

Total Protein Urine/CSF Gen.3

Order information

REF		CONTENT		Analyzer(s) on which cobas c pack(s) can be used
08058679190*	08058679500	Total Protein Urine/CSF Gen.3 (650 tests)	System-ID 2112 001	cobas c 303, cobas c 503, cobas c 703
08058679214*	08058679500	Total Protein Urine/CSF Gen.3 (650 tests)	System-ID 2112 001	cobas c 303, cobas c 503, cobas c 703

Materials required (but not provided):

03121305122	C.f.a.s. PUC (5 x 1 mL)	Code 20489	
03121313122	Precinorm PUC (4 x 3 mL)	Code 20240	
03121291122	Precipath PUC (4 x 3 mL)	Code 20241	
08063494190	Diluent NaCl 9 % (123 mL)	System-ID 2906 001	

* Some kits shown may not be available in all countries.

English

System information

TPU3: ACN 21122 (Urine)

TPC3: ACN 21123 (CSF)

Intended use

In vitro test for the quantitative determination of protein in human urine and cerebrospinal fluid on **cobas c** systems.

Summary

Protein measurements in human urine with this assay are used in the diagnosis and monitoring of disease conditions characterized by proteinuria or albuminuria, including renal or heart diseases, or thyroid disorders.

Protein measurements in human cerebrospinal fluid (CSF) with this assay are used to assess functional blood-brain barriers disorders in case of conditions such as meningitis, brain tumors and infections of the central nervous system.

During glomerular filtration proteins larger than albumin (66 kDa, diameter 3.5 nm, charge -23) are retained by the healthy glomerulus. Proteins with a lower molecular weight are usually able to freely pass through the glomerular membrane but are actively reabsorbed within the tubular system. Therefore, the presence of significant amounts of protein in the urine is suggestive of renal disease.¹ In addition to its role as a marker for renal disease and renal disease risk,² proteinuria has been shown to be an independent predictor of cardiovascular morbidity and mortality.³ Thyroid hormones can directly affect kidney function, and impaired renal function can also contribute to thyroid disorders, most likely due to urinary loss of protein-bound thyroid hormone.^{4,5} For this reason, hypothyroid patients often present with proteinuria and viceversa chronic kidney disease patients have higher risk of developing hypothyroidism.^{6,7} Most CSF constituents (about 80 %) originate by diffusion from plasma across the blood-CSF barrier.⁸ The concentration of CSF proteins is influenced by multiple factors most significantly by blood concentration, protein size, blood-CSF barrier integrity, and intrathecal production.⁹ Many acute inflammatory diseases of the CNS, including infections and malignancies of the CNS, are characterized by a mild to moderate increase in total protein concentration.^{9,10} Low CSF protein levels can occur in conditions such as repeated lumbar puncture or a chronic leak. Additionally, low CSF protein levels are observed in some children (between 6 months and 2 years of age), in cases of acute water intoxication, and in a minority of patients with idiopathic intracranial hypertension.¹⁰

The Roche Diagnostics Urinary/CSF Protein assay is based on the method described by Iwata and Nishikaze,¹¹ later modified by Luxton, Patel, Keir, and Thompson.¹² In this method, benzethonium chloride reacts with protein in a basic medium to produce a turbidity that is more stable and evenly distributed than that observed with the SSA or TCA methodologies. This assay shows an under recovery of γ -globulin compared to albumin of about 30 %, ¹³ and no interference from magnesium ions due to the addition of EDTA.

Test principle

Turbidimetric method.

The sample is preincubated in an alkaline solution containing EDTA, which denatures the protein and eliminates interference from magnesium ions. Benzethonium chloride is then added, producing turbidity.

Reagents - working solutions

R1 Sodium hydroxide: 677 mmol/L; EDTA-Na: 74 mmol/L

R3 Benzethonium chloride: 32 mmol/L

R1 is in position B and R3 is in position C.

Precautions and warnings

For in vitro diagnostic use for laboratory professionals. Exercise the normal precautions required for handling all laboratory reagents.

Infectious or microbial waste:

Warning: handle waste as potentially biohazardous material. Dispose of waste according to accepted laboratory instructions and procedures.

Environmental hazards:

Apply all relevant local disposal regulations to determine the safe disposal.

Safety data sheet available for professional user on request.

This kit contains components classified as follows in accordance with the Regulation (EC) No. 1272/2008:



Danger

H290 May be corrosive to metals.

H314 Causes severe skin burns and eye damage.

H412 Harmful to aquatic life with long lasting effects.

Prevention:

P273 Avoid release to the environment.

P280 Wear protective gloves/ protective clothing/ eye protection/ face protection/ hearing protection.

Response:

P301 + P330 IF SWALLOWED: Rinse mouth. Do NOT induce vomiting. + P331

P303 + P361 IF ON SKIN (or hair): Take off immediately all contaminated clothing. Rinse skin with water. + P353

P304 + P340 IF INHALED: Remove person to fresh air and keep comfortable for breathing. Immediately call a POISON CENTER/ doctor.

P305 + P351 IF IN EYES: Rinse cautiously with water for several
 + P338 minutes. Remove contact lenses, if present and easy to do.
 + P310 Continue rinsing. Immediately call a POISON CENTER/
 doctor.

Hazardous components:

- sodium hydroxide

Product safety labeling follows EU GHS guidance.

Contact phone: all countries: +49-621-7590

Reagent handling

Ready for use

Storage and stability

Shelf life at 15-25 °C: See expiration date on
cobas c pack label.

On-board in use and refrigerated on the
 analyzer: 26 weeks

Specimen collection and preparation

For specimen collection and preparation only use suitable tubes or
 collection containers.

Only the specimens listed below were tested and found acceptable.

Urine

Use random or 24-hour urine specimens. Use no preservatives. Refrigerate
 specimen during collection.

CSF

No special additives are required. Blood in a CSF specimen invalidates the
 protein value.¹⁴

Samples for urinary/CSF protein should be collected before fluorescein is
 given or at least 24 hours later.¹⁵

Note: Urine, CSF and control samples with a protein concentration above
 7000 mg/L must not be measured with TPUC3 as this may clog the
 instrument lines.

Stability:¹⁶

Urine:	1 day at 15-25 °C
	7 days at 2-8 °C
	1 month at -20 °C (± 5 °C)
CSF:	1 day at 15-25 °C
	6 days at 2-8 °C
	> 1 year at -20 °C (± 5 °C)

Freeze only once.

The sample types listed were tested with a selection of sample collection
 tubes that were commercially available at the time of testing, i.e. not all
 available tubes of all manufacturers were tested. Sample collection systems
 from various manufacturers may contain differing materials which could
 affect the test results in some cases. When processing samples in primary
 tubes (sample collection systems), follow the instructions of the tube
 manufacturer.

Centrifuge samples containing precipitates before performing the assay.

See the limitations and interferences section for details about possible
 sample interferences.

Non centrifuged samples may produce elevated results.

Materials provided

See "Reagents – working solutions" section for reagents.

Materials required (but not provided)

See "Order information" section

General laboratory equipment

Assay

For optimum performance of the assay follow the directions given in this
 document for the analyzer concerned. Refer to the appropriate operator's
 manual for analyzer-specific assay instructions.

The performance of applications not validated by Roche is not warranted
 and must be defined by the user.

Application for urine and CSF

Test definition

Reporting time	10 min		
Wavelength (sub/main)	700/505 nm		
Reagent pipetting		Diluent (H ₂ O)	
R1	75 µL	–	
R3	30 µL	–	
<i>Sample volumes</i>	<i>Sample</i>	<i>Sample dilution</i>	
		<i>Sample</i>	<i>Diluent (NaCl)</i>
Normal	4.5 µL	–	–
Decreased	4.5 µL	25 µL	50 µL
Increased	4.5 µL	–	–

For further information about the assay test definitions refer to the
 application parameters setting screen of the corresponding analyzer and
 assay.

Calibration

Application for urine (ACN 21122)

Calibrators	S1: H ₂ O
	S2-S6: C.f.a.s. PUC
Calibration mode	Non-linear
Calibration frequency	Full calibration
	- after reagent lot change
	- as required following quality control procedures

Application for CSF (ACN 21123)

Calibrators	S1: H ₂ O
	S2-S6: C.f.a.s. PUC
Calibration mode	Non-linear
Calibration frequency	Full calibration
	- after reagent lot change
	- as required following quality control procedures

Calibration interval may be extended based on acceptable verification of
 calibration by the laboratory.

Traceability:¹⁷ This method has been standardized against a primary
 standard traceable to NIST.

Quality control

For quality control, use control materials as listed in the "Order information"
 section. In addition, other suitable control material can be used.

Urine:	Precinorm PUC, Precipath PUC
CSF:	Precinorm PUC, Precipath PUC

The control intervals and limits should be adapted to each laboratory's
 individual requirements. It is recommended to perform quality control
 always after lot calibration and subsequently at least every 26 weeks.

Values obtained should fall within the defined limits. Each laboratory should
 establish corrective measures to be taken if values fall outside the defined
 limits.

Follow the applicable government regulations and local guidelines for
 quality control.

Calculation

cobas c systems automatically calculate the analyte concentration of each
 sample in the unit mg/L (mg/dL, g/L).

Repeatability	Mean	SD	CV
	mg/L	mg/L	%
Precinorm PUC	242	9.60	4.0
Precipath PUC	1615	9.06	0.6
Human CSF 1	130	9.52	7.3
Human CSF 2	357	7.72	2.2
Human CSF 3	501	7.16	1.4
Human CSF 4	1087	8.56	0.8
Human CSF 5	1715	10.9	0.6
Intermediate precision	Mean	SD	CV
	mg/L	mg/L	%
Precinorm PUC	242	11.3	4.7
Precipath PUC	1615	10.7	0.7
Human CSF 1	130	9.88	7.6
Human CSF 2	357	9.57	2.7
Human CSF 3	503	8.29	1.6
Human CSF 4	1063	13.0	1.2
Human CSF 5	1715	12.4	0.7

The data obtained on **cobas c** 503 analyzer(s) are representative for **cobas c** 303 analyzer(s) and **cobas c** 703 analyzer(s).

Method comparison

Total protein values for human urine and CSF samples obtained on a **cobas c** 503 analyzer (y) were compared with those determined using the corresponding reagent on a **cobas c** 501 analyzer (x).

Urine

Sample size (n) = 77

Passing/Bablok ²⁵	Linear regression
$y = 0.952x + 19.3 \text{ mg/L}$	$y = 0.948x + 24.0 \text{ mg/L}$
$\tau = 0.983$	$r = 0.999$

The sample concentrations were between 40.8 and 1784 mg/L.

CSF

Sample size (n) = 75

Passing/Bablok ²⁵	Linear regression
$y = 0.968x + 31.3 \text{ mg/L}$	$y = 0.954x + 38.6 \text{ mg/L}$
$\tau = 0.990$	$r = 1.000$

The sample concentrations were between 43.4 and 1890 mg/L.

Total protein values for human urine and CSF samples obtained on a **cobas c** 303 analyzer (y) were compared with those determined using the corresponding reagent on a **cobas c** 501 analyzer (x).

Urine

Sample size (n) = 71

Passing/Bablok ²⁵	Linear regression
$y = 1.012x + 3.81 \text{ mg/L}$	$y = 1.011x + 6.01 \text{ mg/L}$
$\tau = 0.981$	$r = 0.999$

The sample concentrations were between 45.5 and 1879 mg/L.

CSF

Sample size (n) = 77

Passing/Bablok ²⁵	Linear regression
$y = 1.046x + 17.5 \text{ mg/L}$	$y = 1.029x + 25.4 \text{ mg/L}$
$\tau = 0.987$	$r = 1.000$

The sample concentrations were between 89.8 and 1926 mg/L.

Total protein values for human urine and CSF samples obtained on a **cobas c** 703 analyzer (y) were compared with those determined using the corresponding reagent on a **cobas c** 503 analyzer (x).

Urine

Sample size (n) = 72

Passing/Bablok ²⁵	Linear regression
$y = 0.985x - 2.33 \text{ mg/L}$	$y = 0.990x - 3.37 \text{ mg/L}$
$\tau = 0.979$	$r = 1.000$

The sample concentrations were between 41.8 and 1958 mg/L.

CSF

Sample size (n) = 72

Passing/Bablok ²⁵	Linear regression
$y = 0.987x + 7.13 \text{ mg/L}$	$y = 0.985x + 8.20 \text{ mg/L}$
$\tau = 0.983$	$r = 1.000$

The sample concentrations were between 56.2 and 1970 mg/L.

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A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Any serious incident that has occurred in relation to the device shall be reported to the manufacturer and the competent authority of the Member State in which the user and/or the patient is established.

Symbols

Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard:

CONTENT	Contents of kit
→	Volume for reconstitution
GTIN	Global Trade Item Number
Rx only	For USA: Caution: Federal law restricts this device to sale by or on the order of a physician.

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