® PCR Media Uni or Dual Swab Sample Kit	**Swab**
	Anorectal swab	**cobas**® PCR Media Uni or Dual Swab Sample Kit	**Swab**
	Urine	**cobas**® PCR Urine Sample Kit or **cobas**® PCR Media Kit	**Urine**
	Cervical swab	PreservCyt® Solution (ThinPrep)	**PreservCyt®**
Male	Oropharyngeal swab	**cobas**® PCR Media Uni or Dual Swab Sample Kit	**Swab**
	Anorectal swab	**cobas**® PCR Media Uni or Dual Swab Sample Kit	**Swab**
	Urine	**cobas**® PCR Urine Sample Kit or **cobas**® PCR Media Kit	**Urine**

Figure 1 cobas® CT/NG procedure on **cobas®** 5800 System

1	Log onto the system
2	Loading samples onto the system ● For each urine or swab in **cobas®** PCR Media o Uncap tube o Transfer tube directly to rack ● For each primary PreservCyt® specimen vial: o If loading primary vial: ▪ Vortex for 10 seconds ▪ Uncap vial ▪ Transfer vial to rack o If loading secondary tube: ▪ Vortex primary vial for 10 seconds ▪ Aliquot a minimum of 1 mL of PreservCyt® specimen into a **cobas®** PCR Media Secondary tube ▪ Transfer tube to rack ● Load sample racks Confirm samples have been accepted into the system The system prepares automatically Order Tests ● Choose “Swab” for ordering swab specimens collected in **cobas®** PCR Media ● Choose “Urine” for ordering urine specimens collected in **cobas®** PCR Media ● Choose “PreservCyt®” for ordering PreservCyt® Solution specimens Choose the Test name
3	Refill reagents and consumables ● Load test specific reagent cassette ● Load control mini racks ● Load processing tips ● Load elution tips ● Load processing plates ● Load amplification plates ● Load Liquid waste plates ● Load MGP Reagent Refill Specimen Diluent Refill Lysis Reagent Refill Wash Reagent
4	Start the run by choosing the Start processing button on the user interface, all subsequent runs will start automatically if not manually postponed
5	Review and export results
6	Remove sample tubes. If needed, cap any sample tubes meeting the minimum volume requirements for future use. Clean up instrument ● Unload empty control mini racks ● Unload empty test specific reagent cassette(s) ● Empty amplification plate drawer ● Empty liquid waste ● Empty solid waste

Running cobas® CT/NG on cobas® 6800/8800 Systems

cobas® CT/NG can be run with a minimum required sample volume of 1.0 mL for swab and PreservCyt® specimens, and 1.2 mL for urine specimens. The operation of the instrument is described in detail in the **cobas**® 6800/8800 Systems User Guide. Figure 2 below summarizes the procedure.

- Swab and Urine specimens should be uncapped and loaded directly onto racks for processing on the **cobas**® 6800/8800 Systems.
- It is necessary to aliquot specimens collected in PreservCyt® Solution. Refer to the preparation instructions for cervical specimens found in section: "Cervical specimens in PreservCyt® Solution"
- A single run can have any combination of specimens (Swab, Urine, and PreservCyt®) and each specimen can be tested with either the CT/NG, CT, or NG ASAPs.
- Specimens collected in **cobas**® PCR Media or PreservCyt® Solution should be processed using the sample type selection in the user interface (UI) of the **cobas**® CT/NG as described in Table 15.

Table 15 Sample type selection in the user interface of the **cobas**® CT/NG

Specimen		**Collection kit type**	**Process as Sample Type**
Female	Vaginal swab	**cobas**® PCR Media Uni or Dual Swab Sample Kit	**Swab**
	Endocervical swab	**cobas**® PCR Media Dual Swab Sample Kit	**Swab**
	Oropharyngeal swab	**cobas**® PCR Media Uni or Dual Swab Sample Kit	**Swab**
	Anorectal swab	**cobas**® PCR Media Uni or Dual Swab Sample Kit	**Swab**
	Urine	**cobas**® PCR Urine Sample Kit or **cobas**® PCR Media Kit	**Urine**
	Cervical swab	PreservCyt® Solution (ThinPrep)	**PreservCyt**®
Male	Oropharyngeal swab	**cobas**® PCR Media Uni or Dual Swab Sample Kit	**Swab**
	Anorectal swab	**cobas**® PCR Media Uni or Dual Swab Sample Kit	**Swab**
	Urine	**cobas**® PCR Urine Sample Kit or **cobas**® PCR Media Kit	**Urine**

Figure 2 **cobas®** CT/NG procedure on **cobas®** 6800/8800 Systems

1

Log onto the system
Press Start to prepare the system
Order Tests
- Choose “Swab” for ordering swab specimens collected in **cobas®** PCR Media
- Choose “Urine” for ordering urine specimens collected in **cobas®** PCR Media
- Choose “PreservCyt” for ordering PreservCyt® Solution specimens
- Choose the Test

2

Refill reagents and consumables as prompted by the system
- Load test specific reagent cassette
- Load control cassettes
- Load pipette tips
- Load processing plates
- Load MGP Reagent
- Load amplification plates
- Refill Specimen Diluent
- Refill Lysis Reagent
- Refill Wash Reagent

3

Loading specimens onto the system
- For each primary urine or swab in **cobas®** PCR Media
  - Uncap tube
  - Transfer tube directly to rack
- For each primary PreservCyt® specimen vial:
  - Vortex for 10 seconds
  - Aliquot a minimum of 1 mL of PreservCyt® specimen into a 13 mL round-bottom secondary tube
  - Transfer tube to rack
- Load sample rack and clot tip racks into the sample supply module
- Confirm samples have been accepted into the transfer module

4

Start run by choosing the Start manually button on the user interface or have it start automatically after 120 minutes or if the batch is full

5

Review and export results

6

Remove sample tubes. If needed, cap any sample tubes meeting the minimum volume requirements for future use.

Clean up instrument
- Unload empty control cassettes
- Empty amplification plate drawer
- Empty liquid waste
- Empty solid waste

Results

The **cobas**® 5800 System and **cobas**® 6800/8800 Systems automatically detects and discriminates CT and/or NG DNA simultaneously for samples and controls, displaying individual target results for samples as well as validity for controls.

Quality control and validity of results on the cobas® 5800 System

- One **cobas**® Buffer Negative Control [(-) Ctrl] and one CT/NG Positive Control [CT/NG (+) C] must be processed at least every 72 hours or with every new kit lot. Positive and/or negative controls can be scheduled more frequently based on laboratory procedures and/or local regulations.
- The results of the controls are shown in the **cobas**® 5800 software in the "Controls" app.
- In the **cobas**® 5800 System software and/or report, check for flags to ensure the validity of the corresponding test results (Refer to the x800 Data Manager User Assistance for a 'List of flag codes').
- The controls are valid if no flags appear for either control.
- Controls are marked with "Valid" in the column "Control result" if all Targets of the control are reported valid. Controls are marked with 'Invalid' in the column "Control result" if one or both Targets of the control are reported invalid.
- Controls marked with 'Invalid' show a flag in the "Flags" column. More information on why the control is reported invalid including flag information will be shown in the detail view. If the positive control is invalid, repeat testing the positive control and all associated samples. If the negative control is invalid, repeat testing all controls and all associated samples.

Invalidation of results is performed automatically by the **cobas**® 5800 software based on control results.

NOTE: The **cobas**® 5800 System will be delivered with the standard setting of running a set of controls (positive and negative) with every run, but can be configured to a less frequent scheduling up to every 72 hours based on laboratory procedures and/or local regulations. Please contact your Roche service engineer or Roche customer technical support for more information.

Quality control and validity of results on cobas® 6800/8800 Systems

- One **cobas**® Buffer Negative Control [(-) Ctrl] and one CT/NG Positive Control [CT/NG (+) C] are processed with each batch of a requested result type.
- In the **cobas**® 6800/8800 software and/or report, check for flags and their associated results to ensure batch validity.
- All flags are described in the **cobas**® 6800/8800 Systems User Assistance.
- The batch is valid if no flags appear for all controls. If the batch is invalid, repeat testing of the entire batch.

Validation of results is performed automatically by the **cobas**® 6800/8800 software based on negative and positive control performance.

cobas® CT/NG for cobas® 5800 System

The results of the samples are shown in the **cobas®** 5800 software in the "Results" app. Display examples **cobas®** CT/NG for **cobas®** 5800 System Software (Figure 3).

Figure 3 Example of **cobas®** CT/NG results display for **cobas®** 5800 System

Sample ID*	Test	Control results	Flag**	Result
CT/NG_01	CT/NG	Valid		**CT Negative NG Negative**
NG 01_	NG	Valid		**NG Positive (Ct 38.52)**
CT/NG_02	CT/NG	Valid		**CT Invalid NG Invalid**
CT 01	CT	Valid		**CT Negative**
CT/NG_03	CT/NG	Valid		**CT Positive (Ct 38.44) NG positive (Ct 37.00)**

* Table applies for all sample types used.

** The result overview shows a flag symbol in case of invalid results. Detailed flag descriptions are available in the result details.

Check each individual sample for flags in the **cobas®** 5800 System software and/or report. The result interpretation should be as follows:

- Samples associated with valid controls are shown as 'Valid' in the "Control result" column.
- Samples associated with a failed control are shown as 'Invalid' in the "Control result" column.
- If the associated controls of a sample result are invalid, a specific flag will be added to the sample result as follows:
    - Q05D : Result validation failure because of an invalid positive control
    - Q06D : Result validation failure because of an invalid negative control
- The values in "Results" column for individual sample target result should be interpreted as shown in Table 16, Table 17 and Table 18.
- If one or more sample targets are marked with "Invalid" the **cobas®** 5800 software shows a flag in the "Flags" column. More information on why the sample target(s) is reported invalid including flag information is shown in the detail view.
- Invalid results for one or more target combinations are possible with the CT/NG test and are reported out specifically for each channel. Refer to retesting instructions for the respective specimen type.
- Results of this test should only be interpreted in conjunction with information available from clinical evaluation of the patient and patient history.

cobas® CT/NG for cobas® 6800/8800 Systems

Display examples **cobas**® CT/NG **cobas**® 6800/8800 Systems are shown in Figure 4, Figure 5, and Figure 6 respectively.

Figure 4 Example of **cobas**® CT/NG results display for the CT/NG result request on **cobas**® 6800/8800 Systems

Test	Sample ID	Valid	Flags	Sample type	Overall result	Target 1	Target 2
CT/NG	C161420284084196207422	Yes		CT/NG (+) C	**Valid**	Valid	Valid
CT/NG	C161420284090419545972	Yes		(-) Ctrl	**Valid**	Valid	Valid
CT/NG 400 ul	CTNG_PC1	NA		PreservCyt®	**NA**	CT Positive	NG Positive
CT/NG 400 ul	CTNG_PC2	NA		PreservCyt®	**NA**	CT Negative	NG Positive
CT/NG 400 ul	CTNG_Swab1	NA		Swab	**NA**	CT Negative	NG Negative
CT/NG 400 ul	CTNG_Swab2	NA		Swab	**NA**	CT Positive	NG Positive
CT/NG 850 ul	CTNG_Urine1	NA		Urine	**NA**	CT Positive	NG Negative
CT/NG 850 ul	CTNG_Urine2	NA		Urine	**NA**	CT Negative	NG Negative
CT/NG 850 ul	CTNG_Urine3	NA	Y40T	Urine	**NA**	Invalid	Invalid

Figure 5 Example of **cobas**® CT results display for the CT result request on **cobas**® 6800/8800 Systems

Test	Sample ID	Valid	Flags	Sample type	Overall result	Target 1	Target 2
CT	C161420284084196207423	Yes		CT/NG (+) C	**Valid**	Valid	
CT	C161420284090419545973	Yes		(-) Ctrl	**Valid**	Valid	
CT 400 ul	CT_PC1	NA		PreservCyt®	**NA**	CT Positive	
CT 400 ul	CT_PC2	NA		PreservCyt®	**NA**	CT Positive	
CT 400 ul	CT_Swab1	NA		Swab	**NA**	CT Negative	
CT 400 ul	CT_Swab2	NA		Swab	**NA**	CT Positive	
CT 400 ul	CT_Swab3	NA	P02T	Swab	**NA**	Invalid	
CT 850 ul	CT_Urine1	NA		Urine	**NA**	CT Negative	
CT 850 ul	CT_Urine2	NA		Urine	**NA**	CT Positive	

Note: The Target 2 column is reserved for NG results.

Figure 6 Example of **cobas**® NG results display for the NG result request on **cobas**® 6800/8800 Systems

Test	Sample ID	Valid	Flags	Sample type	Overall result	Target 1	Target 2
NG	C161420284084196207424	Yes		CT/NG (+) C	**Valid**		Valid
NG	C161420284090419545974	Yes		(-) Ctrl	**Valid**		Valid
NG 400 ul	NG_PC1	NA		PreservCyt®	**NA**		NG Negative
NG 400 ul	NG_PC2	NA		PreservCyt®	**NA**		NG Positive
NG 400 ul	NG_PC3	NA	Y40T	PreservCyt®	**NA**		Invalid
NG 400 ul	NG_Swab1	NA		Swab	**NA**		NG Positive
NG 400 ul	NG_Swab2	NA		Swab	**NA**		NG Negative
NG 850 ul	NG_Urine1	NA		Urine	**NA**		NG Negative
NG 850 ul	NG_Urine2	NA		Urine	**NA**		NG Positive

Note: The Target 1 column is reserved for CT results.

For a valid batch, check each individual sample for flags in the **cobas**® 6800/8800 software and/or report. The result interpretation should be as follows:

- A valid batch may include both valid and invalid sample results.
- The "Valid" and "Overall Result" columns are not applicable (NA) to sample results for the **cobas**® CT/NG Test and are marked with "NA".
- Reported target results for individual samples are valid unless indicated as "Invalid" within the individual target result column.
- Invalid results for one or more target combinations are possible with the CT/NG test and are reported out specifically for each channel. Refer to retesting instructions for the respective specimen type.
- Results of this test should only be interpreted in conjunction with information available from clinical evaluation of the patient and patient history.

Interpretation of results

Results and their corresponding interpretation for detecting CT and NG (Table 16), CT only (Table 17) and NG only (Table 18) are shown below.

Table 16 cobas® CT/NG results and interpretation for the CT/NG result request

Target 1	Target 2	Interpretation
CT Positive	NG Positive	All requested results were valid. Target signal detected for CT and NG DNA.
CT Positive	NG Negative	All requested results were valid. Target signal detected for CT DNA. No target signal detected for NG DNA.
CT Negative	NG Positive	All requested results were valid. No target signal detected for CT DNA. Target signal detected for NG DNA.
CT Negative	NG Negative	All requested results were valid. No target signal detected for CT or NG DNA.
CT Positive	Invalid	Not all requested results were valid. CT result is valid. Target signal detected for CT DNA. NG result is invalid. Original specimen should be re-tested to obtain valid NG results. If the result is still invalid, a new specimen should be obtained.
Invalid	NG Positive	Not all requested results were valid. CT result is invalid. Original specimen should be re-tested to obtain valid CT results. If the result is still invalid, a new specimen should be obtained. NG result is valid. Target signal detected for NG DNA.
CT Negative	Invalid	Not all requested results were valid. CT result is valid. No target signal detected for CT DNA. NG result is invalid. Original specimen should be re-tested to obtain valid NG results. If the result is still invalid, a new specimen should be obtained.
Invalid	NG Negative	Not all requested results were valid. CT result is invalid. Original specimen should be re-tested to obtain valid CT results. If the result is still invalid, a new specimen should be obtained. NG result is valid. No target signal detected for NG DNA.
Invalid	Invalid	Both CT and NG results are invalid. Original specimen should be re-tested to obtain valid CT and NG results. If the results are still invalid, a new specimen should be obtained.

Table 17 cobas® CT/NG results and interpretation for the CT result request

Result	Interpretation
CT Positive	The requested result was valid. Target signal detected for CT DNA.
CT Negative	The requested result was valid. No target signal detected for CT DNA
CT Invalid	CT result is invalid. Original specimen should be re-tested to obtain valid CT results. If the result is still invalid, a new specimen should be obtained.

Table 18 cobas® CT/NG results and interpretation for the NG result request

Result	Interpretation
NG Positive	The requested result was valid. Target signal detected for NG DNA.
NG Negative	The requested result was valid. No target signal detected for NG DNA
Invalid	NG result is invalid. Original specimen should be re-tested to obtain valid NG results. If the result is still invalid, a new specimen should be obtained.

Procedural limitations

- Use of this product must be limited to personnel trained in the techniques of PCR and the use of the **cobas**® 5800 System or **cobas**® 6800/8800 Systems.
- **cobas**® CT/NG has been evaluated only for use in combination with the **cobas**® CT/NG Positive Control Kit, **cobas**® Buffer Negative Control Kit, **cobas**® **omni** MGP Reagent, **cobas**® **omni** Lysis Reagent, **cobas**® **omni** Specimen Diluent, and **cobas**® **omni** Wash Reagent for use on the **cobas**® 5800 System or **cobas**® 6800/8800 Systems.
- Reliable results depend on proper sample collection, storage and handling procedures.
- Products containing carbomer(s), including vaginal lubricants, creams and gels may interfere with the test and should not be used during or prior to collecting urogenital specimens. See Interference results (Table 23) for further details.
- **cobas**® CT/NG has only been validated for use with male and female urine, clinician-instructed self-collected vaginal swab specimens, clinician-collected vaginal swab specimens, anorectal swab specimens, oropharyngeal swab specimens and endocervical swab specimens, all collected in **cobas**® PCR Media (Roche Molecular Systems, Inc.) and cervical specimens collected in PreservCyt® Solution. Assay performance has not been validated for use with other collection media and/or specimen types.
- Detection of *C. trachomatis* and *N. gonorrhoeae* is dependent on the number of organisms present in the specimen and may be affected by specimen collection methods, patient factors (i.e., age, history of STD, presence of symptoms), stage of infection and/or infecting *C. trachomatis* and *N. gonorrhoeae* strains.
- Though rare, mutations within the highly conserved regions of the cryptic plasmid or genomic DNA of *C. trachomatis* or the genomic DNA of *N. gonorrhoeae* covered by **cobas**® CT/NG primers and/or probes may result in failure to detect the presence of the bacterium.

- *Neisseria gonorrhoeae* may occasionally exchange genetic material with commensal bacteria commonly found in the normal microflora of the mouth and throat. It is possible that this exchange may include isolated DNA sequences which could, on rare occasion, produce a positive signal with this assay.[22]
- Due to inherent differences between technologies, it is recommended that, prior to switching from one technology to the next, users perform method correlation studies in their laboratory to qualify technology differences. Users should follow their own specific policies/procedures.
- **cobas**® CT/NG is not intended to replace other exams or tests for diagnosis of urogenital infection. Patients may have cervicitis, urethritis, urinary tract infections, or vaginal infections due to other causes or concurrent infections with other agents.
- **cobas**® CT/NG is not recommended for evaluation of suspected sexual abuse and for other medico-legal indications.
- **cobas**® CT/NG should not be used to determine therapeutic success as nucleic acids may be present after antimicrobial therapy.
- **cobas**® CT/NG for urine testing is recommended to be performed on first catch urine specimens (defined as the first 10 to 50 mL of the urine stream). The effects of other variables such as first-catch vs. mid-stream, post-douching, etc. have not been evaluated.
- The effects of other potential variables such as vaginal discharge, use of tampons, douching, etc. and specimen collection variables have not been evaluated.
- **cobas**® CT/NG has not been evaluated with patients who were currently being treated with antimicrobial agents active against CT or NG as well as patients with a history of hysterectomy.
- False negative or invalid results may occur due to polymerase inhibition. The CT/NG Internal Control is included in **cobas**® CT/NG to help identify the specimens containing substances that may interfere with nucleic acid isolation and PCR amplification.
- The addition of AmpErase enzyme into the **cobas**® CT/NG Master Mix reagent enables selective amplification of target DNA; however, good laboratory practices and careful adherence to the procedures specified in this Package Insert are necessary to avoid contamination of reagents.
- **cobas**® CT/NG has not been evaluated in patients younger than 14 years of age.

Non-clinical performance evaluation

Key performance characteristics performed on cobas® 6800/8800 Systems

Limit of Detection (LoD)

The *Chlamydia trachomatis* analytical sensitivity claim for the assay is 40 Elementary Bodies (EB) per mL for all serovars (A, B, Ba, C, D, E, F, G, H, I, J, K, L1, L2, L3) as well as for the Swedish variant nvCT, in all claimed specimen types. However, dilutions of some serovars below 40 EB/mL have tested positive using **cobas**® CT/NG for use on the **cobas**® 6800/8800 Systems.

The *N. gonorrhoeae* analytical sensitivity claim for the assay is 1.0 Colony Forming Units (CFU) per mL (45 gonorrhoeae strains tested) in all claimed specimen types. However, dilutions of gonorrhoeae strains below 1.0 CFU/mL have tested positive using **cobas**® CT/NG for use on the **cobas**® 6800/8800 Systems.

Precision

In-house precision was examined using a panel composed of CT and NG cultures diluted into a pool of negative endocervical swab specimen matrix collected in **cobas**® PCR Media, a pool of negative urine matrix plus **cobas**® PCR Media and a pool of negative cervical specimen matrix collected in PreservCyt® Solution. Endocervical swabs were intended to represent all swab samples collected in **cobas**® PCR Media (endocervical, vaginal, oropharyngeal, and anorectal). Four levels were tested using CT serovar D and NG strain 2948 (ATCC 19424) as the target organisms.

The precision panel was designed to include members with very low, low and medium concentrations of CT and NG (≤ 0.7 EB/mL and ≤ 0.07 CFU/mL, ≤ 4 EB/mL and ≤ 0.4 CFU/mL and ≤ 12 EB/mL and ≤ 1.2 CFU/mL) for each panel matrix. Testing was performed with three lots of **cobas**® CT/NG reagents and two instruments for a total of 24 runs. A description of the precision panels and the study performance hit rate is shown in Table 19. All negative panel members tested negative throughout the study. Analysis of standard deviation and percent coefficient of variation of the Ct values from valid tests performed on positive panel members (see Table 20 and Table 21) yielded overall CV (%) ranges from 1.62% to 4.05% for CT and from 1.17% to 3.55% for NG.

Table 19 Summary of within laboratory precision

Target Concentration		N Tested	N positive CT	N positive NG	Hit Rate		95% CI CT		95% CI NG	
CT	**NG**				**CT**	**NG**	**LL**	**UL**	**LL**	**UL**
Endocervical Swab in **cobas**® PCR Media										
Neg	Neg	72	0	0	0%	0%	0.0	5.0	0.0	5.0
0.7 EB/mL	0.07 CFU/mL	72	51	32	71%	44%	59	81	33	57
2 EB/mL	0.4 CFU/mL	72	69	68	96%	94%	88	99	86	98
6 EB/mL	1.2 CFU/mL	72	72	72	100%	100%	95	100	95	100
cobas® PCR Media with Urine										
Neg	Neg	72	0	0	0%	0%	0.0	5.0	0.0	5.0
0.3 EB/mL	0.05 CFU/mL	72	38	47	53%	65%	66	87	66	87
1 EB/mL	0.2 CFU/mL	72	72	69	100%	96%	92	100	95	100
3 EB/mL	0.6 CFU/mL	72	72	72	100%	100%	95	100	95	100
Cervical samples collected into PreservCyt® Solution										
Neg	Neg	72	0	0	0%	0%	0.0	5.0	0.0	5.0
0.7 EB/mL	0.07 CFU/mL	72	56	56	78%	78%	41	65	53	76
4 EB/mL	0.2 CFU/mL	72	71	72	99%	100%	95	100	88	99
12 EB/mL	0.6 CFU/mL	72	72	72	100%	100%	95	100	95	100

Table 20 Overall mean, standard deviations and coefficients of variation (%) for cycle threshold, CT panel members 2, 3, and 4

Hit Rate	Mean Ct	Between instrument		Between lot		Within run		Between run		Between day		Total	
		SD	CV%	SD	CV%	SD	CV%	SD	CV%	SD	CV%	SD	CV%
Endocervical Swab in **cobas®** PCR Media													
71%	39.7	0.00	0.00	0.00	0.00	1.27	3.21	0.00	0.00	0.34	0.85	1.32	3.32
96%	38.5	0.00	0.00	0.04	0.10	1.14	2.96	0.00	0.00	0.48	1.25	1.24	3.22
100%	36.9	0.00	0.00	0.25	0.69	0.54	1.45	0.07	0.18	0.00	0.00	0.60	1.62
Cervical Samples collected into PreservCyt® Solution													
53%	38.3	0.60	1.57	0.52	1.37	1.12	2.92	0.00	0.00	0.00	0.00	1.37	3.58
100%	36.9	0.21	0.56	0.28	0.76	0.68	1.85	0.00	0.00	0.00	0.00	0.77	2.08
100%	35.6	0.00	0.00	0.20	0.56	0.52	1.46	0.09	0.24	0.02	0.05	0.56	1.59
cobas® PCR Media with Urine													
78%	38.9	0.00	0.00	0.12	0.30	1.25	3.22	0.39	1.01	0.00	0.00	1.32	3.39
99%	38.3	0.11	0.28	0.00	0.00	1.52	3.97	0.00	0.00	0.29	0.77	1.55	4.05
100%	37.1	0.00	0.00	0.00	0.00	1.05	2.84	0.00	0.00	0.28	0.77	1.09	2.94

Table 21 Overall mean, standard deviations and coefficients of variation (%) for cycle threshold, NG panel members 2, 3, and 4

Hit Rate	Mean Ct	Between instrument		Between lot		Within run		Between run		Between day		Total	
		SD	CV%	SD	CV%	SD	CV%	SD	CV%	SD	CV%	SD	CV%
Endocervical Swab in **cobas®** PCR Media													
44%	39.1	0.00	0.00	0.31	0.79	0.84	2.14	0.72	1.85	0.57	1.46	1.28	3.28
94%	38.1	0.00	0.00	0.00	0.00	1.27	3.34	0.00	0.00	0.00	0.00	1.27	3.34
100%	36.5	0.00	0.00	0.24	0.67	0.69	1.89	0.00	0.00	0.15	0.40	0.74	2.04
Cervical Samples collected into PreservCyt® Solution													
65%	39.0	0.34	0.87	0.00	0.00	1.11	2.85	0.08	0.20	0.45	1.16	1.25	3.21
96%	38.0	0.00	0.00	0.00	0.00	1.25	3.28	0.00	0.00	0.00	0.00	1.25	3.28
100%	35.8	0.00	0.00	0.28	0.78	0.76	2.13	0.00	0.00	0.00	0.00	0.81	2.27
cobas® PCR Media with Urine													
78%	39.1	0.00	0.00	0.26	0.66	1.35	3.46	0.00	0.00	0.18	0.45	1.39	3.55
100%	36.7	0.14	0.38	0.16	0.42	0.71	1.92	0.00	0.00	0.00	0.00	0.74	2.00
100%	34.9	0.00	0.00	0.16	0.47	0.37	1.06	0.06	0.18	0.00	0.00	0.41	1.17

Analytical specificity/cross reactivity

A panel of 151 bacteria, fungi and viruses, including those commonly found in the male and female urogenital tract, 17 representatives of non-*gonorrhoeae Neisseria* strains and other phylogenetically unrelated organisms, were tested with **cobas**® CT/NG to assess analytical specificity. The organisms listed in Table 22 were spiked at concentrations of approximately 1 x 10^6 units*/mL for bacteria and approximately 1 x 10^5 units*/mL for viruses into pools of negative swab specimens in **cobas**® PCR Media (endocervical, oropharyngeal, and anorectal), urine stabilized in **cobas**® PCR Media and cervical specimens in PreservCyt® Solution. Testing was performed with each potential interfering organism alone as well as with each organism mixed with CT and NG cultures at ≤ 12 EB/mL and ≤ 1.2 CFU/mL. Results indicated that none of these organisms interfered with the detection of CT and NG or produced false positive results in the CT/NG negative matrices.

*All bacteria were quantified as Colony Forming Units (CFU) except *Chlamydophila pneumonia* and *Chlamydophila psittaci* as Elementary Bodies (EB). All viruses were quantified as units/mL as determined by $TCID_{50}$ Endpoint Dilution Assay. *Trichomonas vaginalis* and HPV16 were quantified as cells/mL.

Table 22 Microorganisms tested for analytical specificity/cross reactivity

Achromobacter xerosis	*Haemophilus ducreyi*	*Neisseria polysaccharea*
Acinetobacter calcoaceticus	*Haemophilus influenzae*	*Neisseria sicca*
Acinetobacter lwoffi	*Helicobacter pylori*	*Neisseria subflava*
Actinomyces israelii	HPV 16	*Neisseria weaverii*
Actinomyces pyogenes	HSV-1	*Paracoccus denitrificans*
Aerococcus viridans	HSV-2	*Peptostreptococcus anaerobius*
Aeromonas hydrophila	Human Adenovirus 40	*Peptostreptococcus asaccharolyticus*
Alcaligenes faecalis	Human Enterovirus 71	*Peptostreptococcus magnus*
Bacillus subtilis	Human Rotavirus	*Plesiomonas shigelloides*
Bacteriodes fragilis	*Kingella dentrificans*	*Propionibacterium acnes*
Bacteroides caccae	*Kingella kingae*	*Proteus mirabilis*
Bacteroides ureolyticus	*Klebsiella oxytoca*	*Proteus penneri*
Bifidobacterium adolescentis	*Klebsiella pneumoniae*	*Proteus vulgaris*
Bifidobacterium breve	*Lactobacillus acidophillus*	*Providencia rettgeri*
Bifidobacterium longum	*Lactobacillus brevis*	*Providencia stuartii*
Blautia producta	*Lactobacillus crispatus*	*Pseudomonas aeruginosa*
Branhamella catarrhalis	*Lactobacillus delbrueckii subsp. lactis*	*Pseudomonas fluorescens*
Brevibacterium linens	*Lactobacillus jensenii*	*Pseudomonas putida*
Campylobacter coli	*Lactobacillus lactis*	*Rahnella aquatilis*
Campylobacter jejuni	*Lactobacillus oris*	*Rhizobium radiobacter*
Candida albicans	*Lactobacillus parabuchnerri*	*Rhodospirillum rubrum*
Candida glabrata	*Lactobacillus reuteri*	*Saccharomyces cerevisiae*
Candida parapsilosis	*Lactobacillus vaginalis*	*Salmonella choleraesuis*
Candida tropicalis	*Lactococcus lactis cremoris*	*Salmonella minnesota*
Chlamydia psittaci	*Legionella pneumophila*	*Salmonella typhimurium*
Chlamydophila pneumoniae	*Leuconostoc paramensenteroides aka. Weissella*	*Serratia denitrificans*
Chromobacter violaceum	*Listeria monocytogenes*	*Serratia marcescens*
Citrobacter freundii	*Micrococcus luteus*	*Shigella dysenteriae*

Clostridioides difficile (Serogroup B)	*Moraxella lacunata*	*Staphylococcus aureus*
Clostridium perfringens	*Moraxella osloensis*	*Staphylococcus epidermidis*
Corynebacterium genitalium	*Morganella morganii*	*Staphylococcus saprophyticus*
Corynebacterium xerosis	*Mycobacterium smegmatis*	*Streptococcus agalactiae*
Cryptococcus neoformans	*Mycoplasma genitalium*	*Streptococcus anginosus*
Cytomegalovirus	*Mycoplasma hominis*	*Streptococcus bovis*
Deinococcus radiodurans	*Neisseria cinerea*	*Streptococcus dysgalactiae*
Derxia gummosa	*Neisseria dentrificans*	*Streptococcus equinis*
Eikenella corrodens	*Neisseria elongata subsp. elongata*	*Streptococcus mitis*
Enterobacter aerogenes	*Neisseria elongata subsp. niroreducans*	*Streptococcus mutans*
Enterobacter cloacae	*Neisseria flava*	*Streptococcus pneumoniae*
Enterococcus avium	*Neisseria flavescens*	*Streptococcus pyogenes*
Enterococcus casseliflavus	*Neisseria kochi*	*Streptococcus salivarius*
Enterococcus faecalis	*Neisseria lactamica*	*Streptococcus sanguis*
Enterococcus faecium	*Neisseria macacae*	*Streptomyces griseinus*
Erwinia herbicola	*Neisseria meningitidis Serogroup A*	*Trichomonas vaginalis*
Erysipelothrix rhusiopathiae	*Neisseria meningitidis Serogroup B*	*Ureaplasma urealyticum*
Escherichia coli	*Neisseria meningitidis Serogroup C*	*Veillonela parvula*
Escherichia fergusonii	*Neisseria meningitidis Serogroup D*	*Vibrio cholerae*
Flavobacterium meningosepticum	*Neisseria meningitidis Serogroup W135*	*Vibrio parahaemolyticus*
Fusobacterium nucleatum	*Neisseria meningitidis Serogroup Y*	*Yersinia enterocolitica*
Gardnerella vaginalis	*Neisseria mucosa*	
Gemella haemolysans	*Neisseria perflava*	

Interference

The effect of over-the-counter or prescription feminine products that may be present in urogenital specimens (Table 23), over-the-counter oral hygiene products that may be present in oropharyngeal specimens (Table 24) and of hygiene and prescription products that may be present in anorectal specimens (Table 26) were evaluated. Testing was done using pooled clinical specimens (vaginal swab, urine and PreservCyt® specimens were used to represent urogenital) with spiking of potential interferents at levels expected from normal patient usage. Interferents were tested in CT/NG negative specimen pools as well as in specimen pools with CT/NG at ≤ 120 EB/mL and ≤ 1.2 CFU/mL, depending on the specimen type tested. CT serovars D and I and NG strains 2948 (ATCC 19424) and 891 were used in this study.

Of the over-the-counter (OTC) feminine hygiene and prescription products tested in urogenital specimens, Metronidazole, Replens, RepHresh Odor Eliminating Vaginal Gel and RepHresh Clean Balance produced false negative or invalid results. These products contain carbomer(s). Products containing carbomer(s) have been shown to generate false negative and invalid results. Table 23 is not intended to be a comprehensive list of carbomer containing products. None of the OTC oral hygiene products tested in oropharyngeal swabs or the OTC anorectal hygiene and prescription products tested in anorectal swabs produced interference to the test when examined at concentrations expected through typical product use.

Table 23 List of substances tested for interference in urogenital specimens

Product Name	
Clindamycin Phosphate Vaginal Cream	Norforms Suppositories
CVS Tioconazole 1 (Equate tioconazole 1)	Premarin
Equate Vagicaine Anti-Itch Cream	Replens Long-Lasting Vaginal Moisturizer*
Estrace	Summer's Eve Feminine Deodorant Spray
K-Y Ultra Gel (Replaces K-Y Silk E)	VCF - Vaginal Contraceptive Foam
Metronidazole Vaginal Gel *	Yeast Gard Advanced
Monistat 3 Vaginal Antifungal Combination Pack	Azo Standard (urine only)
Monistat Complete Care Itch Relief Cream	RepHresh Odor Eliminating Vaginal Gel*‡
Gyne-Lotrimin 7	RepHresh Clean Balance*‡

* Metronidazole, Replens and RepHresh showed interference at levels that may potentially be present in clinical specimens

‡ RepHresh products were tested using simulated swab specimen

Table 24 List of substances tested for interference in oropharyngeal swab specimens

Product Name
Cepacol Maximum Strength Throat Drop Lozenges
Colgate Total Toothpaste
Robitussin Cough / Chest Congestion Cough Syrup
Listerine Ultra Clean Antiseptic Mouthwash
Scope Mouthwash
Sucrets Complete Lozenges
Vicks - Chloraseptic Sore Throat Spray Menthol
Zicam Oral Mist

Table 25 List of substances tested for interference in anorectal swab specimens

Product Name
ANUSOL® Plus Ointment
CB Fleet® Mineral Oil Enema
Doproct Suppositories/ Hemorrhoidal Treatment
K-Y Jelly
Lotrimin Antifungal Cream
Preparation H Hemorrhoidal Ointment
PREPARATION H Hemorrhoidal Suppositories
Driminate Generic for Dramamine Motion Sickness - Major Pharmaceuticals
Target - Triple Paste Diaper Rash Ointment
Tucks Medicated Cooling Hemorrhoidal Pads
Vaseline Original Petroleum Jelly

Endogenous substances that may be present in urogenital, oropharyngeal and anorectal specimens were tested for interference. Testing was done using pooled clinical specimens (endocervical swab, urine and PreservCyt® specimens were used to represent urogenital) with spiking of potential endogenous interferents. Interferents were tested in CT/NG negative specimen pools as well as in the presence of CT/NG at ≤ 120 EB/mL and ≤ 1.2 CFU/mL, depending on the specimen type tested. CT serovars D and I and NG strains 2948 (ATCC 19424) and 891 were used in this study.

Interference was noted with whole blood at 10% for urine and PreservCyt® specimens, with stool at 0.4% in anorectal specimens and with cervical mucus at 1% in endocervical specimens. Levels of endogenous substances tolerated by the assay for all specimen types are shown in Table 26.

Table 26 Summary of endogenous substance concentrations that do not show interference

Interferent	Endocervical Swab	Anorectal Swab	Oropharyngeal Swab	PreservCyt®	Urine
Albumin (% w/v)	N/A	N/A	N/A	N/A	5%
Bilirubin (% w/v)	N/A	N/A	N/A	N/A	0.5%
Mucus (% w/v)	0.5%	1.0%	1.0%	1.0%	0.5%
Glucose (% w/v)	N/A	N/A	N/A	N/A	1.0%
Peripheral Blood Mononuclear Cells (PBMCs as cells/mL)	1.0E+06	1.0E+06	1.0E+06	1.0E+06	1.0E+06
pH (acidic and alkaline)	N/A	N/A	N/A	N/A	pH 4 and pH 9
Saliva (% w/v)	N/A	N/A	2.0%	N/A	N/A
Semen (% w/v)	1.5%	N/A	N/A	1.5%	N/A
Stool (% w/v)	N/A	0.3%	N/A	N/A	N/A
Whole Blood (% v/v)	10%	10%	10%	5%	5%

Competitive inhibition

To assess competitive inhibition between CT and NG, samples of each specimen type (vaginal, oropharyngeal, and anorectal swabs in **cobas**® PCR Media, urine stabilized in **cobas**® PCR Media and cervical specimens in PreservCyt® Solution) were tested where low and moderate concentrations of one target were mixed with very high concentrations of the opposite target. Low and moderate concentrations were defined as ~1x LoD and ~3x LoD, respectively, and high concentrations were defined as generating a signal greater than in 95% of target positive specimens.

Testing results indicated that when NG was present at a high concentration, CT was detected in all specimen types, at both low (~1x LoD) and moderate (~3x LoD) levels. Results also indicated that when CT was present at a high concentration, NG was detected in all specimen types at moderate (~3x LoD) levels, and in all but one (oropharyngeal) specimen type, at low (~1x LoD) levels.

Whole system failure

The samples tested in the whole system failure study were pooled CT and NG negative clinical cervical specimens collected in PreservCyt® Solution, vaginal swab specimens collected in **cobas**® PCR Media and urine specimens stabilized in **cobas**® PCR Media. Each pool of clinical specimens was spiked with cultures of CT, serovar D (D-UW3) (CT) and NG 2948 (ATCC 19424) (NG) to a concentration of ≤ 12 EB/mL and ≤ 1.2 CFU/mL, depending on the sample type. The results of this study determined that all replicates were valid and positive for CT/NG, resulting in a whole system failure rate of 0%. The two-sided 95% exact confidence interval was 0% for the lower bound and 3.6% for the upper bound [0%: 3.6%].

Cross contamination

Studies were performed to evaluate potential cross contamination for **cobas**® CT/NG. Cross-contamination can cause false positive results. In this performance study the sample to sample cross-contamination rate of **cobas**® CT/NG has been determined to be 0.5% (2/432) when alternating very high positive and negative samples were tested over multiple runs. Run to run cross-contamination has not been observed (0/282). Testing was done using samples prepared with **cobas**® PCR Media, urine stabilized in **cobas**® PCR Media and with PreservCyt® Solution on the **cobas**® 6800/8800 Systems. High positive samples in the study were prepared to generate a Ct value that exceeds 95% or more of signal obtained from specimens of infected patients in the intended use population. The likelihood of encountering such specimens in the routine use of **cobas**® CT/NG is proportional to CT and NG prevalence in the testing population. Therefore the sample to sample cross-contamination rate in routine use of **cobas**® CT/NG will likely be less than 0.5% x 5% x CT prevalence in the testing population. Even at a maximum prevalence of 100%, the cross-contamination rate would be 0.5% x 5% x 100% = 0.025%.

Clinical performance evaluation

Prospective collection method correlation

The performance of **cobas**® CT/NG and the **cobas**® 4800 CT/NG Test were compared by analysis of the following specimen types:

- Endocervical swabs in **cobas**® PCR Media
- Vaginal swabs (clinician-collected) in **cobas**® PCR Media
- Vaginal swabs (self-collected) in **cobas**® PCR Media
- Oropharyngeal swabs in **cobas**® PCR Media
- Anorectal swabs in **cobas**® PCR Media
- Male and female urine mixed with **cobas**® PCR Media
- Cervical specimens collected in PreservCyt® Solution

A total of 6,318 subjects were recruited from 19 clinical sites in Germany and the US, producing 13,433 valid CT results and 13,398 valid NG results which were used for the correlation study analysis. The correlation results for all specimen types are shown in Table 27 and the calculated PPA, NPA, OPA with 95% Confidence Intervals are shown in Table 28. Across all specimen types, there were 125 discrepant specimens for *Chlamydia trachomatis*; of which 120 were positive on the **cobas**® 6800/8800 Systems and 5 were positive on the **cobas**® 4800 System. Also across all specimen types, there were 42 discrepant specimens for *Neisseria gonorrhoeae*, of which 40 were positive on the **cobas**® 6800/8800 Systems and 2 were positive on the **cobas**® 4800 System.

Correlation analysis between **cobas**® CT/NG and the **cobas**® 4800 CT/NG Test shows Positive Percent Agreements (PPA) greater than 95% for both CT and NG in all specimen types with the majority of specimen types having a PPA of 100% for both CT and NG. Negative and Overall Percent Agreements were greater than 98% for both CT and NG in all specimen types.

Table 27 Results summary for correlation of **cobas**® CT/NG and the **cobas**® 4800 CT/NG Test

Specimen Type	Chlamydia trachomatis				Neisseria gonorrhoeae			
	Con +	Con -	68+ /48 -	68 -/48 +	Con +	Con -	68+ /48 -	68 -/48 +
Endocervical Swab	114	1778	15	0	22	1883	1	1
Vaginal Swab	87	1040	15	0	20	1111	1	0
SC-Vaginal Swab	90	1028	14	0	18	1100	3	0
Oropharyngeal Swab	37	1915	14	0	74	1864	22	0
Anorectal Swab	100	1871	30	0	71	1923	8	0
Female Urine	272	2083	18	0	23	2340	4	0
Male Urine	114	717	3	0	30	803	0	1
PreservCyt®	157	1905	11	5	25	2049	1	0
All Specimens Total	971	12337	120	5	283	13073	40	2

Con = Concordant; + = Positive; - = Negative; SC = Self-Collected

Table 28 Agreement calculations for correlation of **cobas**® CT/NG and the **cobas**® 4800 CT/NG Test

Specimen Type	*Chlamydia trachomatis*			*Neisseria gonorrhoeae*		
	Result (%)		95% CI	Result		95% CI
Endocervical Swab	PPA	100%	96.8%-100%	PPA	95.7%	78.1%-99.9%
	NPA	99.2%	98.6%-99.5%	NPA	99.9%	99.7%-100%
	OPA	99.2%	98.7%-99.6%	OPA	99.9%	99.6%-100%
Vaginal Swab	PPA	100%	95.8%-100%	PPA	100%	83.2%-100%
	NPA	98.6%	97.7%-99.2%	NPA	99.9%	99.5%-100%
	OPA	98.7%	97.8%-99.3%	OPA	99.9%	99.5%-100%
SC-Vaginal Swab	PPA	100%	96.0%-100%	PPA	100%	81.5%-100%
	NPA	98.7%	97.8%-99.3%	NPA	99.7%	99.2%-99.9%
	OPA	98.8%	97.9%-99.3%	OPA	99.7%	99.2%-99.9%
Oropharyngeal Swab	PPA	100%	90.5%-100%	PPA	100%	95.1%-100%
	NPA	99.3%	98.8%-99.6%	NPA	98.8%	98.2%-99.3%
	OPA	99.3%	98.8%-99.6%	OPA	98.9%	98.3%-99.3%
Anorectal Swab	PPA	100%	96.4%-100%	PPA	100%	94.9%-100%
	NPA	98.4%	97.8%-98.9%	NPA	99.6%	99.2%-99.8%
	OPA	98.5%	97.9%-99.0%	OPA	99.6%	99.2%-99.8%
Female Urine	PPA	100%	98.7%-100%	PPA	100%	85.2%-100%
	NPA	99.1%	98.6%-99.5%	NPA	99.8%	99.6%-100%
	OPA	99.2%	98.8%-99.5%	OPA	99.8%	99.6%-100%
Male Urine	PPA	100%	96.8%-100%	PPA	96.8%	83.3%-99.9%
	NPA	99.6%	98.8%-99.9%	NPA	100%	99.5%-100%
	OPA	99.6%	99.0%-99.9%	OPA	99.9%	99.3%-100%
PreservCyt®	PPA	96.9%	92.9%-99.0%	PPA	100%	86.3%-100%
	NPA	99.4%	99.0%-99.7%	NPA	99.9%	99.7%-100%
	OPA	99.2%	98.8%-99.6%	OPA	99.9%	99.7%-100%
All Specimens Total	PPA	99.5%	98.8%-99.8%	PPA	99.3%	97.5%-99.9%
	NPA	99.0%	98.8%-99.2%	NPA	99.7%	99.6%-99.8%
	OPA	99.1%	98.9%-99.2%	OPA	99.7%	99.6%-99.8%

PPA = Positive Percent Agreement; NPA = Negative Percent Agreement; OPA = Overall Percent Agreement; SC = Self-Collected

Clinical Performance – Clinical Study

Extragenital (anorectal and oropharyngeal) specimens

The clinical utility and performance of **cobas**® CT/NG establish additionally in a multi-site, prospective study by comparing the results to an Infection Status (IS) that used a combination of commercially available CT/NG assay with anorectal or oropharyngeal swab samples. The subjects were enrolled from 8 geographically diverse clinic sites (STD, HIV, Family Planning, and STD Research) in the US. Specimens were tested for CT and NG using **cobas**® CT/NG and commercially available NAATs. All tests were run according to the respective manufacturers' Instructions For Use. The clinical performance of **cobas**® CT/NG was determined by comparing the results to the IS. A positive IS interpretation was derived when at least 2 of the 3 comparator reference assays were positive. The IS interpretations are further outlined in Table 29 below.

Table 29 Comparator Interpretation of Infection Status from "NAAT A", "NAAT B", and "NAAT C" results for each specimen type.

NAAT A	NAAT B	NAAT C	IS Interpretation
+	+	+	+
+	+	-	+
+	-	+	+
-	+	+	+
U	+	+	+
+	U	+	+
+	+	U	+
+	-	-	-
-	+	-	-
-	-	+	-
-	-	-	-
+	+	+	+
+	+	-	+
+	-	+	+
-	+	+	+
+	-	-	-
-	+	-	-
-	-	+	-
-	-	-	-
U	-	-	-
-	U	-	-
-	-	U	-
+	U	-	U
+ or -	U	U	U

IS = Infection status; + denotes Positive, - denotes Negative; U = uninterpretable test result.

Note: Uninterpretable test results occurred when retesting of invalid or equivocal results failed to yield a positive or negative result.

Results

A total of 2,439 subjects were consented to participate in this study, however, a total of 49 subjects were excluded, based on exclusion/inclusion criteria, that led to a total subject enrollment of 2,390. Of the 2,390 subjects contributing specimens, 2,365 anorectal and 2,382 oropharyngeal specimens were tested. Out of the 4,747 samples (2,365 rectal swabs and 2,382 oropharyngeal swabs), there were 4 final invalid results due to processing errors.

Chlamydia trachomatis: extragenital specimens infection status summary

Table 30 summarizes the results from evaluable subjects designated as CT positive or negative according to the IS algorithm for both Anorectal (AR) and Oropharyngeal (OP) specimens. Of the 2,365 contributing subjects for Rectum, 18 subjects had CT uninterpretable IS, 12 were excluded due to protocol deviations and the remaining 2,335 were evaluable. Similarly, of the 2,382 contributing subjects for Oropharyngeal, 23 subjects had CT uninterpretable IS, 11 were excluded due to protocol deviations, 3 were excluded due to failed test results and the remaining 2,345 were evaluable.

Table 30 *Chlamydia trachomatis*: summary of Infection status Interpretation for anorectal and oropharyngeal specimen types

Specimen Type[a]	NAAT A	NAAT B	NAAT C	IS[b] Interpretation	cobas® CT/NG	SS[c] Symp[d]	SS[c] Asymp[d]	SS[c] Unkn[d]	Total
AR	Inv	+	+	Positive	+	1	3	0	4
AR	-	+	+	Positive	-	0	3	0	3
AR	-	+	+	Positive	+	4	12	0	16
AR	+	-	+	Positive	-	0	1	0	1
AR	+	+	-	Positive	+	1	0	0	1
AR	+	+	+	Positive	-	2	1	0	3
AR	+	+	+	Positive	+	47	68	0	115
AR				**Total Positive**		**55**	**88**	**0**	**143**
AR	Inv	-	-	Negative	-	29	45	1	75
AR	-	NA	-	Negative	-	7	2	3	12
AR	-	NA	-	Negative	+	0	1	0	1
AR	-	Inv	-	Negative	-	3	3	0	6
AR	-	-	Inv	Negative	-	0	1	0	1
AR	-	-	Inv	Negative	+	1	0	0	1
AR	-	-	-	Negative	-	635	1,411	13	2,059
AR	-	-	-	Negative	+	5	2	0	7
AR	-	-	+	Negative	-	4	12	0	16
AR	-	-	+	Negative	+	0	6	0	6
AR	-	+	-	Negative	-	1	5	0	6
AR	-	+	-	Negative	+	1	1	0	2
AR				**Total Negative**		**686**	**1,489**	**17**	**2,192**
OP	Inv	+	+	Positive	+	0	1	0	1
OP	-	+	+	Positive	+	1	2	0	3
OP	+	+	-	Positive	+	0	1	0	1
OP	+	+	+	Positive	+	8	15	0	23
OP				**Total Positive**		**9**	**19**	**0**	**28**
OP	Inv	-	-	Negative	-	32	46	1	79
OP	-	NA	-	Negative	-	5	7	2	14
OP	-	Inv	-	Negative	-	1	6	0	7
OP	-	-	Inv	Negative	-	1	0	0	1
OP	-	-	-	Negative	-	679	1,486	14	2,179
OP	-	-	-	Negative	+	2	0	0	2
OP	-	-	+	Negative	-	13	16	0	29
OP	-	+	-	Negative	-	1	1	0	2
OP	-	+	-	Negative	+	0	2	0	2
OP	+	-	-	Negative	-	1	1	0	2
OP				**Total Negative**		**735**	**1,565**	**17**	**2,317**

[a]AR = anorectal; OP=oropharyngeal

[b]IS = infection status.

[c]SS = symptom status;

[d]Symp = symptomatic, Asymp = asymptomatic, Unkn = Unknown symptom status.

Note: NA = Not available, Inv = Invalid.

Note: Infection Status (IS) is determined for each specimen type. The IS of a sample will be established by the concordance results from at least 2 out of 3 comparator assays (NAAT A, NAAT B, NAAT C). If one of the comparator assays is Uninterpretable/Invalid/Failed, the two remaining assays must be concordant to define the IS as Positive (+) or Negative (-). Any other combination of Uninterpretable/Invalid/Failed and valid results are excluded from the analyses.

Note: Of the 2,365 contributing subjects for Rectum, 18 subjects had CT uninterpretable IS. Similarly, of the 2,382 contributing subjects for Oropharyngeal, 23 subjects had CT uninterpretable IS.

Note: Any IS interpretation that is Uninterpretable/Invalid/Failed/Protocol deviations are excluded from performance analyses.

Chlamydia trachomatis: performance results

The overall point estimate of **cobas®** CT/NG sensitivity for CT detection was 95.1% with a 95% CI of 90.2% to 97.6% for anorectal specimens and 100.0% with a 95% CI of 87.9% to 100% for oropharyngeal specimens. The sensitivity estimates were similar between asymptomatic and symptomatic subjects with overlapping two-sided 95% CIs (Table 31). The overall point estimate of **cobas®** CT/NG specificity for CT was 99.2% with a 95% CI of 98.8% to 99.5% for anorectal specimens and 99.8% with a 95% CI of 99.6% to 99.9% for oropharyngeal specimens. The specificity estimates were similar between asymptomatic and symptomatic subjects with overlapping two-sided 95% CIs (Table 31).

Table 31 *Chlamydia trachomatis*: overall clinical performance compared with Infection Status by sample type and symptom status

Sample Type[a]	Symptom Status[b]	Total (N)	SENS	95% Score CI	SPEC	95% Score CI	PREV (%)	PPV	NPV
AR	Symp	741	96.4% (53/55)	(87.7%, 99.0%)	99.0% (679/686)	(97.9%, 99.5%)	7.4	88.3% (53/60)	99.7% (679/681)
AR	Asymp	1,577	94.3% (83/88)	(87.4%, 97.5%)	99.3% (1479/1489)	(98.8%, 99.6%)	5.6	89.2% (83/93)	99.7% (1479/1484)
AR	Unknown	17	NE	NE	100.0% (17/17)	(81.6%, 100.0%)	0.0	NE	100.0% (17/17)
AR	Overall	2,335	95.1% (136/143)	(90.2%, 97.6%)	99.2% (2175/2192)	(98.8%, 99.5%)	6.1	88.9% (136/153)	99.7% (2175/2182)
OP	Symp	744	100.0% (9/9)	(70.1%, 100.0%)	99.7% (733/735)	(99.0%, 99.9%)	1.2	81.8% (9/11)	100.0% (733/733)
OP	Asymp	1,584	100.0% (19/19)	(83.2%, 100.0%)	99.9% (1563/1565)	(99.5%, 100.0%)	1.2	90.5% (19/21)	100.0% (1563/1563)
OP	Unknown	17	NE	NE	100.0% (17/17)	(81.6%, 100.0%)	0.0	NE	100.0% (17/17)
OP	Overall	2,345	100.0% (28/28)	(87.9%, 100.0%)	99.8% (2313/2317)	(99.6%, 99.9%)	1.2	87.5% (28/32)	100.0% (2313/2313)

[a]AR = anorectal, OP = oropharyngeal; [b]Symp = symptomatic, Asymp = asymptomatic.
Note: CI = confidence interval, PREV = prevalence; SENS = sensitivity; SPEC = specificity; PPV = positive predictive value; NPV = negative predictive value; NE =non-estimable.

Note: The predictive values shown above reflect performance specific to the clinical study population and may not be applicable to all individuals in the intended use population.

Neisseria gonorrhoeae: extragenital specimens infection status summary

Table 32 summarizes the results from evaluable subjects designated as NG positive or negative according to the IS algorithm for both Anorectal (AR) and Oropharyngeal (OP) specimens. Of the 2,365 contributing subjects for Rectum, 15 subjects had NG uninterpretable IS, 12 were excluded due to protocol deviations and the remaining 2,338 were evaluable. Similarly, of the 2,382 contributing subjects for Oropharyngeal, 19 subjects had NG uninterpretable IS, 11 were excluded due to protocol deviations, 3 were excluded due to failed test results and the remaining 2,349 were evaluable.

Table 32 *Neisseria gonorrhoeae*: Summary of Infection Status interpretation for anorectal and oropharyngeal specimen types

Specimen Type[a]	NAAT A	NAAT B	NAAT C	IS[b] Interpretation	cobas® CT/NG	SS[d] Symp[c]	SS[d] Asymp[c]	SS[d] Unkn[c]	Total
AR	Inv	+	+	Positive	+	2	0	0	2

Specimen Type[a]	NAAT A	NAAT B	NAAT C	IS[b] Interpretation	cobas® CT/NG	SS[d] Symp[c]	SS[d] Asymp[c]	SS[d] Unkn[c]	Total
AR	-	+	+	Positive	-	1	0	0	1
AR	-	+	+	Positive	+	3	4	0	7
AR	+	NA	+	Positive	+	0	1	0	1
AR	+	Inv	+	Positive	+	0	1	0	1
AR	+	-	+	Positive	+	0	1	0	1
AR	+	+	+	Positive	+	37	50	1	88
AR				**Total Positive**		**43**	**57**	**1**	**101**
AR	Inv	-	-	Negative	-	27	50	1	78
AR	Inv	-	-	Negative	+	1	0	0	1
AR	-	NA	-	Negative	-	7	2	3	12
AR	-	Inv	-	Negative	-	3	2	0	5
AR	-	-	-	Negative	-	646	1,461	12	2,119
AR	-	-	-	Negative	+	3	3	0	6
AR	-	-	+	Negative	-	5	0	0	5
AR	-	-	+	Negative	+	6	1	0	7
AR	-	+	-	Negative	-	0	2	0	2
AR	+	-	-	Negative	-	0	1	0	1
AR	+	-	-	Negative	+	1	0	0	1
AR				**Total Negative**		**699**	**1,522**	**16**	**2,237**
OP	Inv	+	+	Positive	+	1	1	0	2
OP	-	+	+	Positive	+	9	8	0	17
OP	+	NA	+	Positive	+	0	1	0	1
OP	+	Inv	+	Positive	+	0	1	0	1
OP	+	-	+	Positive	+	1	2	0	3
OP	+	+	-	Positive	+	0	1	0	1
OP	+	+	+	Positive	+	41	30	0	71
OP				**Total Positive**		**52**	**44**	**0**	**96**
OP	Inv	-	-	Negative	-	30	53	1	84
OP	-	NA	-	Negative	-	5	6	2	13
OP	-	Inv	-	Negative	-	0	5	0	5
OP	-	-	Inv	Negative	-	2	0	0	2
OP	-	-	Inv	Negative	+	1	0	0	1
OP	-	-	-	Negative	-	631	1,452	13	2,096
OP	-	-	-	Negative	+	6	7	1	14
OP	-	-	+	Negative	-	4	3	0	7
OP	-	-	+	Negative	+	4	4	0	8
OP	-	+	-	Negative	-	5	15	0	20
OP	-	+	-	Negative	+	0	1	0	1
OP	+	-	-	Negative	-	1	0	0	1
OP	+	-	-	Negative	+	0	1	0	1
OP				**Total Negative**		**689**	**1,547**	**17**	**2,253**

[a]AR = anorectal; OP = oropharyngeal.

[b] IS= infection status

[c]Symp = symptomatic, Asymp = asymptomatic, Unkn = Unknown symptom status.

[d]SS=symptom status
Note: NA = Not available, Inv = Invalid.
Note: Infection status (IS) is determined for each specimen type. The IS of a sample will be established by the concordance results from at least 2 out of 3 comparator assays (NAAT A, NAAT B, NAAT C).
If one of the comparator assays is Uninterpretable/Invalid/Failed, the two remaining assays must be concordant to define the IS as Positive (+) or Negative (-).

Note: Of the 2,365 contributing subjects for Rectum, 15 subjects had NG uninterpretable IS. Similarly, of the 2,382 contributing subjects for Oropharyngeal, 19 subjects had NG uninterpretable IS.

Note: Any IS interpretation that is Uninterpretable/Invalid/Failed/Protocol deviations are excluded from performance analyses.

Neisseria gonorrhoeae: performance results

The overall point estimate of **cobas**® CT/NG sensitivity for NG detection was 99.0%, with a 95% CI of 94.6% to 99.8% for anorectal specimens and 100.0% with a 95% CI of 96.2% to 100% for oropharyngeal specimens. The sensitivity estimates were 97.7% (42/43) and 100% (57/57) in anorectal specimens for symptomatic and asymptomatic subjects respectively (Table 29). The sensitivity estimates were both 100% in oropharyngeal specimens for symptomatic (52/52) and asymptomatic (44/44) subjects.

The overall point estimate of **cobas**® CT/NG specificity for NG was 99.3% with a 95% CI of 98.9% to 99.6% for anorectal specimens and 98.9% with a 95% CI of 98.4% to 99.2% for oropharyngeal specimens. The specificity estimates were similar between asymptomatic and symptomatic subjects (Table 33).

Table 33 *Neisseria gonorrhoeae*: overall clinical performance compared with infection status by sample type and symptom status

Sample Type[a]	Symptom Status[b]	Total (N)	SENS	95% Score CI	SPEC	95% Score CI	PREV (%)	PPV	NPV
AR	Symp	742	97.7% (42/43)	(87.9%, 99.6%)	98.4% (688/699)	(97.2%, 99.1%)	5.8	79.2% (42/53)	99.9% (688/689)
AR	Asymp	1,579	100.0% (57/57)	(93.7%, 100.0%)	99.7% (1518/1522)	(99.3%, 99.9%)	3.6	93.4% (57/61)	100.0% (1518/1518)
AR	Unknown	17	100.0% (1/1)	(20.7%, 100.0%)	100.0% (16/16)	(80.6%, 100.0%)	5.9	100.0% (1/1)	100.0% (16/16)
AR	Overall	2,338	99.0% (100/101)	(94.6%, 99.8%)	99.3% (2222/2237)	(98.9%, 99.6%)	4.3	87.0% (100/115)	100.0% (2222/2223)
OP	Symp	741	100.0% (52/52)	(93.1%, 100.0%)	98.4% (678/689)	(97.2%, 99.1%)	7.0	82.5% (52/63)	100.0% (678/678)
OP	Asymp	1,591	100.0% (44/44)	(92.0%, 100.0%)	99.2% (1534/1547)	(98.6%, 99.5%)	2.8	77.2% (44/57)	100.0% (1534/1534)
OP	Unknown	17	NE	NE	94.1% (16/17)	(73.0%, 99.0%)	0.0	0.0% (0/1)	100.0% (16/16)
OP	Overall	2,349	100.0% (96/96)	(96.2%, 100.0%)	98.9% (2228/2253)	(98.4%, 99.2%)	4.1	79.3% (96/121)	100.0% (2228/2228)

Expected values for extragenital specimens

Prevalence

Positive and negative predictive values for hypothetical prevalence rates

The hypothetical PPVs and NPVs of **cobas**® CT/NG derived from CT disease prevalence of 1% to 50% are shown in Table 34 for anorectal specimens and Table 35 for oropharyngeal specimens.

Table 34 Positive Predictive Value and Negative Predictive Value for hypothetical *Chlamydia trachomatis* prevalence for anorectal specimens

Hypothetical Prevalence (%)	Sensitivity (%) [a]	Specificity (%) [a]	PPV (%)	NPV (%)
1	95.1	99.2	55.33	99.95
3	95.1	99.2	79.14	99.85
5	95.1	99.2	86.59	99.74
10	95.1	99.2	93.16	99.45
15	95.1	99.2	95.58	99.14
20	95.1	99.2	96.84	98.78
30	95.1	99.2	98.13	97.93
50	95.1	99.2	99.19	95.30

[a]The overall sensitivity and specificity were estimated by comparing the test results with **cobas**® CT/NG to the Infection Status across all genders for CT anorectal.

Note: PPV = Positive predictive value, NPV = Negative predictive value.

Table 35 Positive Predictive Value and Negative Predictive Value for hypothetical *Chlamydia trachomatis* prevalence for oropharyngeal specimens

Hypothetical Prevalence (%)	Sensitivity (%) [a]	Specificity (%) [a]	PPV (%)	NPV (%)
1	100.0	99.8	85.41	100.0
3	100.0	99.8	94.71	100.0
5	100.0	99.8	96.82	100.0
10	100.0	99.8	98.47	100.0
15	100.0	99.8	99.03	100.0
20	100.0	99.8	99.31	100.0
30	100.0	99.8	99.60	100.0
50	100.0	99.8	99.83	100.0

[a]The overall sensitivity and specificity were estimated by comparing the test results with **cobas**® CT/NG to the Infection Status across all genders for CT oropharyngeal.

Note: PPV = Positive predictive value, NPV = Negative predictive value.

The hypothetical PPVs and NPVs of **cobas**® CT/NG derived from NG disease prevalence of 1% to 50% are shown in Table 36 for anorectal specimens and Table 37 for oropharyngeal specimens.

Table 36 Positive Predictive Value and Negative Predictive Value for hypothetical *Neisseria gonorrhoeae* prevalence for anorectal specimens

Hypothetical Prevalence (%)	Sensitivity (%) [a]	Specificity (%) [a]	PPV (%)	NPV (%)
1	99.0	99.3	59.86	99.99
3	99.0	99.3	82.04	99.97
5	99.0	99.3	88.60	99.95
10	99.0	99.3	94.26	99.89
15	99.0	99.3	96.30	99.82
20	99.0	99.3	97.36	99.75
30	99.0	99.3	98.44	99.57
50	99.0	99.3	99.33	99.01

[a] The overall sensitivity and specificity were estimated by comparing the test results with **cobas**® CT/NG to the Infection Status across all genders for NG Anorectal.
Note: PPV = Positive predictive value, NPV = Negative predictive value.

Table 37 Positive Predictive Value and Negative Predictive Value for hypothetical *Neisseria gonorrhoeae* prevalence for oropharyngeal specimens

Hypothetical Prevalence (%)	Sensitivity (%) [a]	Specificity (%) [a]	PPV (%)	NPV (%)
1	100.0	98.9	47.65	100.0
3	100.0	98.9	73.60	100.0
5	100.0	98.9	82.59	100.0
10	100.0	98.9	90.92	100.0
15	100.0	98.9	94.08	100.0
20	100.0	98.9	95.75	100.0
30	100.0	98.9	97.48	100.0
50	100.0	98.9	98.90	100.0

Table 38 and Table 39 present the positivity rate as determined by the **cobas**® CT/NG test for *C. trachomatis* and *N. gonorrhoeae* respectively. Overall, the observed positivity rate for *C. trachomatis* during the clinical study, when testing with the **cobas**® CT/NG Test, was 1.4% and 6.6% for oropharyngeal and anorectal swabs, respectively; similarly, the observed positivity rate for *N. gonorrhoeae*, as determined by the **cobas**® CT/NG test, was 5.2% and 4.9% for oropharyngeal and anorectal swabs, respectively. The positivity rate for each site and overall is shown below.

Table 38 CT Positivity rate as determined by **cobas®** CT/NG for oropharyngeal and anorectal specimen types

Collection Site ID	OP[a] Number of Samples Tested with cobas® valid results (N)	OP[a] Number of Positive Results by cobas® CT/NG (n)	OP[a] Positivity Rate (%)[c] (n/N)	AR[b] Number of Samples Tested (N)	AR[b] Number of Positive Results by cobas® CT/NG (n)	AR[b] Positivity Rate (%)[c] (n/N)
10	170	2	1.2%	170	6	3.5%
11	90	1	1.1%	89	4	4.5%
12	388	6	1.5%	385	49	12.7%
13	171	1	0.6%	170	3	1.8%
14	259	2	0.8%	256	18	7.0%
15	433	10	2.3%	426	36	8.5%
16	394	6	1.5%	395	19	4.8%
17	457	5	1.1%	456	21	4.6%
Total	2362	33	1.4%	2347	156	6.6%

[a] OP= Oropharyngeal / Throat specimen type.

[b] AR= Anorectal specimen type.

[c] Positivity Rate (%) = (Number of valid Positive **cobas** results/Total number of **cobas** valid results) x100.

Table 39 NG Positivity rate as determined by **cobas**® CT/NG for oropharyngeal and anorectal specimen types

Collection Site ID	OP[a] Number of Samples Tested with cobas® valid results (N)	OP[a] Number of Positive Results by cobas® CT/NG (n)	OP[a] Positivity Rate (%)[c] (n/N)	AR[b] Number of Samples Tested (N)	AR[b] Number of Positive Results by cobas® CT/NG (n)	AR[b] Positivity Rate (%)[c] (n/N)
10	170	6	3.5%	170	4	2.4%
11	90	7	7.8%	89	4	4.5%
12	388	51	13.1%	385	48	12.5%
13	171	0	0 %	170	3	1.8%
14	259	21	8.1%	256	18	7.0%
15	433	14	3.2%	426	15	3.5%
16	394	10	2.5%	395	8	2.0%
17	457	14	3.1%	456	15	3.3%
Total	2362	123	5.2%	2347	115	4.9%

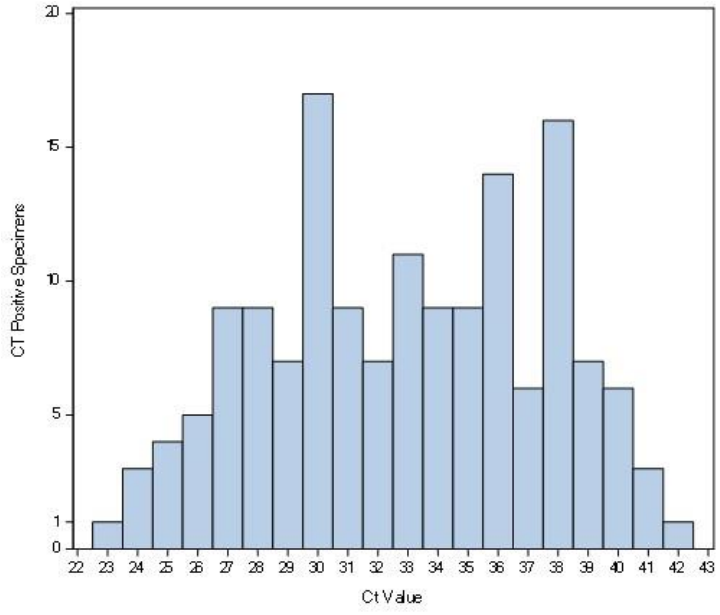
[a] OP= Oropharyngeal/Throat specimen type.

[b] AR= Anorectal / Rectum specimen type.

[c] Positivity Rate (%) = (Number of valid Positive **cobas** results/Total number of **cobas** valid results) x100.

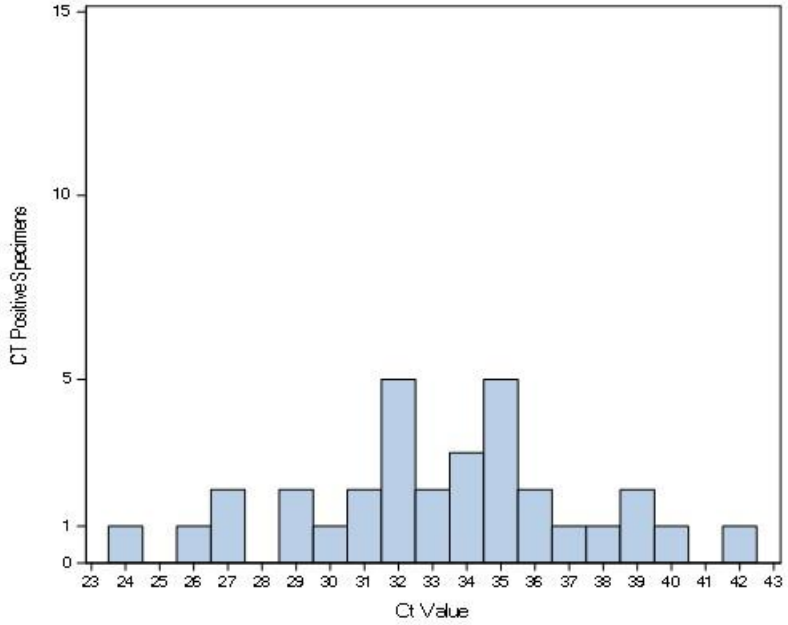
For extragenital specimens, the frequency distribution of **cobas**® CT/NG–positive results for CT anorectal, CT oropharyngeal, NG anorectal, and NG oropharyngeal are shown in Figure 7, Figure 8, Figure 9 and Figure 10 respectively.

Figure 7 Distribution of cycle threshold values for **cobas**® CT/NG (*Chlamydia trachomatis* anorectal specimens)



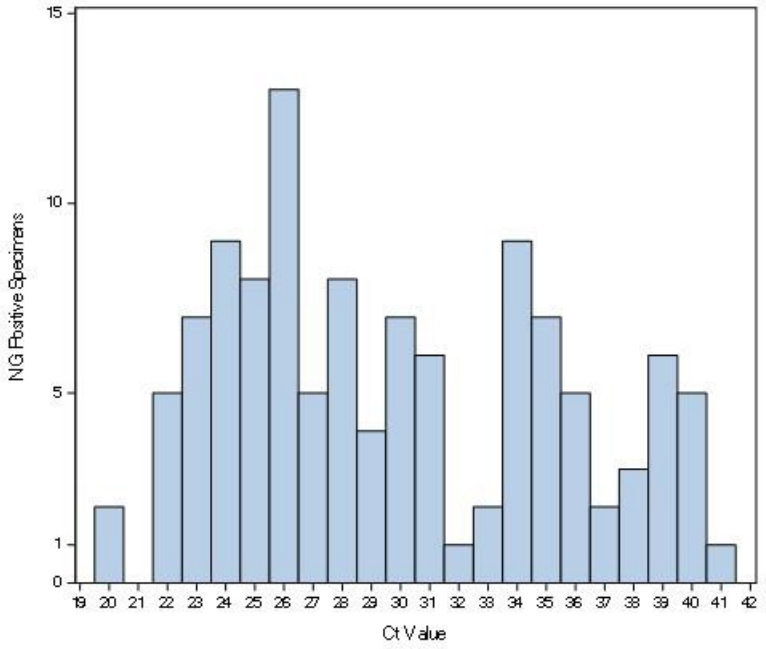
Ct = cycle threshold; CT = Chlamydia trachomatis.

Figure 8 Distribution of cycle threshold values for **cobas**® CT/NG (*Chlamydia trachomatis* oropharyngeal specimens)



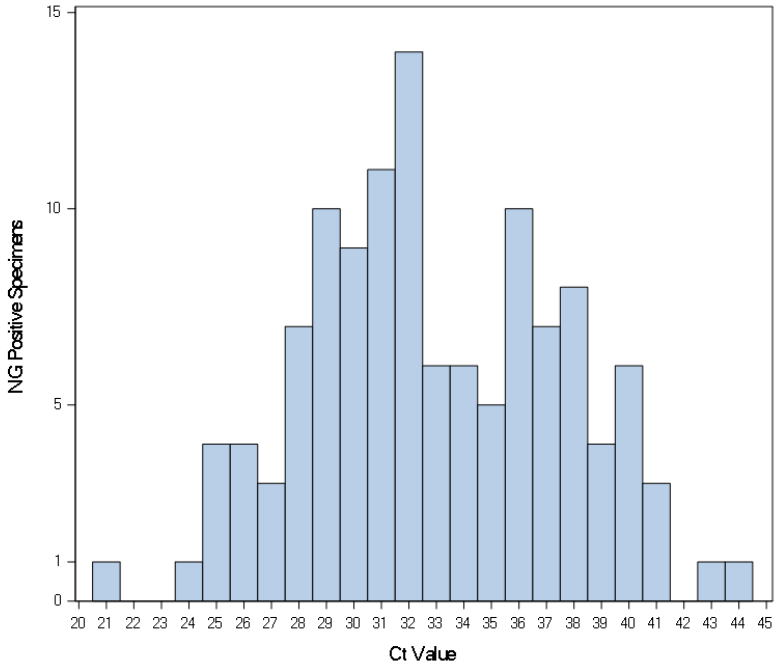
Ct = cycle threshold; CT = Chlamydia trachomatis.

Figure 9 Distribution of cycle threshold values for **cobas**® CT/NG (*Neisseria gonorrhoeae* anorectal specimens)



Ct = cycle threshold; NG = Neisseria gonorrhoeae.

Figure 10 Distribution of cycle threshold values for **cobas**® CT/NG (*Neisseria gonorrhoeae* oropharyngeal specimens)



Ct = cycle threshold; NG = Neisseria gonorrhoeae.

System equivalency

System equivalency of the **cobas**® 5800, **cobas**® 6800 and **cobas**® 8800 Systems was demonstrated via performance studies. The data presented in this Instructions for Use support equivalent performance for all systems.

Additional information

Key assay features

Sample types	• Endocervical swab collected in **cobas**® PCR Media • Vaginal swab collected in **cobas**® PCR Media • Self-collected Vaginal swab collected in **cobas**® PCR Media • Oropharyngeal swab collected in **cobas**® PCR Media • Anorectal swab collected in **cobas**® PCR Media • Male and female urine stabilized in **cobas**® PCR Media • Cervical specimen collected in PreservCyt® Solution
Amount of sample required/processed	• ≥1000 µL required in sample tube for all swab samples, instrument processes 400 µL • ≥1000 µL required in sample tube for PreservCyt® samples, instrument processes 400 µL • ≥1200 µL required in sample tube for urine samples, instrument processes 850 µL • On **cobas**® 5800 System, ≥3000 µL required in sample tube for PreservCyt® samples in primary tubes, instrument processes 400 µL
Test duration	• < 3.5 hours to first result

Symbols

The following symbols are used in labeling for Roche PCR diagnostic products.

Table 40 Symbols used in labeling for Roche PCR diagnostics products

Age/DOB Age or Date of Birth	Device not for near-patient testing	QS IU/PCR QS IU per PCR reaction, use the QS International Units (IU) per PCR reaction in calculation of the results.
SW Ancillary Software	Device not for self-testing	
Assigned Range [copies/mL] Assigned Range (copies/mL)	Distributor *(Note: The applicable country/region may be designated beneath the symbol)*	SN Serial number
Assigned Range [IU/mL] Assigned Range (IU/mL)	Do not re-use	Site Site
EC REP Authorized representative in the European Community	Female	Procedure Standard Standard Procedure
BARCODE Barcode Data Sheet	For IVD performance evaluation only	STERILE EO Sterilized using ethylene oxide
LOT Batch code	GTIN Global Trade Item Number	Store in dark
Biological risks	Importer	Temperature limit
REF Catalogue number	IVD In vitro diagnostic medical device	TDF Test Definition File
CE CE marking of conformity; this device is in conformity with the applicable requirements for CE marking of an in vitro diagnostic medical device	LLR Lower Limit of Assigned Range	This way up
	Male	Procedure UltraSensitive Ultrasensitive Procedure
Collect Date Collect date	Manufacturer	UDI Unique Device Identifier
Consult instructions for use	CONTROL − Negative control	ULR Upper Limit of Assigned Range
Σ Contains sufficient for <*n*> tests	NON STERILE Non-sterile	Urine Fill Line Urine Fill Line
CONTENT Content of kit	Patient Name	Rx Only US Only: Federal law restricts this device to sale by or on the order of a physician.
CONTROL Control	Patient number	Use-by date
Date of manufacture	Peel here	
Device for near-patient testing	CONTROL + Positive control	
Device for self-testing	QS copies / PCR QS copies per PCR reaction, use the QS copies per PCR reaction in calculation of the results.	

Technical support

For technical support (assistance) please reach out to your local affiliate:
https://www.roche.com/about/business/roche_worldwide.htm

Manufacturer

Table 41 Manufacturer

Manufactured in the United States

Roche Molecular Systems, Inc.
1080 US Highway 202 South
Branchburg, NJ 08876 USA
www.roche.com

Made in USA

Trademarks and patents

See https://diagnostics.roche.com/us/en/about-us/patents

Copyright

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Document revision

Document Revision Information	
Doc Rev. 1.0 10/2023	First Publishing