

REF			SYSTEM
08443432190*	08443432500	300	cobas e 402
08443432214*			cobas e 801

* Some kits shown may not be available in all countries.

English

System information

Short name	ACN (application code number)
TSH	10172

Intended use

Immunoassay for the in vitro quantitative determination of thyrotropin in human serum and plasma.

The electrochemiluminescence immunoassay "ECLIA" is intended for use on **cobas e** immunoassay analyzers.

Summary

Thyroid-stimulating hormone (TSH, thyrotropin) is a glycoprotein having a molecular weight of approximately 30000 daltons and consisting of two subunits. The β -subunit carries the TSH-specific immunological and biological information, whereas the α -chain carries species-specific information and has an identical amino acid sequence to the α -chains of LH, FSH and hCG.¹

TSH is formed in specific basophil cells of the anterior pituitary and is subject to a circadian secretion sequence. The hypophyseal release of TSH (thyrotropic hormone) is the central regulating mechanism for the biological action of thyroid hormones. TSH has a stimulating action in all stages of thyroid hormone formation and secretion; it also has a proliferative effect.¹

The determination of TSH serves as the initial test in thyroid diagnostics. Even very slight changes in the concentrations of the free thyroid hormones bring about much greater opposite changes in the TSH level. Accordingly, TSH is a very sensitive and specific parameter for assessing thyroid function and is particularly suitable for early detection or exclusion of disorders in the central regulating circuit between the hypothalamus, pituitary and thyroid.^{2,3,4,5,6}

The Elecsys TSH assay employs monoclonal antibodies specifically directed against human TSH. The antibodies labeled with ruthenium complex^{a)} consist of a chimeric construct from human and mouse-specific components. As a result, interfering effects due to HAMA (human anti-mouse antibodies) are largely eliminated.

a) Tris(2,2'-bipyridyl)ruthenium(II)-complex (Ru(bpy)₃²⁺)

Test principle

Sandwich principle. Total duration of assay: 18 minutes.

- 1st incubation: 30 μ L of sample, a biotinylated monoclonal TSH-specific antibody and a monoclonal TSH-specific antibody labeled with a ruthenium complex react to form a sandwich complex.
- 2nd incubation: After addition of streptavidin-coated microparticles, the complex becomes bound to the solid phase via interaction of biotin and streptavidin.
- The reaction mixture is aspirated into the measuring cell where the microparticles are magnetically captured onto the surface of the electrode. Unbound substances are then removed with ProCell II M. Application of a voltage to the electrode then induces chemiluminescent emission which is measured by a photomultiplier.
- Results are determined via a calibration curve which is instrument-specifically generated by 2-point calibration and a master curve provided via the **cobas** link.

Reagents - working solutions

The **cobas e** pack is labeled as TSH.

- M Streptavidin-coated microparticles, 1 bottle, 14.1 mL:
Streptavidin-coated microparticles 0.72 mg/mL; preservative.
- R1 Anti-TSH-Ab-biotin, 1 bottle, 15.8 mL:
Biotinylated monoclonal anti-TSH antibody (mouse) 2.0 mg/L;
phosphate buffer 100 mmol/L, pH 7.2; preservative.

- R2 Anti-TSH-Ab~Ru(bpy)₃²⁺, 1 bottle, 13.9 mL:
Monoclonal anti-TSH antibody (mouse/human) labeled with ruthenium complex 1.5 mg/L; phosphate buffer 100 mmol/L, pH 7.2; preservative.

Precautions and warnings

For in vitro diagnostic use for health care professionals. Exercise the normal precautions required for handling all laboratory reagents.

Infectious or microbial waste:

Warning: handle waste as potentially biohazardous material. Dispose of waste according to accepted laboratory instructions and procedures.

Environmental hazards:

Apply all relevant local disposal regulations to determine the safe disposal.

Safety data sheet available for professional user on request.

This kit contains components classified as follows in accordance with the Regulation (EC) No. 1272/2008:



Warning

- H317 May cause an allergic skin reaction.

Prevention:

- P261 Avoid breathing dust/fume/gas/mist/vapours/spray.
- P272 Contaminated work clothing should not be allowed out of the workplace.
- P280 Wear protective gloves.

Response:

- P333 + P313 If skin irritation or rash occurs: Get medical advice/attention.
- P362 + P364 Take off contaminated clothing and wash it before reuse.

Disposal:

- P501 Dispose of contents/container to an approved waste disposal plant.

Product safety labeling follows EU GHS guidance.

Contact phone: all countries: +49-621-7590

Avoid foam formation in all reagents and sample types (specimens, calibrators and controls).

Reagent handling

The reagents in the kit have been assembled into a ready-for-use unit that cannot be separated.

All information required for correct operation is available via the **cobas** link.

Storage and stability

Store at 2-8 °C.

Do not freeze.

Store the **cobas e** pack **upright** in order to ensure complete availability of the microparticles during automatic mixing prior to use.

Stability:	
unopened at 2-8 °C	up to the stated expiration date
on the analyzers	16 weeks

Specimen collection and preparation

Only the specimens listed below were tested and found acceptable.

Serum collected using standard sampling tubes or tubes containing separating gel.

Li-heparin, K₂-EDTA and K₃-EDTA plasma.

Plasma tubes containing separating gel can be used.

Criterion: Slope 0.9-1.1 + coefficient of correlation \geq 0.95 and with a bias \leq 10 % at medical decision points (0.27 μ IU/mL and 4.2 μ IU/mL).

Stable for 8 days at 20-25 °C, 14 days at 2-8 °C, 24 months at -20 °C (\pm 5 °C). Freeze only once.

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube manufacturer.

Centrifuge samples containing precipitates before performing the assay.

Do not use heat-inactivated samples.

Do not use samples and controls stabilized with azide.

Ensure the samples and calibrators are at 20-25 °C prior to measurement.

Due to possible evaporation effects, samples and calibrators on the analyzers should be analyzed/measured within 2 hours.

Materials provided

See "Reagents – working solutions" section for reagents.

Materials required (but not provided)

- [REF] 08443459190, TSH CalSet, 4 x 1.3 mL
- [REF] 06445918190, PreciControl Thyro Sensitive, for 4 x 2.0 mL
- [REF] 11731416190, PreciControl Universal, for 4 x 3.0 mL
- [REF] 07299010190, Diluent MultiAssay, 36 mL sample diluent
- General laboratory equipment
- **cobas e** analyzer

Additional materials for **cobas e** 402 and **cobas e** 801 analyzers:

- [REF] 06908799190, ProCell II M, 2 x 2 L system solution
- [REF] 04880293190, CleanCell M, 2 x 2 L measuring cell cleaning solution
- [REF] 07485409001, Reservoir Cup, 8 cups to supply ProCell II M and CleanCell M
- [REF] 06908853190, PreClean II M, 2 x 2 L wash solution
- [REF] 05694302001, Assay Tip/Assay Cup tray, 6 magazines x 6 magazine stacks x 105 assay tips and 105 assay cups, 3 wasteliners
- [REF] 07485425001, Liquid Flow Cleaning Cup, 2 adaptor cups to supply ISE Cleaning Solution/Elecsys SysClean for Liquid Flow Cleaning Detection Unit
- [REF] 07485433001, PreWash Liquid Flow Cleaning Cup, 1 adaptor cup to supply ISE Cleaning Solution/Elecsys SysClean for Liquid Flow Cleaning PreWash Unit
- [REF] 11298500316, ISE Cleaning Solution/Elecsys SysClean, 5 x 100 mL system cleaning solution

Assay

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.

Resuspension of the microparticles takes place automatically prior to use.

Place the cooled (stored at 2-8 °C) **cobas e** pack on the reagent manager. Avoid foam formation. The system automatically regulates the temperature of the reagents and the opening/closing of the **cobas e** pack.

Calibration

Traceability: This method has been standardized against the 2nd IRP WHO Reference Standard 80/558.

The predefined master curve is adapted to the analyzer using the relevant CalSet.

Calibration frequency: Calibration must be performed once per reagent lot using fresh reagent (i.e. not more than 24 hours since the **cobas e** pack was registered on the analyzer).

Calibration interval may be extended based on acceptable verification of calibration by the laboratory.

Renewed calibration is recommended as follows:

- after 12 weeks when using the same reagent lot
- after 28 days when using the same **cobas e** pack on the analyzer
- as required: e.g. quality control findings outside the defined limits

Quality control

For quality control, use PreciControl Universal or PreciControl Thyro Sensitive.

In addition, other suitable control material can be used.

Controls for the various concentration ranges should be run individually at least once every 24 hours when the test is in use, once per **cobas e** pack, and following each calibration.

The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

If necessary, repeat the measurement of the samples concerned.

Follow the applicable government regulations and local guidelines for quality control.

Calculation

The analyzer automatically calculates the analyte concentration of each sample either in μ IU/mL or mIU/L (selectable).

Limitations - interference

The effect of the following endogenous substances and pharmaceutical compounds on assay performance was tested. Interferences were tested up to the listed concentrations and no impact on results was observed.

Endogenous substances

Compound	Concentration tested
Bilirubin	\leq 701 μ mol/L or \leq 41 mg/dL
Hemoglobin	\leq 0.621 mmol/L or \leq 1000 mg/dL
Intralipid	\leq 1500 mg/dL
Biotin	\leq 4912 nmol/L or \leq 1200 ng/mL
Rheumatoid factors	\leq 1500 IU/mL
IgG	\leq 2 g/dL
IgM	\leq 0.5 g/dL

Criterion: For concentrations \leq 0.2 μ IU/mL the deviation is \leq 0.02 μ IU/mL. For concentrations $>$ 0.2 μ IU/mL the deviation is \leq 10 %.

There is no high-dose hook effect at TSH concentrations up to 1000 μ IU/mL.

Pharmaceutical substances

In vitro tests were performed on 17 commonly used pharmaceuticals. No interference with the assay was found.

In addition, the following special drugs were tested. No interference with the assay was found.

Special drugs

Drug	Concentration tested mg/L
Iodide	0.2
Carbimazole	30
Methimazole	80
Propylthiouracil	60
Perchlorate	2000
Propranolol	240

Drug	Concentration tested mg/L
Amiodarone	200
Prednisolone	100
Hydrocortisone	200
Fluocortolone	100
Octreotide	0.3
Levothyroxine	0.25
Liothyronine	0.015

Drug interferences are measured based on recommendations given in CLSI guidelines EP07 and EP37 and other published literature. Effects of concentrations exceeding these recommendations have not been characterized.

The presence of autoantibodies may induce high molecular weight complexes (macro-TSH) which may cause unexpectedly high values of TSH.⁷

In rare cases, interference due to extremely high titers of antibodies to analyte-specific antibodies, streptavidin or ruthenium can occur. These effects are minimized by suitable test design.

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

Limits and ranges

Measuring range

0.005-100 $\mu\text{IU/mL}$ (defined by the Limit of Detection and the maximum of the master curve). Values below the Limit of Detection are reported as $< 0.005 \mu\text{IU/mL}$. Values above the measuring range are reported as $> 100 \mu\text{IU/mL}$ (or up to 1000 $\mu\text{IU/mL}$ for 10-fold diluted samples).

Lower limits of measurement

Limit of Blank, Limit of Detection and Limit of Quantitation

Limit of Blank = 0.0025 $\mu\text{IU/mL}$

Limit of Detection = 0.005 $\mu\text{IU/mL}$

Limit of Quantitation = 0.005 $\mu\text{IU/mL}$

The Limit of Blank, Limit of Detection and Limit of Quantitation were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A2 requirements.

The Limit of Blank is the 95th percentile value from $n \geq 60$ measurements of analyte-free samples over several independent series. The Limit of Blank corresponds to the concentration below which analyte-free samples are found with a probability of 95 %.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low concentration samples. The Limit of Detection corresponds to the lowest analyte concentration which can be detected (value above the Limit of Blank with a probability of 95 %).

The Limit of Quantitation is the lowest analyte concentration that can be reproducibly measured with an intermediate precision CV of $\leq 20 \%$.

Dilution

Samples with TSH concentrations above the measuring range can be diluted with Diluent MultiAssay. The recommended dilution is 1:10 (either automatically by the analyzers or manually). The concentration of the diluted sample must be $\geq 10 \mu\text{IU/mL}$.

After manual dilution, multiply the result by the dilution factor.

After dilution by the analyzers, the software automatically takes the dilution into account when calculating the sample concentration.

Expected values

0.270-4.20 $\mu\text{IU/mL}$ ⁸

These values correspond to the 2.5th and 97.5th percentiles of results obtained from a total of 516 healthy test subjects examined.

For detailed information about reference intervals in children, adolescents and pregnant women, refer to the brochure "Reference Intervals for Children and Adults", English: [REF] 04640292.

This booklet also contains results of a detailed study about influencing factors on thyroid parameters in a well characterized reference group of

adults. Different inclusion and exclusion criteria were applied (e.g. sonographic results (thyroid volume and density) as well as criteria according to the guidelines of the National Academy of Clinical Biochemistry - NACB).

Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

Specific performance data

Representative performance data on the analyzers are given below. Results obtained in individual laboratories may differ.

Precision

Precision was determined using Elecsys reagents, samples and controls in a protocol (EP05-A3) of the CLSI (Clinical and Laboratory Standards Institute): 2 runs per day in duplicate each for 21 days ($n = 84$). The following results were obtained:

cobas e 402 and cobas e 801 analyzers					
Sample	Mean $\mu\text{IU/mL}$	Repeatability		Intermediate precision	
		SD $\mu\text{IU/mL}$	CV %	SD $\mu\text{IU/mL}$	CV %
Human serum 1	0.015	0.001	3.4	0.002	11.2
Human serum 2	0.281	0.003	1.1	0.005	1.8
Human serum 3	4.09	0.030	0.7	0.090	2.2
Human serum 4	59.8	0.494	0.8	1.10	1.8
Human serum 5	95.6	0.706	0.7	3.81	4.0
PC ^b Universal 1	1.39	0.013	1.0	0.021	1.5
PC Universal 2	7.99	0.064	0.8	0.132	1.6
PC Thyro Sensitive	0.176	0.002	1.2	0.004	2.1

b) PC = PreciControl

Method comparison

a) A comparison of the Elecsys TSH assay, [REF] 08443432190 (cobas e 801 analyzer; y), with the Elecsys TSH assay, [REF] 07028091190 (cobas e 801 analyzer; x), gave the following correlations ($\mu\text{IU/mL}$):

Number of samples measured: 138

Passing/Bablok⁹ Linear regression
 $y = 0.974x - 0.000$ $y = 0.944x + 0.089$
 $\tau = 0.972$ $r = 0.999$

The sample concentrations were between 0.006 and 99.2 $\mu\text{IU/mL}$.

b) A comparison of the Elecsys TSH assay, [REF] 08443432190 (cobas e 402 analyzer; y), with the Elecsys TSH assay, [REF] 08443432190 (cobas e 801 analyzer; x), gave the following correlations ($\mu\text{IU/mL}$):

Number of samples measured: 138

Passing/Bablok⁹ Linear regression
 $y = 0.972x - 0.003$ $y = 0.968x + 0.014$
 $\tau = 0.987$ $r = 1.00$

The sample concentrations were between 0.006 and 90.6 $\mu\text{IU/mL}$.

Analytical specificity

The following cross-reactivities were found, tested with a TSH concentration of approximately 0.35 $\mu\text{IU/mL}$.

Cross-reactant	Concentration tested mU/mL	Cross-reactivity %
LH	10000	0.000
FSH	10000	0.000
hGH	1000	n. d. ^{c)}
hCG	50000	0.000

c) n. d. = not detectable

References

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- 2 Wu AHB. Tietz Clinical Guide To Laboratory Tests. Saunders Elsevier, Philadelphia, 4th edition, 2006, section II, p. 1040-1043.
- 3 Surks MI, Chopra IJ, Mariash CN, et al. American Thyroid Association Guidelines for the Use of Laboratory Tests in Thyroid Disorders. JAMA 1990;263:1529-1532.
- 4 Keffer JH. Preanalytical Considerations in Testing Thyroid Function. Clin Chem 1996;42(1):125-135.
- 5 Ladenson PW. Optimal laboratory testing for diagnosis and monitoring of thyroid nodules, goiter and thyroid cancer. Clin Chem 1996;42(1):183-187.
- 6 Nicoloff JT, Spencer CA. The use and misuse of the sensitive thyrotropin assays. J Clin Endocr Metab 1990;71:553-558.
- 7 Sakai H, Fukuda G, Suzuki N, et al. Falsely Elevated Thyroid-Stimulating Hormone (TSH) Level Due to Macro-TSH. Endocr J 2009;56(3):435-440.
- 8 Ebert C, Bieglmayer C, Igari J, et al. Elecsys TSH, FT4, T4, T-uptake, FT3 and T3. Clinical results of a multicentre study. Wien Klin Wochenschr 1998;110 Suppl 3:27-40.
- 9 Bablok W, Passing H, Bender R, et al. A general regression procedure for method transformation. Application of linear regression procedures for method comparison studies in clinical chemistry, Part III. J Clin Chem Clin Biochem 1988 Nov;26(11):783-790.







For further information, please refer to the appropriate operator's manual for the analyzer concerned, the respective application sheets and the Method Sheets of all necessary components (if available in your country).

A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Any serious incident that has occurred in relation to the device shall be reported to the manufacturer and the competent authority of the Member State in which the user and/or the patient is established.

Symbols

Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard (for USA: see dialog.roche.com for definition of symbols used):

	Contents of kit
	Analyzers/Instruments on which reagents can be used
	Reagent
	Calibrator
	Volume for reconstitution
	Global Trade Item Number



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