

REF		CONTENT		Analyzer(s) on which <b>cobas c</b> pack(s) can be used
05589061190	05589061500	Bilirubin Direct Gen.2 350 test	System-ID 07 7479 0	<b>cobas c 311</b> , <b>cobas c 501/502</b> , COBAS INTEGRA 400 plus

Materials required (but not provided):

		<b>cobas c 311</b> , <b>cobas c 501/502</b>	COBAS INTEGRA 400 plus
10759350190	Calibrator f.a.s. (12 x 3 mL)	Code 401	System-ID 07 3718 6
12149435122	Precinorm U plus (10 x 3 mL)	Code 300	System-ID 07 7999 7
12149443122	Precipath U plus (10 x 3 mL)	Code 301	System-ID 07 8000 6
05117003190	PreciControl ClinChem Multi 1 (20 x 5 mL)	Code 391	System-ID 07 7469 3
05947626190	PreciControl ClinChem Multi 1 (4 x 5 mL)	Code 391	System-ID 07 7469 3
05117216190	PreciControl ClinChem Multi 2 (20 x 5 mL)	Code 392	System-ID 07 7470 7
05947774190	PreciControl ClinChem Multi 2 (4 x 5 mL)	Code 392	System-ID 07 7470 7
10158046122	Precibil (4 x 2 mL)	Code 306	System-ID 07 6604 6
20756350322	NaCl Diluent 9 % (6 x 22 mL)	n.a.	System-ID 07 5635 0

## English

### Intended use

In vitro test for the quantitative determination of direct bilirubin in human serum and plasma on **cobas c** and COBAS INTEGRA systems.

### Summary

Measurements of direct bilirubin, performed with this assay in human serum and plasma of adults and neonates, are used for the diagnosis of hyperbilirubinemia (such as observed with abnormal destruction of red blood cells, liver diseases, and metabolic disorders, including hepatitis and gallbladder block), and in newborn screening for severe hyperbilirubinemia.

Bilirubin is formed in the reticuloendothelial system during the degradation of aged erythrocytes. The heme portion from hemoglobin and from other heme-containing proteins is removed, metabolized to bilirubin, and transported as a complex with serum albumin to the liver. In the liver, bilirubin is conjugated with glucuronic acid for solubilization and subsequent transport through the bile duct and elimination via the digestive tract. Diseases or conditions which, through hemolytic processes, produce bilirubin faster than the liver can metabolize it, cause the levels of unconjugated (indirect) bilirubin to increase in the circulation. Liver immaturity and several other diseases in which the bilirubin conjugation mechanism is impaired cause similar elevations of circulating unconjugated bilirubin. Bile duct obstruction or damage to hepatocellular structure causes increases in the levels of both conjugated (direct) and unconjugated (indirect) bilirubin in the circulation.<sup>1,2,3,4</sup>

In newborns, several mechanisms lead to an increased bilirubin load, such as increased turnover in fetal red blood cells, reduced bilirubin clearance, and increased enterohepatic circulation of bilirubin. Screening neonates for severe hyperbilirubinemia, especially in newborns with infant jaundice, has been proposed to help preventing chronic bilirubin encephalopathy.<sup>5,6</sup>

### Test principle

Diazo method.<sup>7</sup>

Conjugated bilirubin and  $\delta$ -bilirubin (direct bilirubin) react directly with 3,5-Dichlorophenyl diazonium salt in acid buffer to form the red-colored azobilirubin.

bilirubin + 3,5 DPD  $\longrightarrow$  azobilirubin

The color intensity of the red azo dye formed is directly proportional to the direct (conjugated) bilirubin concentration and can be determined photometrically.

Remark: Under the influence of blue light, e.g. during phototherapy of newborn children, unconjugated bilirubin is partly transformed into a water-soluble isomer called photobilirubin, a substrate for direct bilirubin tests. This fraction is detected by BILD2 and may lead to above-normal results in healthy children.

### Precautions and warnings

For in vitro diagnostic use for health care professionals. Exercise the normal precautions required for handling all laboratory reagents.

Infectious or microbial waste:

Warning: handle waste as potentially biohazardous material. Dispose of waste according to accepted laboratory instructions and procedures.

Environmental hazards:

Apply all relevant local disposal regulations to determine the safe disposal.

Safety data sheet available for professional user on request.

### Reagent handling

Ready for use

### Specimen collection and preparation

For specimen collection and preparation only use suitable tubes or collection containers.

Only the specimens listed below were tested and found acceptable.

Serum: Collect serum using standard sample tubes.

Plasma: Li-heparin, K<sub>2</sub>-, K<sub>3</sub>-EDTA.

Protect specimens from exposure to light.

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube manufacturer.

Centrifuge samples containing precipitates before performing the assay.

See the limitations and interferences section for details about possible sample interferences.

Stability:<sup>a,8,9</sup>

2 days at 15-25 °C

7 days at 2-8 °C

6 months at (-15)-(-25) °C

Freeze only once.

### Materials provided

See "Reagents – working solutions" section for reagents.

### Materials required (but not provided)

- See "Order information" section
- General laboratory equipment

### Assay

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.

The performance of applications not validated by Roche is not warranted and must be defined by the user.

**Calculation**

The systems automatically calculate the analyte concentration of each sample.

Conversion factors:  $\mu\text{mol/L} \times 0.0585 = \text{mg/dL}$   
 $\text{mg/dL} \times 10 = \mu\text{mol/L}$   
 $\text{mg/dL} \times 17.1 = \mu\text{mol/L}$

**Expected values***Jendrassik-Grof method*

Direct bilirubin  $\leq 5 \mu\text{mol/L}$  ( $\leq 0.30 \text{ mg/dL}$ )<sup>1</sup>

An upper limit of  $10 \mu\text{mol/L}$  direct bilirubin for neonates has been cited in the literature, although this has not been confirmed by internal data.<sup>10</sup>

*Doumas method*

Direct bilirubin  $\leq 3.4 \mu\text{mol/L}$  ( $\leq 0.20 \text{ mg/dL}$ )<sup>11</sup>

An upper limit of  $10 \mu\text{mol/L}$  direct bilirubin for neonates has been cited in the literature, although this has not been confirmed by internal data.<sup>10</sup>

Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

**cobas c systems****System information***Jendrassik-Grof method*

For **cobas c** 311/501 analyzers:

**BILD2:** ACN 734

For **cobas c** 502 analyzer:

**BILD2:** ACN 8734

*Doumas method*

For **cobas c** 311/501 analyzers:

**DBIL2:** ACN 735

For **cobas c** 502 analyzer:

**DBIL2:** ACN 8735

**Reagents - working solutions**

**R1** Phosphoric acid: 85 mmol/L; HEDTA: 4.0 mmol/L; NaCl: 50 mmol/L; detergent; pH 1.9

**R2** 3,5 Dichlorophenyl diazonium: 1.5 mmol/L; pH 1.3

R1 is in position B and R2 is in position C.

**Storage and stability**

Shelf life at 2-8 °C: See expiration date on **cobas c** pack label.

On-board in use and refrigerated on the analyzer: 6 weeks

**Application for serum and plasma****cobas c 311 test definition**

Assay type	2-Point End	
Reaction time / Assay points	10 / 6-8	
Wavelength (sub/main)	800/546 nm	
Reaction direction	Increase	
Units	$\mu\text{mol/L}$ (mg/dL, mg/L)	
Reagent pipetting		Diluent (NaCl)
R1	120 $\mu\text{L}$	–
R2	24 $\mu\text{L}$	–
<i>Sample volumes</i>	<i>Sample</i>	<i>Sample dilution</i>

		Sample	Diluent (H <sub>2</sub> O)
Normal	6.7 $\mu\text{L}$	–	–
Decreased	3.4 $\mu\text{L}$	–	–
Increased	6.7 $\mu\text{L}$	–	–

**cobas c 501/502 test definition**

Assay type	2-Point End		
Reaction time / Assay points	10 / 10-13		
Wavelength (sub/main)	800/546 nm		
Reaction direction	Increase		
Units	$\mu\text{mol/L}$ (mg/dL, mg/L)		
Reagent pipetting		Diluent (NaCl)	
R1	120 $\mu\text{L}$	–	
R2	24 $\mu\text{L}$	–	

<i>Sample volumes</i>	<i>Sample</i>	<i>Sample dilution</i>	
		Sample	Diluent (H <sub>2</sub> O)
Normal	6.7 $\mu\text{L}$	–	–
Decreased	3.4 $\mu\text{L}$	–	–
Increased	6.7 $\mu\text{L}$	–	–

**Calibration**

Calibrator	S1: H <sub>2</sub> O S2: Calibrator f.a.s.
Calibration mode	Linear regression
Calibration frequency	2-point calibration - after reagent lot change - as required following quality control procedures

Traceability: This method has been standardized against the manual test performance using the Jendrassik-Grof or Doumas method.<sup>12,13</sup>

**Quality control**

For quality control, use control materials as listed in the "Order information" section.

In addition, other suitable control material can be used.

The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

Follow the applicable government regulations and local guidelines for quality control.

**Limitations - interference**

Criterion: Recovery within  $\pm 10 \%$  of initial values at a direct bilirubin concentration of  $34.2 \mu\text{mol/L}$  ( $2.0 \text{ mg/dL}$ ).

Hemolysis:<sup>14</sup> No significant interference up to an H index of 25 (approximate hemoglobin concentration:  $15.5 \mu\text{mol/L}$  or  $25 \text{ mg/dL}$ ).

Lipemia (Intralipid):<sup>14</sup> No significant interference up to an L index of 750. There is poor correlation between the L index (corresponds to turbidity) and triglycerides concentration.

Drugs: No interference was found at therapeutic concentrations using common drug panels.<sup>15,16</sup>

Exception: Phenylbutazone causes artificially low bilirubin results.

Samples containing indocyanine green must not be measured.

In very rare cases, gammopathy, in particular type IgM (Waldenström's macroglobulinemia), may cause unreliable results.<sup>17</sup>

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

In certain cases specimens may give a direct bilirubin result slightly greater than the total bilirubin result. This is observed in patient samples when nearly all the reacting bilirubin is in the direct form. In such cases the result for the total bilirubin should be reported for both direct bilirubin and total bilirubin values.

**ACTION REQUIRED**

**Special Wash Programming:** The use of special wash steps is mandatory when certain test combinations are run together on **cobas c** systems. The latest version of the carry-over evasion list can be found with the NaOHD-SMS-SmpCln1+2-SCCS Method Sheets. For further instructions refer to the operator's manual. **cobas c** 502 analyzer: All special wash programming necessary for avoiding carry-over is available via the **cobas** link, manual input is required in certain cases.

**Where required, special wash/carry-over evasion programming must be implemented prior to reporting results with this test.**

**Limits and ranges****Measuring range***Jendrassik-Grof method*

1.5-291 µmol/L (0.09-17 mg/dL)

*Doumas method*

1.4-236 µmol/L (0.08-13.8 mg/dL)

Determine samples having higher concentrations via the rerun function. Dilution of samples via the rerun function is a 1:2 dilution. Results from samples diluted using the rerun function are automatically multiplied by a factor of 2.

**Lower limits of measurement***Jendrassik-Grof method**Limit of Blank, Limit of Detection and Limit of Quantitation*

Limit of Blank = 1.0 µmol/L (0.06 mg/dL)

Limit of Detection = 1.5 µmol/L (0.09 mg/dL)

Limit of Quantitation = 3.0 µmol/L (0.18 mg/dL)

The Limit of Blank, the Limit of Detection and the Limit of Quantitation were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A requirements.

The Limit of Blank is the 95<sup>th</sup> percentile value from  $n \geq 60$  measurements of analyte-free samples over several independent series. The Limit of Blank corresponds to the concentration below which analyte-free samples are found with a probability of 95 %.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low concentration samples.

The Limit of Detection corresponds to the lowest analyte concentration which can be detected (value above the Limit of Blank with a probability of 95 %).

The Limit of Quantitation is the lowest analyte concentration that can be reproducibly measured with a total error of 30 %. It has been determined using low concentration bilirubin samples.

**Lower limits of measurement***Doumas method**Limit of Blank, Limit of Detection and Limit of Quantitation*

Limit of Blank = 0.8 µmol/L (0.05 mg/dL)

Limit of Detection = 1.2 µmol/L (0.07 mg/dL)

Limit of Quantitation = 1.4 µmol/L (0.08 mg/dL)

The Limit of Blank, the Limit of Detection and the Limit of Quantitation were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A2 requirements.

The Limit of Blank is the 95<sup>th</sup> percentile value from  $n \geq 60$  measurements of analyte-free samples over several independent series. The Limit of Blank corresponds to the concentration below which analyte-free samples are found with a probability of 95 %.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low concentration samples.

The Limit of Detection corresponds to the lowest analyte concentration which can be detected (value above the Limit of Blank with a probability of 95 %).

The Limit of Quantitation is the lowest analyte concentration that can be reproducibly measured with a precision of 20 % CV. It has been determined using low concentration direct bilirubin samples.

**Specific performance data**

Representative performance data on the analyzers are given below. Results obtained in individual laboratories may differ.

**Precision***Jendrassik-Grof method*

Precision was determined using human samples and controls in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP5 requirements with repeatability ( $n = 21$ ) and intermediate precision (4 aliquots per run, 1 run per day, 21 days). The following results were obtained on the **cobas c** 501 analyzer:

Repeatability	Mean	SD	CV
	µmol/L (mg/dL)	µmol/L (mg/dL)	%
Precinorm U	16.2 (0.948)	0.1 (0.006)	0.6
Precipath U	42.0 (2.46)	0.1 (0.01)	0.3
Human serum 1	2.5 (0.146)	0.1 (0.006)	2.9
Human serum 2	174 (10.2)	1 (0.1)	0.3
Human serum 3	280 (16.4)	1 (0.1)	0.3
Intermediate precision	Mean	SD	CV
	µmol/L (mg/dL)	µmol/L (mg/dL)	%
Precinorm U	14.9 (0.872)	0.4 (0.023)	2.6
Precipath U	38.8 (2.27)	0.5 (0.03)	1.4
Human serum 1	1.8 (0.105)	0.2 (0.018)	10
Human serum 2	179 (10.5)	2.6 (0.15)	1.5
Human serum 3	260 (15.2)	4.0 (0.23)	1.5

*Doumas method*

Repeatability and intermediate precision were determined using human samples and controls in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP5 requirements (2 aliquots per run, 2 runs per day, 21 days). The following results were obtained on the **cobas c** 501 analyzer:

Repeatability	Mean	SD	CV
	µmol/L (mg/dL)	µmol/L (mg/dL)	%
Precinorm U	12.0 (0.7)	0.2 (0.01)	1.7
Precipath U	31.4 (1.8)	0.2 (0.01)	0.5
Human serum 1	1.5 (0.09)	0.1 (0.01)	4.4
Human serum 2	145 (8.5)	0.7 (0.04)	0.5
Human serum 3	211 (12.3)	0.8 (0.05)	0.4
Intermediate precision	Mean	SD	CV
	µmol/L (mg/dL)	µmol/L (mg/dL)	%
Precinorm U	12.0 (0.7)	0.3 (0.02)	2.6
Precipath U	31.4 (1.8)	0.4 (0.02)	1.4
Human serum 1	1.5 (0.09)	0.2 (0.01)	10
Human serum 2	145 (8.5)	2.1 (0.12)	1.5
Human serum 3	211 (12.3)	3.2 (0.19)	1.5

The data obtained on **cobas c** 501 analyzer(s) are representative for **cobas c** 311 analyzer(s).

**Method comparison***Jendrassik-Grof method*

# BILD2

## Bilirubin Direct Gen.2

Bilirubin values for human serum and plasma samples obtained with the Roche BILD2 reagent on a **cobas c 501** analyzer (y) were compared to those determined with the previous Roche DBIL reagent on a Roche/Hitachi MODULAR P analyzer (x).

Sample size (n) = 65

Passing/Bablok <sup>18</sup>	Linear regression
$y = 1.010x + 1.17 \mu\text{mol/L}$	$y = 0.998x + 2.38 \mu\text{mol/L}$
$\tau = 0.950$	$r = 0.997$

The sample concentrations were between 2.4 and 161  $\mu\text{mol/L}$  (0.14 and 9.42 mg/dL).

The data obtained on **cobas c 501** analyzer(s) are representative for **cobas c 311** analyzer(s).

### Doumas method

Bilirubin values for human serum and plasma samples obtained using the Roche Direct Bilirubin Gen.2 reagent on a **cobas c 501** analyzer (x) were compared with those determined with the Roche BILD2 reagent on a COBAS INTEGRA 800 analyzer (y).

Sample size (n) = 75

Passing/Bablok <sup>18</sup>	Linear regression
$y = 0.995x + 0.339 \mu\text{mol/L}$	$y = 0.993x - 0.158 \mu\text{mol/L}$
$\tau = 0.962$	$r = 0.999$

The sample concentrations were between 2.2 and 200  $\mu\text{mol/L}$  (0.13 and 11.7 mg/dL).

The data obtained on **cobas c 501** analyzer(s) are representative for **cobas c 311** analyzer(s).

## COBAS INTEGRA systems

### System information

#### Jendrassik-Grof method

**BILD2:** Test ID 0-734

#### Doumas method

**DBIL2:** Test ID 0-735

### Reagents - working solutions

**R1** Phosphoric acid: 85 mmol/L; HEDTA: 4.0 mmol/L; NaCl: 50 mmol/L; detergent; pH 1.9

**SR** 3,5-Dichlorophenyl diazonium: 1.5 mmol/L; pH 1.3

R1 is in position B and SR is in position C.

### Storage and stability

Shelf life at 2-8 °C	See expiration date on <b>cobas c</b> pack label
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On-board in use at 10-15 °C	6 weeks
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### Application for serum and plasma

### Test definition

Measuring mode	Absorbance
Abs. calculation mode	Endpoint
Reaction mode	R1-S-SR
Reaction direction	Increase
Wavelength A/B	552/659 nm
Calc. first/last	MP 33-36
Unit	$\mu\text{mol/L}$

### Pipetting parameters

		Diluent (H <sub>2</sub> O)
R1	120 $\mu\text{L}$	–
SR	24 $\mu\text{L}$	–

Sample	7 $\mu\text{L}$	2 $\mu\text{L}$
Total volume	153 $\mu\text{L}$	

### Calibration

Calibrator	Calibrator f.a.s. Use deionized water as zero calibrator.
Calibration mode	Linear regression
Calibration replicate	Duplicate recommended
Calibration interval	Each lot and as required following quality control procedures

Traceability: This method has been standardized against the manual test performance using the Jendrassik Grof or Doumas method.<sup>12,13</sup>

### Quality control

Reference range	Precinorm U plus or PreciControl ClinChem Multi 1
Pathological range	Precipath U plus, PreciControl ClinChem Multi 2 or Precibil
Control interval	24 hours recommended
Control sequence	User defined
Control after calibration	Recommended

For quality control, use control materials as listed in the "Order information" section. In addition, other suitable control material can be used.

The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

Follow the applicable government regulations and local guidelines for quality control.

### Limitations - interference

Criterion: Recovery within  $\pm 10\%$  of initial value of direct bilirubin concentration of 34.2  $\mu\text{mol/L}$  (2.0 mg/dL).

Hemolysis:<sup>14</sup> No significant interference up to an H index of 25 (approximate hemoglobin concentration: 15.5  $\mu\text{mol/L}$  or 25 mg/dL).

Lipemia (Intralipid):<sup>14</sup> No significant interference up to an L index of 750. There is poor correlation between the L index (corresponds to turbidity) and triglycerides concentration.

Drugs: No interference was found at therapeutic concentrations using common drug panels.<sup>15,16</sup>

Exception: Phenylbutazone causes artificially low bilirubin results.

Samples containing indocyanine green must not be measured.

In very rare cases, gammopathy, in particular type IgM (Waldenström's macroglobulinemia), may cause unreliable results.<sup>17</sup>

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

In certain cases specimens may give a direct bilirubin result slightly greater than the total bilirubin result. This is observed in patient samples when nearly all the reacting bilirubin is in the direct form. In such cases the result for the total bilirubin should be reported for both direct bilirubin and total bilirubin values.

### ACTION REQUIRED

**Special Wash Programming:** The use of special wash steps is mandatory when certain test combinations are run together on COBAS INTEGRA analyzers. Refer to the CLEAN Method Sheet for further instructions and for the latest version of the Extra wash cycle list.

**Where required, special wash/carry-over evasion programming must be implemented prior to reporting results with this test.**

**Limits and ranges****Measuring range***Jendrassik-Grof method*

1.5-291 µmol/L (0.09-17 mg/dL)

*Doumas method*

1.2-236 µmol/L (0.07-13.8 mg/dL)

Determine samples having higher concentrations via the rerun function. Dilution of samples via the rerun function is a 1:2 dilution. Results from samples diluted using the rerun function are automatically multiplied by a factor of 2.

**Lower limits of measurement***Jendrassik-Grof method**Limit of Blank, Limit of Detection and Limit of Quantitation*

Limit of Blank = 1.0 µmol/L (0.06 mg/dL)

Limit of Detection = 1.5 µmol/L (0.09 mg/dL)

Limit of Quantitation = 3.0 µmol/L (0.18 mg/dL)

The Limit of Blank, the Limit of Detection and the Limit of Quantitation were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A requirements.

The Limit of Blank is the 95<sup>th</sup> percentile value from  $n \geq 60$  measurements of analyte-free samples over several independent series. The Limit of Blank corresponds to the concentration below which analyte-free samples are found with a probability of 95 %.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low concentration samples.

The Limit of Detection corresponds to the lowest analyte concentration which can be detected (value above the Limit of Blank with a probability of 95 %).

The Limit of Quantitation is the lowest analyte concentration that can be reproducibly measured with a total error of 30 %. It has been determined using low concentration direct bilirubin samples.

**Lower limits of measurement***Doumas method**Limit of Blank, Limit of Detection and Limit of Quantitation*

Limit of Blank = 0.8 µmol/L (0.05 mg/dL)

Limit of Detection = 1.2 µmol/L (0.07 mg/dL)

Limit of Quantitation = 1.2 µmol/L (0.07 mg/dL)

The Limit of Blank, the Limit of Detection and the Limit of Quantitation were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A requirements.

The Limit of Blank is the 95<sup>th</sup> percentile value from  $n \geq 60$  measurements of analyte-free samples over several independent series. The Limit of Blank corresponds to the concentration below which analyte-free samples are found with a probability of 95 %.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low concentration samples.

The Limit of Detection corresponds to the lowest analyte concentration which can be detected (value above the Limit of Blank with a probability of 95 %).

The Limit of Quantitation is the lowest analyte concentration that can be reproducibly measured with a precision of 20 % CV. It has been determined using low concentration direct bilirubin samples.

**Specific performance data**

Representative performance data on the analyzers are given below. Results obtained in individual laboratories may differ.

**Precision***Jendrassik-Grof method*

Precision was determined using human samples and controls in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP5 requirements with repeatability ( $n = 21$ ) and intermediate precision (4 aliquots per run, 1 run per day, 21 days). The following results were obtained on the COBAS INTEGRA 800 analyzer:

<i>Repeatability</i>	<i>Mean µmol/L (mg/dL)</i>	<i>SD µmol/L (mg/dL)</i>	<i>CV %</i>
Precinorm U	16.2 (0.95)	0.1 (0.01)	0.9
Precipath U	41.5 (2.43)	0.2 (0.01)	0.5
Human serum 1	2.6 (0.15)	0.2 (0.01)	6.4
Human serum 2	80.5 (4.71)	0.2 (0.01)	0.2
Human serum 3	282 (16.5)	1 (0.1)	0.2

<i>Intermediate precision</i>	<i>Mean µmol/L (mg/dL)</i>	<i>SD µmol/L (mg/dL)</i>	<i>CV %</i>
Precinorm U	15.9 (0.93)	0.3 (0.02)	1.6
Precipath U	40.5 (2.34)	0.4 (0.02)	1.0
Human serum 1	2.4 (0.14)	0.2 (0.01)	7.7
Human serum 2	79.2 (4.63)	0.8 (0.05)	1.0
Human serum 3	278 (16.3)	1 (0.1)	0.4

*Doumas method*

Precision was determined using human samples and controls in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP5 requirements with repeatability ( $n = 21$ ) and intermediate precision (2 aliquots per run, 2 runs per day, 21 days). The following results were obtained on the COBAS INTEGRA 800 analyzer:

<i>Repeatability</i>	<i>Mean µmol/L (mg/dL)</i>	<i>SD µmol/L (mg/dL)</i>	<i>CV %</i>
Precinorm U	12.9 (0.75)	0.2 (0.01)	1.2
Precipath U	32.8 (1.9)	0.2 (0.01)	0.6
Human serum 1	2.0 (0.12)	0.1 (0.01)	7.4
Human serum 2	64.2 (3.8)	0.2 (0.01)	0.4
Human serum 3	225 (13.2)	0.7 (0.04)	0.3

<i>Intermediate precision</i>	<i>Mean µmol/L (mg/dL)</i>	<i>SD µmol/L (mg/dL)</i>	<i>CV %</i>
Precinorm U	12.9 (0.75)	0.2 (0.01)	1.6
Precipath U	32.8 (1.9)	0.3 (0.02)	1.0
Human serum 1	2.0 (0.12)	0.2 (0.01)	7.7
Human serum 2	64.2 (3.8)	0.7 (0.04)	1.0
Human serum 3	225 (13.2)	0.8 (0.05)	0.4

The data obtained on COBAS INTEGRA 800 analyzer(s) are representative for COBAS INTEGRA 400 analyzer(s).

**Method comparison***Jendrassik-Grof method*

Bilirubin values for human serum and plasma samples obtained with the BILD2 reagent (y) on a COBAS INTEGRA 800 analyzer were compared to those determined using the previous Roche BIL-D reagent (x) on the same analyzer.

Sample size ( $n$ ) = 56Passing/Bablok<sup>18</sup> $y = 1.012x + 2.62 \mu\text{mol/L}$  $\tau = 0.964$ 

Linear regression

 $y = 0.997x + 2.89 \mu\text{mol/L}$  $r = 0.997$ 

The sample concentrations were between 7.79 and 273 µmol/L (0.456 and 16.0 mg/dL).

The data obtained on COBAS INTEGRA 800 analyzer(s) are representative for COBAS INTEGRA 400 analyzer(s).

*Doumas method*

Bilirubin values for human serum and plasma samples obtained with the BILD2 reagent (y) on a COBAS INTEGRA 800 analyzer were compared to

those determined using the previous Roche BIL-D reagent (x) on the same analyzer.

Sample size (n) = 71

Passing/Bablok<sup>18</sup> Linear regression  
 $y = 1.049x + 1.20 \mu\text{mol/L}$   $y = 1.020x + 2.09 \mu\text{mol/L}$

$\tau = 0.962$   $r = 0.998$

The sample concentrations were between 1.4 and 235  $\mu\text{mol/L}$  (0.08 and 13.7 mg/dL).

The data obtained on COBAS INTEGRA 800 analyzer(s) are representative for COBAS INTEGRA 400 analyzer(s).

### References




- McPherson RA, Pincus MR. Henry's Clinical Diagnosis and Management by Laboratory Methods. 21st ed. Saunders Elsevier, 2007:1405.
- Sticova E, Jirsa M. New insights in bilirubin metabolism and their clinical implications. World J Gastroenterol 2013;19(38):6398-6407.
- Kwo PY, Cohen SM, Lim JK. ACG Clinical Guideline: Evaluation of Abnormal Liver Chemistries. Am J Gastroenterol 2017;112(1):18-35.
- Newsome PN, Cramb R, Davidson SM, et al. Guidelines on the management of abnormal liver blood tests. Gut 2018;67(1):6-19.
- American Academy of Pediatrics Subcommittee on Hyperbilirubinemia. Management of hyperbilirubinemia in the newborn infant 35 or more weeks of gestation. Pediatrics 2004;114(1):297-316.
- Fawaz R, Baumann U, Ekong U, et al. Guideline for the Evaluation of Cholestatic Jaundice in Infants: Joint Recommendation of the North American Society for Pediatric Gastroenterology, Hepatology, and Nutrition and the European Society for Pediatric Gastroenterology, Hepatology, and Nutrition. J Pediatr Gastroenterol Nutr 2017;64(1):154-168.
- Malloy HT, Evelyn KA. The determination of bilirubin with the photoelectric colorimeter. J Biol Chem 1937;119:481-490.
- Quality of Diagnostic Samples, Recommendations of the Working Group on Preanalytical Quality of the German Society for Clinical Chemistry and Laboratory Medicine, 3rd completely revised ed. 2010.
- Use of Anticoagulants in Diagnostic Laboratory Investigations. WHO Publication WHO/DIL/LAB/99.1 Rev. 2: Jan 2002.
- Soldin JS, Brugnara C, Wong EC. Pediatric Reference Intervals. AACCPress, 5th ed., 2005.
- Balistreri WF, Shaw LM. Liver function. In: Tietz NW, ed. Fundamentals of Clinical Chemistry. 3rd ed. Philadelphia: WB Saunders 1987;729-761.
- Jendrassik L, Grof P. Vereinfachte photometrische Methode zur Bestimmung des Bilirubins. Biochem Z 1938;297:81-89.
- Doumas BT, Perry BW, Jendrzeczak B, et al. Pitfalls in the American Monitor Kit Methods for Determination of Total and Direct Bilirubin. Clin Chem 1982;28(11):2305-2308.
- Glick MR, Ryder KW, Jackson SA. Graphical Comparisons of Interferences in Clinical Chemistry Instrumentation. Clin Chem 1986;32:470-475.
- Breuer J. Report on the Symposium "Drug effects in Clinical Chemistry Methods". Eur J Clin Chem Clin Biochem 1996;34:385-386.
- Sonntag O, Scholer A. Drug interference in clinical chemistry: recommendation of drugs and their concentrations to be used in drug interference studies. Ann Clin Biochem 2001;38:376-385.
- Bakker AJ, Mücke M. Gammopathy interference in clinical chemistry assays: mechanisms, detection and prevention. Clin Chem Lab Med 2007;45(9):1240-1243.
- Bablok W, Passing H, Bender R, et al. A general regression procedure for method transformation. Application of linear regression procedures for method comparison studies in clinical chemistry, Part III. J Clin Chem Clin Biochem 1988 Nov;26(11):783-790.

A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Any serious incident that has occurred in relation to the device shall be reported to the manufacturer and the competent authority of the Member State in which the user and/or the patient is established.

### Symbols

Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard (for USA: see [navifyportal.roche.com](http://navifyportal.roche.com) for definition of symbols used):

	Contents of kit
	Volume for reconstitution
	Global Trade Item Number

Rx only

For USA: Caution: Federal law restricts this device to sale by or on the order of a physician.



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Additions, deletions or changes are indicated by a change bar in the margin.

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