


## Homocysteine Enzymatic Assay

### Order information

REF		CONTENT		Analyzer(s) on which <b>cobas c</b> pack(s) can be used
06542921190	06542921500	Homocysteine Enzymatic Assay (200 tests)	System-ID 03 7487 1	<b>cobas c</b> 701/702
			System-ID 04 7487 1	

Materials required (but not provided):

05385504190	HCYS Calibrator Kit (2 x 3 mL)	Code 590	
05142423190	HCYS Control Kit Control 1 (2 x 3 mL)	Code 254	
	HCYS Control Kit Control 2 (2 x 3 mL)	Code 255	
05172152190	Diluent NaCl 9 % (119 mL)	System-ID 08 6869 3	

### English

#### System information

HCYS: ACN 8778

#### Intended use

In vitro test for the quantitative determination of total L-homocysteine in human serum and plasma on **cobas c** systems. The assay can assist in the diagnosis of patients suspected of having hyperhomocysteinemia or homocystinuria.

#### Summary

Homocysteine (Hcy) measurements, performed with this assay in human serum and plasma, are used as an aid in the diagnosis of patients suspected of having hyperhomocysteinemia or homocystinuria.

Homocysteine (Hcy) is a thiol-containing amino acid produced by the intracellular demethylation of methionine. Hcy is metabolized by one of two pathways: transsulfuration and remethylation. B-vitamins and folate are needed as substrates in the homocysteine metabolism. Elevation of plasma Hcy levels is called hyperhomocysteinemia. Total homocysteine (tHcy) represents the sum of all forms of Hcy including oxidized, protein-bound, thiolactone and free forms. Hyperhomocysteinemia can be defined in relation to tHcy levels as either moderate, intermediate, or severe.<sup>1,2</sup> Elevations in Hcy levels can be due to:

- genetic deficiencies in enzymes involved in Hcy metabolism such as cystathionine beta-synthase (CBS), methionine synthase, and methylenetetrahydrofolate reductase;
- nutritional deficiency in B-vitamins (B6, B12 and folate), levels of which correlate inversely with Hcy levels;
- renal failure due to defective clearance of Hcy from the kidney;
- other causes including thyroid dysfunction, cancer, psoriasis, and diabetes as well as various drugs, alcohol, tobacco, coffee, older age and menopause.<sup>2,3</sup>

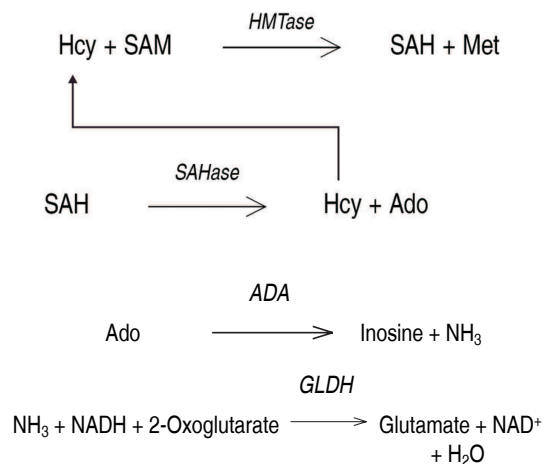
Severe hyperhomocysteinemia occurs in homocystinuria, an innate metabolic disorder characterized by deficiencies in cystathionine β-synthase (CBS), one of the enzymes involved in the metabolism of Hcy to cysteine.<sup>4</sup> For diagnosis of CBS deficiency, measuring tHcy levels is recommended. The diagnosis is very likely if elevated tHcy is accompanied by high or borderline-high plasma methionine-concentrations, and further supported if sensitive methods demonstrate low plasma-cystathionine concentrations with an increased methionine-to-cystathionine ratio.<sup>5,6</sup> Hyperhomocysteinemia is also the hallmark of remethylation inherited disorders. Plasma tHcy is recommended as the first biochemical parameter to assess when a remethylation disorder is suspected. Individuals with remethylation disorders typically have elevated tHcy concentrations but – in contrast to those with CBS deficiency – methionine levels are usually normal or decreased.<sup>6,7</sup> Measurement of tHcy is also recommended to rule out CBS deficiency in the differential diagnosis of inherited methylation disorders such as methionine adenosyltransferase I/III deficiency and glycine N-methyltransferase deficiency. In this case, a low or only slightly increased tHcy level can help exclude CBS deficiency.<sup>8</sup> Elevated levels of tHcy have been found to be associated with numerous pathological conditions, including cardiovascular disease (CVD), stroke, Alzheimer's disease, eye diseases, chronic kidney disease, bone tissue damages, gastrointestinal disorders, cancer, congenital defects, and pregnancy complications.<sup>2,3,9,10,11,12,13,14,15,16,17,18,19,20,21</sup>

- Hcy has been considered as a marker for CVD risk by clinical societies, further investigations are required in order to evaluate if it is also to be seen as a true causal risk factor.<sup>9,10,11,22,23</sup>

Guidelines for tHcy determination in clinical laboratories have been established.<sup>24,25</sup>

#### Test principle

Homocysteine Enzymatic Assay is based on a novel enzyme cycling assay principle that assesses the co-substrate conversion product instead of assessing co-substrate or Hcy conversion products of Hcy. In this assay, oxidized Hcy is first reduced to free Hcy which then reacts with a co-substrate, S-adenosylmethionine (SAM), to form methionine (Met) and S-adenosylhomocysteine (SAH), catalyzed by a Hcy S-methyltransferase. SAH is assessed by coupled enzyme reactions where SAH is hydrolyzed into adenosine (Ado) and Hcy by SAH hydrolase, and Hcy is cycled into the Hcy conversion reaction to form a reaction cycle that amplifies the detection signal. The formed Ado is immediately hydrolyzed into inosine and ammonia. In the last step, the enzyme glutamate dehydrogenase (GLDH) catalyzes the reaction of ammonia with 2-oxoglutarate and NADH to form NAD<sup>+</sup>. The concentration of Hcy in the sample is directly proportional to the amount of NADH converted to NAD<sup>+</sup> ( $\Delta A_{340\text{ nm}}$ ).



#### Reagents - working solutions

##### R1 NADH reagent

S-adenosylmethionine 0.1 mmol/L, TCEP\* > 0.5 mmol/L, 2-oxoglutarate < 5.0 mmol/L, NADH > 0.2 mmol/L, buffer, pH 9.1 (25 °C), preservative, stabilizer

##### R2 Enzyme reagent

Homocysteine S-methyltransferase (HMTase): 5.0 kU/L, glutamate dehydrogenase (GLDH) 10 kU/L, casein (bovine) ≤ 0.2 %, buffer, pH 7.2 (25 °C), preservative, detergent

##### R3 Start reagent

Adenosine deaminase (bovine) 5.0 kU/L, S-adenosyl-homocysteine hydrolase (SAHase) 3.0 kU/L, casein (bovine) ≤ 0.2 %, buffer, pH 7.2 (25 °C), preservative, stabilizer

\*Tris(2-carboxyethyl)phosphine

## Homocysteine Enzymatic Assay

Cat. No. 06542921190 consists of 2 **cobas c** packs: 1 x R1 + R2 and 1 x R3.

R1 is in position B and R2 is in position C of **cobas c** pack 1. R3 is in position C of **cobas c** pack 2.

### Precautions and warnings

For in vitro diagnostic use for health care professionals. Exercise the normal precautions required for handling all laboratory reagents.

Infectious or microbial waste:

Warning: handle waste as potentially biohazardous material. Dispose of waste according to accepted laboratory instructions and procedures.

Environmental hazards:

Apply all relevant local disposal regulations to determine the safe disposal.

Safety data sheet available for professional user on request.

### Reagent handling

Ready for use

### Storage and stability

Shelf life at 2-8 °C: See expiration date on **cobas c** pack label.

On-board in use and refrigerated on the analyzer: 4 weeks

On-board on the reagent manager: 1 hour

### Do not freeze.

### Specimen collection and preparation

For specimen collection and preparation only use suitable tubes or collection containers.

Only the specimens listed below were tested and found acceptable.  
Serum.

Plasma: Li-heparin, K<sub>2</sub>-EDTA and K<sub>3</sub>-EDTA plasma.

It is important to centrifuge blood samples immediately after collection to separate the plasma from the blood cells. If immediate centrifugation is not possible, collected blood specimens should be kept on ice and centrifuged within an hour. Hemolysed or turbid specimens or severely lipemic specimens are not recommended for the Hcy assay.

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube manufacturer.

Centrifuge samples containing precipitates before performing the assay.

See the limitations and interferences section for details about possible sample interferences.

Stability: 4 days at 15-25 °C<sup>25,26</sup>

4 weeks at 2-8 °C<sup>27</sup>

10 months at -20 °C (± 5°C)<sup>27</sup>

Freeze only once.

### Materials provided

See "Reagents – working solutions" section for reagents.

### Materials required (but not provided)

See "Order information" section

General laboratory equipment

### Assay

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.

The performance of applications not validated by Roche is not warranted and must be defined by the user.

### Application for serum and plasma

#### cobas c 701/702 test definition

Assay type 2-Point End

Reaction time / Assay points 10 / 28-38

Wavelength (sub/main) 700/340 nm

Reaction direction Decrease

Units µmol/L

Reagent pipetting Diluent (H<sub>2</sub>O)

R1 176 µL –

R2 28 µL –

R3 20 µL –

	Sample volumes	Sample dilution	
		Sample	Diluent (NaCl)
Normal	14 µL	–	–
Decreased	14 µL	30 µL	120 µL
Increased	14 µL	–	–

### Calibration

Calibrators S1-5: HCYS Calibrator Kit

Multiply the lot-specific HCYS Calibrator Kit calibrator value by the factors below to determine the standard concentrations for the 5-point calibration curve:

S1: 0.050 S4: 0.500

S2: 0.100 S5: 1.00

S3: 0.250

Calibration mode RCM

Calibration frequency Full calibration

- every 7 days on-board
- after reagent lot change
- as required following quality control procedures

Calibration interval may be extended based on acceptable verification of calibration by the laboratory.

Traceability: This method has been standardized against NIST SRM 1955 reference material.

### Quality control

For quality control, use control materials as listed in the "Order information" section.

In addition, other suitable control material can be used.

The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

Follow the applicable government regulations and local guidelines for quality control.

### Calculation

**cobas c** systems automatically calculate the analyte concentration of each sample.

### Limitations - interference

Criterion: Recovery within ± 10 % of initial value for analyte concentrations > 15 µmol/L or ± 1.5 µmol/L for analyte concentrations ≤ 15 µmol/L.

Icterus:<sup>28</sup> No significant interference up to an I index of 20 for conjugated and unconjugated bilirubin (approximate conjugated and unconjugated bilirubin concentration: 342 µmol/L or 20 mg/dL).

Hemolysis:<sup>28</sup> No significant interference up to an H index of 100 (approximate hemoglobin concentration: 62 µmol/L or 100 mg/dL).

Lipemia (Intralipid):<sup>28</sup> No significant interference up to an L index of 250. There is poor correlation between the L index (corresponds to turbidity) and triglycerides concentration.

## Homocysteine Enzymatic Assay

Drugs: No interference was found at therapeutic concentrations using common drug panels.<sup>29,30</sup>

Exceptions: 0.5 mmol/L Glutathione, 100 µmol/L Cystathionine, 0.5 mmol/L Pyruvate.

Patients who are taking methotrexate, carbamazepine, phenytoin, nitrous oxide, anticonvulsants, or 6-azuridine triacetate, may have higher levels of Hcy due to metabolic interference with Hcy metabolism.<sup>24,27</sup>

S-Adenosylhomocysteine (SAH) will cause a significant positive interference. However, SAH is only detectable at sub-nmol/L concentrations in normal plasma, and should not cause concern.<sup>31</sup>

Addition of 3-deazaadenosine to inhibit Hcy production in red cells has been suggested. However, the Homocysteine Enzymatic Assay can not use samples containing 3-deazaadenosine since it inhibits one of the key enzymes used in the assay.

In very rare cases, gammopathy, in particular type IgM (Waldenström's macroglobulinemia), may cause unreliable results.<sup>32</sup>

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

### ACTION REQUIRED

**Special Wash Programming:** The use of special wash steps is mandatory when certain test combinations are run together on **cobas c** systems. All special wash programming necessary for avoiding carry-over is available via the **cobas** link, manual input is required in certain cases. The latest version of the carry-over evasion list can be found with the NaOHD/SMS/SmpCln1+2/SCCS Method Sheet and for further instructions refer to the operator's manual.

**Where required, special wash/carry-over evasion programming must be implemented prior to reporting results with this test.**

### Limits and ranges

#### Measuring range

3-50 µmol/L

Determine samples having higher concentrations via the rerun function. Dilution of samples via the rerun function is a 1:5 dilution. Results from samples diluted using the rerun function are automatically multiplied by a factor of 5.

#### Lower limits of measurement

*Limit of Blank, Limit of Detection and Limit of Quantitation*

Limit of Blank = 3 µmol/L

Limit of Detection = 3 µmol/L

Limit of Quantitation = 5.5 µmol/L

The Limit of Blank and Limit of Detection were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A requirements.

The Limit of Blank is the 95<sup>th</sup> percentile value from  $n \geq 60$  measurements of analyte-free samples over several independent series. The Limit of Blank corresponds to the concentration below which analyte-free samples are found with a probability of 95 %.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low concentration samples.

The Limit of Detection corresponds to the lowest analyte concentration which can be detected (value above the Limit of Blank with a probability of 95 %).

The Limit of Quantitation is the lowest analyte concentration that can be reproducibly measured with a total error of 30 %. It has been determined using low concentration homocysteine samples.

### Expected values

In most of the U.S. clinical laboratories, 15 µmol/L is used as the cut-off value for normal levels of Hcy in adults.

In European laboratories, 12 µmol/L is used as the cut-off value for normal levels of Hcy in adults.<sup>25</sup>

Age, pregnancy, and renal function are important. The intake of folic acid as either supplement or through fortification of foods must also be considered:

Group	Folate supplemented	Nonsupplemented
(fasting/basal tHcy, µmol/L)		
Pregnancy	8	10
Children < 15 years	8	10
Adults 15-65 years	12	15
Elderly > 65 years	16	20

Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

### Specific performance data

Representative performance data on the analyzers are given below. Results obtained in individual laboratories may differ.

#### Precision

Precision was determined using human samples and controls in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP5 requirements with repeatability ( $n = 21$ ) and intermediate precision (2 aliquots per run, 2 runs per day, 21 days). The following results were obtained on the **cobas c 701** analyzer:

Repeatability	Mean	SD	CV
	µmol/L	µmol/L	%
Homocysteine Control 1	12.3	0.2	1.3
Homocysteine Control 2	38.9	0.4	1.1
Human serum A	6.15	0.13	2.1
Human serum B	16.9	0.2	1.4
Human serum C	23.3	0.3	1.3

Intermediate precision	Mean	SD	CV
	µmol/L	µmol/L	%
Homocysteine Control 1	12.2	0.3	2.1
Homocysteine Control 2	39.1	0.8	2.0
Human serum 1	8.26	0.19	2.3
Human serum 2	13.1	0.3	2.1
Human serum 3	30.0	0.5	1.8
Human serum 4	44.4	1.0	2.2

Results for intermediate precision were obtained on the **cobas c 501** analyzer.

The data obtained on **cobas c 501** analyzer(s) are representative for **cobas c 701** analyzer(s).

### Method comparison

Hcy values for human serum samples obtained on the **cobas c 701** analyzer (y) were compared with those determined using the corresponding reagent on a **cobas c 501** analyzer (x).

Sample size ( $n$ ) = 62

Passing/Bablok <sup>33</sup>	Linear regression
$y = 0.979x + 0.267 \text{ µmol/L}$	$y = 0.974x + 0.212 \text{ µmol/L}$
$\tau = 0.974$	$r = 0.997$

The sample concentrations were between 3.65 and 49.0 µmol/L.

### References

- Selhub J. Homocysteine metabolism. Annu Rev Nutr 1999;19:217-246. doi: 10.1146/annurev.nutr.19.1.217. PMID: 10448523
- Škovierová H, Vidomanová E, Mahmood S, et al. The Molecular and Cellular Effect of Homocysteine Metabolism Imbalance on Human Health. Int J Mol Sci 2016 Oct 20;17(10):1733.

- 3 Kluijtmans LA, Young IS, Boreham CA, et al. Genetic and nutritional factors contributing to hyperhomocysteinemia in young adults. *Blood* 2003 Apr 1;101(7):2483-2488.
- 4 Pasquali M, Longo N. Newborn screening and inborn errors of metabolism. Rifai N, Chiu RWK, Young I, Burnham CAD, Wittwer CT, editors. *Tietz Textbook of Laboratory Medicine*, Saunders Elsevier, Philadelphia, 7th edition, 2023, ch. 60, p. 886-887.e37.
- 5 Morris AA, Kožich V, Santra S, et al. Guidelines for the diagnosis and management of cystathionine beta-synthase deficiency. *J Inherit Metab Dis* 2017 Jan;40(1):49-74.
- 6 Huemer M, Kožich V, Rinaldo P, et al. Newborn screening for homocystinurias and methylation disorders: systematic review and proposed guidelines. *J Inherit Metab Dis* 2015 Nov;38(6):1007-1019.
- 7 Huemer M, Diodato D, Schwahn B, et al. Guidelines for diagnosis and management of the cobalamin-related remethylation disorders cblC, cblD, cblE, cblF, cblG, cblJ and MTHFR deficiency. *J Inherit Metab Dis* 2017 Jan;40(1):21-48.
- 8 Barić I, Stauffer C, Augoustides-Savvopoulou P, et al. Consensus recommendations for the diagnosis, treatment and follow-up of inherited methylation disorders. *J Inherit Metab Dis* 2017 Jan;40(1):5-20.
- 9 Ospina-Romero M, Cannegieter SC, den Heijer M, et al. Hyperhomocysteinemia and Risk of First Venous Thrombosis: The Influence of (Unmeasured) Confounding Factors. *Am J Epidemiol* 2018 Jul 1;187(7):1392-1400.
- 10 Homocysteine Studies Collaboration. Homocysteine and risk of ischemic heart disease and stroke: a meta-analysis. *JAMA* 2002 Oct 23-30;288(16):2015-2022.
- 11 Bazzano LA, Reynolds K, Holder KN, et al. Effect of folic acid supplementation on risk of cardiovascular diseases: a meta-analysis of randomized controlled trials. *JAMA* 2006 Dec 13;296(22):2720-2726.
- 12 Christine CW, Auinger P, Joslin A, et al. Vitamin B12 and Homocysteine Levels Predict Different Outcomes in Early Parkinson's Disease. *Mov Disord* 2018 May;33(5):762-770.
- 13 Licking N, Murchison C, Cholerton B, et al. Homocysteine and cognitive function in Parkinson's disease. *Parkinsonism Relat Disord* 2017 Nov;44:1-5.
- 14 Clarke R, Bennett D, Parish S, et al. Effects of homocysteine lowering with B vitamins on cognitive aging: meta-analysis of 11 trials with cognitive data on 22,000 individuals. *Am J Clin Nutr* 2014 Aug;100(2):657-666.
- 15 Smith AD, Refsum H, Bottiglieri T, et al. Homocysteine and Dementia: An International Consensus Statement. *J Alzheimers Dis* 2018;62(2):561-570.
- 16 van Meurs JB, Dhonukshe-Rutten RA, Pluijm SM, et al. Homocysteine levels and the risk of osteoporotic fracture. *N Engl J Med* 2004 May 13;350(20):2033-2041.
- 17 McLean RR, Jacques PF, Selhub J, et al. Homocysteine as a predictive factor for hip fracture in older persons. *N Engl J Med* 2004;350(20):2042-2049.
- 18 Dhonukshe-Rutten RA, Pluijm SM, de Groot LC, et al. Homocysteine and vitamin B12 status relate to bone turnover markers, broadband ultrasound attenuation, and fractures in healthy elderly people. *J Bone Miner Res* 2005 Jun;20(6):921-929.
- 19 Gerdhem P, Ivaska KK, Isaksson A, et al. Associations between homocysteine, bone turnover, BMD, mortality, and fracture risk in elderly women. *J Bone Miner Res* 2007 Jan;22(1):127-134.
- 20 Sawka AM, Ray JG, Yi Q, et al. Randomized clinical trial of homocysteine level lowering therapy and fractures. *Arch Intern Med* 2007 Oct 22;167(19):2136-2139.
- 21 Ren A, Wang J. Methylenetetrahydrofolate reductase C677T polymorphism and the risk of unexplained recurrent pregnancy loss: a meta-analysis. *Fertil Steril* 2006 Dec;86(6):1716-1722.
- 22 Kinoshita M, Yokote K, Arai H, et al. Japan Atherosclerosis Society (JAS) Guidelines for Prevention of Atherosclerotic Cardiovascular Diseases 2017. *J Atheroscler Thromb* 2018 Sep 1;25(9):846-984.
- 23 NACB LMPG Committee Members; Myers GL, Christenson RH, Cushman M, et al. National Academy of Clinical Biochemistry Laboratory Medicine Practice guidelines: emerging biomarkers for primary prevention of cardiovascular disease. *Clin Chem* 2009 Feb;55(2):378-384.
- 24 Refsum H. Total Homocysteine: Guidelines for Determination in the Clinical Laboratory. *Clin Lab News* 2002 May;12-14 ([www.aacc.org](http://www.aacc.org)).
- 25 Refsum H, Smith AD, Ueland PM, et al. Facts and Recommendations about Total Homocysteine Determinations: An Expert Opinion. *Clin Chem* 2004;50(1):3-32.
- 26 Fiskerstrand T, Refsum H, Kvalheim G, et al. Homocysteine and other thiols in plasma and urine: Automated determination and sample stability. *Clin Chem* 1993 Feb;39(2):263-271.
- 27 Rasmussen K and Moller J. Total homocysteine measurement in clinical practice. *Ann Clin Biochem* 2000;37:627-648.
- 28 Glick MR, Ryder KW, Jackson SA. Graphical Comparisons of Interferences in Clinical Chemistry Instrumentation. *Clin Chem* 1986;32:470-475.
- 29 Breuer J. Report on the Symposium "Drug effects in Clinical Chemistry Methods". *Eur J Clin Chem Clin Biochem* 1996;34:385-386.
- 30 Sonntag O, Scholer A. Drug interference in clinical chemistry: recommendation of drugs and their concentrations to be used in drug interference studies. *Ann Clin Biochem* 2001;38:376-385.
- 31 Loehrer FM, Angst CP, Brunner FP, et al. Evidence for disturbed S-adenosylmethionine: S-adenosylhomocysteine ratio in patients with end-stage renal failure: a cause for disturbed methylation reactions? *Nephrol Dial Transplant* 1998;13(3):656-661.
- 32 Bakker AJ, Mücke M. Gammopathy interference in clinical chemistry assays: mechanisms, detection and prevention. *Clin Chem Lab Med* 2007;45(9):1240-1243.
- 33 Bablok W, Passing H, Bender R, et al. A general regression procedure for method transformation. Application of linear regression procedures for method comparison studies in clinical chemistry, Part III. *J Clin Chem Clin Biochem* 1988 Nov;26(11):783-790.

A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Any serious incident that has occurred in relation to the device shall be reported to the manufacturer and the competent authority of the Member State in which the user and/or the patient is established.

The Summary of Safety & Performance Report can be found here: <https://ec.europa.eu/tools/eudamed>

### Symbols

Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard (for USA: see [navifyportal.roche.com](http://navifyportal.roche.com) for definition of symbols used):

**CONTENT**

Contents of kit



Volume for reconstitution

**GTIN**

Global Trade Item Number

Rx only

For USA: Caution: Federal law restricts this device to sale by or on the order of a physician.

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# HCYS

Homocysteine Enzymatic Assay

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CE 0123



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