For general laboratory use.

LightCycler® Multiplex DNA Master

Version: 07

Content Version: November 2023

Easy-to-use 5x reaction mix optimized for multiplex qPCR, compatible with the LightCycler® PRO, LightCycler® 480, LightCycler® 96, or the LightCycler® 2.0 Real-Time PCR Systems.

Cat. No. 07 339 585 001	1 kit 200 reactions of 20 µL final volume each
Cat. No. 07 339 577 001	1 kit 1,000 reactions of 20 µL final volume each

Store the kit at −15 to −25°C.

1. General Information

1.1. Contents

Vial / Bottle	Cap	Label	Function / Description	Catalog Number	Content
1	red	LightCycler® Multiplex DNA Master, qPCR Reaction Mix, 5x conc.	Contains qPCR Reaction Buffer, AptaTaq Polymerase, dATP, dCTP, dGTP, dUTP, $MgCl_2$, and proprietary additives.	07 339 585 001	1 vial, 880 µL
				07 339 577 001	5 vials, 880 µL each
2	colorless	LightCycler® Multiplex DNA Master, Water, PCR Grade	To adjust the final reaction volume.	07 339 585 001	3 vials, 1 mL each
				07 339 577 001	15 vials, 1 mL each

1.2. Storage and Stability

Storage Conditions (Product)

The kit is shipped on dry ice.
When stored at −15 to −25°C, the kit is stable through the expiry date printed on the label. The kit is stable at +2 to +8°C for 4 weeks.

Vial / Bottle	Cap	Label	Storage
1	red	qPCR Reaction Mix, 5x conc.	Store at −15 to −25°C. ⚠ ***Avoid repeated freezing and thawing (more than 5 times).*** Aliquot Vial 1 and freeze or store Vial 1 at +2 to +8°C for a maximum of 4 weeks.
2	colorless	Water, PCR Grade	Store at −15 to −25°C or store at +2 to +8°C for a maximum of 4 weeks.

Storage Conditions (Working Solution)

Prepare the reagents right before use. The working solution, see Section, **Protocols, Setup of the qPCR reaction**, is stable at +15 to +25°C for up to 4 hours, and is therefore ideal for use in automated workflows.

1.3. Additional Equipment and Reagents Required

Standard laboratory equipment

- Nuclease-free pipette tips
- 1.5 mL RNase-free microcentrifuge tubes to prepare master mixes and dilutions
- To minimize risk of RNase contamination, autoclave all vessels

For qPCR

- Real-Time PCR systems such as the LightCycler® PRO, LightCycler® 480, LightCycler® 96, LightCycler® 2.0 Systems*, or other Real-Time PCR Systems
- LightCycler® 480 Multiwell Plate 96, white*
- LightCycler® 480 Multiwell Plate 384, white*
- LightCycler® 480 Multiwell Plate 96, white, 4 bar codes*
- LightCycler® 480 Multiwell Plate 384, white, 4 bar codes*
- Sealing Foil Applicator*
- LightCycler® 480 Sealing Foil*
- LightCycler® 8-Tube Strips (white)*
- LightCycler® 8-Tube Strip Adapter Plate*
- Centrifuge with swinging-bucket rotor

1.4. Application

The LightCycler® Multiplex DNA Master is designed for fast, highly sensitive and specific real-time PCR analysis of DNA.
The single-vial master mix allows fast and convenient hot start qPCR without the need for upfront polymerase activation incubation. The mix is optimized for use with hydrolysis probes and does not require optimization of $MgCl_2$ concentration.

1.5. Preparation Time

Typical Run Time

The LightCycler® Multiplex DNA Master can be used for multiplex qPCR protocols. For example, a septaplex protocol using 45 cycles requires less than 75 minutes when using the LightCycler® PRO System.

2. How to Use this Product

2.1. Before you Begin

Sample Materials

Use any template DNA suitable for qPCR in terms of purity, concentration, and absence of PCR inhibitors.
For reproducible isolation of nucleic acids, use:
- Either a MagNA Pure System together with a dedicated reagent kit (for automated isolation),
- or a High Pure Nucleic Acid Isolation Kit (for manual isolation).

Control Reactions

Always run appropriate positive and negative controls with the samples.
- To check for the presence of contamination, prepare and include a negative control by replacing the template DNA with Water, PCR Grade (Vial 2).

Primers

Suitable concentrations of PCR primers range from 0.2 to 0.5 µM final concentration. The recommended starting concentration is 0.5 µM each.

Probe

Suitable concentrations of hydrolysis probes range from 0.05 to 0.5 µM (final concentration in PCR). The recommended starting concentration is 0.25 µM each.

The optimal probe concentration is the lowest concentration that results in the lowest quantification cycle value (Cq) and adequate fluorescence dynamics for a given target concentration.

For a hydrolysis probe hybridization complex, the Tm of the hydrolysis probe has to be higher than the Tm of the primers.

Mg^{2+} Concentration

The reaction mix in this kit already contains an optimal concentration of $MgCl_2$, which works with nearly all primer combinations.

You do not need to adjust the $MgCl_2$ concentration to amplify different sequences.

General Considerations

Precautions

Always use nuclease-free techniques. Nuclease contaminated reagents and reaction vessels will degrade template nucleic acids. Please follow these guidelines to minimize risk of contamination:
- Wear disposable gloves and change them frequently.
- Avoid touching surfaces or materials that could cause nuclease carryover.
- Use only reagents provided in this kit. Substitutions may introduce nucleases.
- Clean and decontaminate work areas and instruments, including pipettes, with commercially available decontamination reagents.
- Use only new nuclease-free aerosol-blocking pipette tips.
- Use a work area specifically designated for nucleic acid work, and if possible, use reaction vessels and pipettes dedicated only for work with template nucleic acid.

2.2. Protocols

LightCycler® PRO, LightCycler® 480, and LightCycler® 96 System protocols

The following procedure is optimized for use with the corresponding LightCycler® System you are using.

⚠ ***Program the LightCycler® Instrument before preparing the reaction mixes.***
For details on how to program the experimental protocol, see the LightCycler® PRO System User Assistance, LightCycler® 480 Instrument II Operator's Manual, or LightCycler® 96 System Operator's Guide.

Protocol for use with the LightCycler® PRO System (Multiwell Plate 96 or 384)

The LightCycler® PRO System protocol contains the following programs:
- **Pre-incubation** for denaturation of the DNA
- **Amplification** of the DNA
- **Cooling** of the thermal cycler

Setup				
Thermal cycler type			Reaction volume [µL]	
96 (384)			10 – 100 (5 – 20)	
Detection format				
Select dyes used in your assays. If the dye is not predefined, use appropriate filter combination in the user-defined detection format.				
Programs				
Program name		Cycles		
Pre-incubation		1		
Amplification		45[1]		
Cooling		1		
Temperature targets				
	Target [°C]	Acquisition mode	Duration [s]	Ramp rate [°C/s]
Pre-incubation	95	None	30	4.4 (4.8)
2-step amplification	95	None	5	4.4 (4.8)
	60[2]	Single	30	2.2 (2.5)
Cooling (automatically added)	40	None	30	2.2 (2.5)

[1] Forty-five cycles is suitable for most assays. If the assay shows steep amplification curves and early crossing points, 40 cycles should be sufficient. Reducing the number of cycles will improve uniformity of the product and reduce the time required for the assay.

[2] The LightCycler® Multiplex DNA Master includes an aptamer for hot start. Anneal/Extend temperatures significantly below 60°C may not be well tolerated because aptamers will inhibit DNA polymerase activity at low temperatures.

Color Compensation protocol for the LightCycler® PRO Instrument

The LightCycler® PRO Instrument does not require the creation of a color compensation object.

Protocol for use with the LightCycler® 480 Instrument II (Multiwell Plate 96 or 384)

The LightCycler® 480 Instrument II protocol contains the following programs:
- **Pre-incubation** for denaturation of the DNA
- **Amplification** of the DNA
- **Cooling** of the thermal cycler

Setup					
Thermal cycler type			Reaction volume [µL]		
96 (384)			20 (10)		
Detection format	Excitation filter		Emission filter		
For example: Mono Color Hydrolysis Probe / UPL Probe					
FAM	465		510		
Programs					
Program name	Cycles		Analysis mode		
Pre-incubation	1		None		
Amplification	45[1]		Quantification		
Cooling	1		None		
Temperature targets					
	Target [°C]	Acquisition mode	Hold [hh:mm:ss]	Ramp rate [°C/s]	Acquisitions [per °C]
Pre-incubation	95	None	00:00:30	4.4 (4.8)	–
Amplification	95	None	00:00:05	4.4 (4.8)	–
	60[2]	Single	00:00:30	2.2 (2.5)	–
Cooling	40	None	00:00:30	2.2 (2.5)	–

[1] Forty-five cycles is suitable for most assays. If the assay shows steep amplification curves and early crossing points, 40 cycles should be sufficient. Reducing the number of cycles will improve uniformity of the product and reduce the time required for the assay.

[2] The LightCycler® Multiplex DNA Master includes an aptamer for hot start. Anneal/Extend temperatures significantly below 60°C may not be well tolerated because aptamers will inhibit DNA polymerase activity at low temperatures.

Color Compensation protocol for the LightCycler® 480 Instrument II

For a multicolor, multiplex amplification, the application of a color compensation file is necessary to compensate for optical crosstalk between the different channels. For the LightCycler® 480 Instrument II, an instrument-specific color compensation file is mandatory for multicolor experiments, and a color compensation object can be generated by performing the following experiment.

The LightCycler® 480 Instrument II protocol contains the following programs:
- **Pre-incubation** for denaturation of the DNA
- **Amplification** of the DNA
- **Temperature gradient step** to create the Color Compensation file
- **Cooling** of the thermal block

For details on how to program the experimental protocol, see the LightCycler® 480 Instrument Operator's Manual, Version 1.5.

Programming a customized detection format for the LightCycler® 480 System filter combination selection

The Detection Format in the LightCycler® 480 Software, Version 1.5 setup must be customized for the applied multicolor hydrolysis probe format used in the qPCR detection. In the **Tool** module, the **Detection Formats** option allows creating a new detection format specified by the user, including a **Detection Format** list, a **Filter Combination** selection area, and a **Selected Filter Combination List**. Different filter settings for the LightCycler® 480 Instrument II are defined. The following table shows the qPCR parameters that must be programmed for a LightCycler® 480 Instrument II Color Compensation file with a LightCycler® 480 Multiwell Plate 96.

Setup					
Thermal cycler type			Reaction volume [µL]		
96			20		
Detection format	Excitation filter		Emission filter		
For example: 3 Color Hydrolysis Probe					
FAM Red 610 Cy5	465 533 618		510 610 660		
For new customized hydrolysis probes detection formats, set for all selected filters in the “Selected Filter Combination List” (under Tools), the following values:					
Melt factor	1				
Quant factor	10				
Max integration time (sec)	2				
Programs					
Program name	Cycles		Analysis mode		
Pre-incubation	1		None		
Amplification	45		Quantification		
Temperature gradient step	1		Color compensation		
Cooling	1		None		
Temperature targets					
	Target [°C]	Acquisition mode	Hold [hh:mm:ss]	Ramp rate [°C/s]	Acquisitions [per °C]
Pre-incubation	95	None	00:00:30	4.4	–
Amplification	95	None	00:00:05	4.4	–
	60	Single	00:00:30	4.4	–
Temperature gradient step	95	None	00:00:10	4.4	–
	40	None	00:00:10	2.2	–
	95	Continuous	–	–	1 – 5
Cooling	40	None	00:00:30	2.2	–

Preparation of the Color Compensation run

Prepare the calibrator qPCR mix for more than one reaction. Multiply the amount in the volume column by the number of reactions you need (minimum of 3 to 5 replicates) plus additional reactions since there will be a slight loss of liquid during the pipetting steps. In order to ensure accuracy, do not pipette volumes less than 1 µL when adding the individual reagents. For each dye, set up the following reactions:

Reagent	1x buffer [µL]	1x for each dye [µL]
qPCR Reaction Mix, 5x conc. (Vial 1)	4.0	4.0
Detection mix for each dye	–	X (depending on the assay)
Water, PCR Grade (Vial 2)	16.0	Y (depending on the assay)
Templates, such as DNA or positive samples eluates	–	5.0
Total volume	**20.0**	**20.0**

1. Pipette the replicates of each different calibrator mix into a LightCycler® 480 Multiwell Plate 96.

2. Seal the LightCycler® 480 Multiwell Plate using a sealing foil.

3. Place the multiwell plate into the centrifuge with a swinging-bucket rotor and suitable adapter, and balance it with a suitable counterweight, such as another multiwell plate.
   – Centrifuge at 1,500 × *g* for 2 minutes.

4. Load the multiwell plate into the LightCycler® 480 Instrument II and start the program.

Create Color Compensation object

When the experiment is finished, open the **Sample Editor**. Select workflow **Color Comp** and designate the used positions of the multiwell plate as “Water” for buffer replicates, and the appropriate dyes respectively (e.g., FAM, Red 610, Cy5 for the example mentioned above).
Open the **Analysis** module “Color Compensation”, click on **Calculate**, and save the color compensation object in the database.
From this point on, you can apply this “CC Object” to each dual-, triple- or quadruple-color experiment performed with FAM, Yellow 555, Red 610, and Cy5 on the same instrument.

Protocol for use with the LightCycler® 96 Instrument

Run editor				
Detection format			Reaction volume (µL)	
Selected dyes used in your assays. If using the DNA Process Control, select Cy5.			20	
Programs				
	Temp. [°C]	Ramp [°C/s]	Duration [s]	Acquisition mode
Pre-incubation	95	4.4	30	None
2-step amplification	No. of cycles: 45[1]			
	95	4.4	5	None
	60[2]	2.2	30	Single

[1] Forty-five cycles is suitable for most assays. If the assay shows steep amplification curves and early crossing points, 40 cycles should be sufficient. Reducing the number of cycles will improve uniformity of the product and reduce the time required for the assay.

[2] The LightCycler® Multiplex DNA Master includes an aptamer for hot start. Anneal/Extend temperatures significantly below 60°C may not be well tolerated because aptamers will inhibit DNA polymerase activity at low temperatures.

Color Compensation protocol for the LightCycler® 96 Instrument

The LightCycler® 96 Instrument does not require the creation of a color compensation object.

Setup of the qPCR reaction for the LightCycler® PRO, LightCycler® 480, and LightCycler® 96 Instruments

Follow the procedure below to prepare at least ten 20 µL standard reactions.
For 384-well plate setups use 10 µL standard reactions.
Always wear gloves during handling.

1. Thaw the solutions and, to ensure recovery of all the contents, briefly spin vials in a microcentrifuge before opening.
   – Mix carefully by pipetting up and down or vortex briefly.

2. Prepare a 20x conc. solution of your primers and a 20x conc. solution of your probes.

3. In a 1.5 mL reaction tube, prepare the qPCR Mix.
   – For best results, prepare at least 10 reactions in order to reduce pipetting errors. To prepare more reactions, multiply the amount in the "Volume 1 reaction" column below by the number of reactions to be run, plus at least one additional reaction.

Reagent[1]	Volume 1 reaction [µL]		Volume 10 reactions [µL]		Final conc.
	96-well plate	384-well plate	96-well plate	384-well plate	
Water, PCR Grade (Vial 2)	9.0	4.5	90.0	45.0	–
qPCR Reaction Mix, 5x conc. (Vial 1)	4.0	2.0	40.0	20.0	1x
Primer Mix, 20x conc.	1.0	0.5	10.0	5.0	1x
Probe Mix 20x conc.	1.0	0.5	10.0	5.0	1x
Total volume	**15.0**	**7.5**	**150.0**	**75.0**	

[1] For eluates derived from stool samples, it is recommended to add 0.2 µg/µL (final) of molecular biology-grade Bovine Serum Albumin*. Adjust for BSA volume by subtracting from PCR Water volume.

4. Mix carefully by pipetting up and down or vortex briefly.

5. Prepare the reagents right before use; the working solution (everything combined except DNA template) is stable at +15 to +25°C up to 4 hours, and is therefore ideal for use in automated workflows.

6. Prepare sample concentration of the DNA.

7. Pipette 15 µL (7.5 µL) qPCR Mix into a multiwell plate.
   – Add 5 µL (2.5 µL) of the DNA template.
   – Seal multiwell plate with a LightCycler® 480 Sealing Foil.

8. Place the multiwell plate into the centrifuge with a swinging-bucket rotor and suitable adapter, and balance it with a suitable counterweight, such as another multiwell plate.
   – Centrifuge at 1,500 × *g* for 2 minutes.

9. Load the multiwell plate into the LightCycler® PRO, LightCycler® 480, or LightCycler® 96 Instrument.

10. Start the PCR program described above.
    – If you use reaction volumes other than 20 µL, be sure to adapt the right volume in the running protocol. To start, we recommend using the same hold times as for the 20 µL volume.

Protocol for use with the LightCycler® 2.0 Instrument

⚠ ***Program the LightCycler® 2.0 Instrument before preparing the reaction mixes.***
ℹ *For details on how to program the experimental protocol, see the LightCycler® 2.0 Instrument Operator's Manual B.*

A LightCycler® 2.0 Instrument protocol that uses the LightCycler® Multiplex DNA Master contains the following programs:
- **Pre-incubation** for denaturation of the DNA
- **Amplification** of the DNA
- **Cooling** the rotor and thermal chamber

The following table shows the PCR parameters that must be programmed for a LightCycler® 2.0 Instrument PCR run with the LightCycler® Multiplex DNA Master using the LightCycler® Capillaries (20 µl).

LightCycler® Software Version 4.1				
Programs				
Setup[1]	Setting			
Default channel	Fluorescence channel			
Seek temperature	30°C			
Max seek pos.	Enter the total number of sample positions for which the instrument should look.			
Instrument type	"6 Ch." for LightCycler® 2.0 Instrument			
Capillary size	Select "20 µL" as the capillary size for the experiment.			
Programs				
Program name	Cycles	Analysis mode		
Pre-incubation	1	None		
Amplification	45[2]	Quantification		
Cooling	1	None		
Temperature targets				
	Target [°C]	Hold [hh:mm:ss]	Ramp rate [°C/s]	Acquisition mode [per °C]
Pre-incubation	95	00:00:30	20	None
Amplification	95	00:00:05	20	None
	60[3]	00:00:30	20	Single
Cooling	40	00:00:30	20	None

[1] The LightCycler® Multiplex DNA Master includes an aptamer for hot start. Anneal/Extend temperatures significantly below 60°C may not be well tolerated because aptamer will inhibit DNA polymerase activity at low temperatures.
[2] Forty-five cycles are suitable for most assays. If the assay shows steep amplification curves and early crossing points, 40 cycles should be sufficient. Reducing the number of cycles will improve uniformity of the product and reduce the time required for the assay.
[3] Most available assays are designed for an annealing temperature of +60°C. If the Tm of the primers and probe indicate using different settings, select an annealing temperature +5°C below the calculated primer Tm for initial experiments. For assay optimization, choose the highest annealing temperature which still results in a high yield of amplicon with a low Cq and adequate fluorescence dynamics.

Color Compensation protocol for the LightCycler® 2.0 Instrument

The following procedure is optimized for use with the LightCycler® 2.0 System. Program the LightCycler® 2.0 Instrument before preparing the reaction mixes. A LightCycler® 2.0 Instrument color compensation protocol that uses the LightCycler® Multiplex DNA Master contains the following programs:

- **Pre-incubation** for denaturation of the DNA
- **Amplification** of the DNA
- **Temperature gradient step** for Color Compensation
- **Cooling** the rotor and thermal chamber

For details on how to program the experimental protocol, see the LightCycler® 2.0 Instrument Operator's Manual B.

Color Compensation protocol

The performance of a color compensation is a prerequisite for running a dual-color experiment. The generated color compensation file is used to compensate for crosstalk between the individual detection channels when performing multi-color experiments. A color compensation calibration run is performed by running a blank capillary (containing Water, PCR Grade), and individual capillaries with one dye each (monocolor PCR reactions), in a qPCR program, followed by a color compensation analysis.

The following tables show the parameters that must be programmed for a LightCycler® Instrument Color Compensation calibration run with the LightCycler® Multiplex DNA Master.

LightCycler® Software Version 4.1				
Programs				
Setup		Setting		
Default channel		Fluorescence channel		
Seek temperature		30°C		
Max seek pos.		Enter the total number of sample positions for which the instrument should look.		
Instrument type		"6 Ch." for LightCycler® 2.0 Instrument		
Capillary size		Select "20 µL" as the capillary size for the experiment.		
Programs				
Program name		Cycles	Analysis mode	
Pre-incubation		1	None	
Amplification		45	Quantification	
Temperature gradient		1	Color compensation	
Cooling		1	None	
Temperature targets				
	Target [°C]	Hold [hh:mm:ss]	Ramp rate [°C/s]	Acquisition mode [per °C]
Pre-incubation	95	00:00:05	20	None
Amplification	95	00:00:01	20	None
	60	00:00:15	20	Single
Temperature gradient	95	00:00:01	20	None
	40	00:01:00	20	None
	95	00:00:00	0.2	Continuous
Cooling	40	00:00:30	20	None

Preparation of the Color Compensation mixes

⚠ ***Do not touch the surface of the LightCycler® Capillaries.***

1. Place three LightCycler® Capillaries (20 µl) into LightCycler® Centrifuge Adapters.

2. Prepare the capillaries (20 µL, each), as shown in Step 3.

3. For each dye, set up the following reactions:

Reagent	Volume for each dye [µL]	Capillary with water [µL]
qPCR Reaction Mix, 5x conc. (Vial 1)	4.0	–
Detection mix for each dye	X (depending on the assay)	–
Water, PCR Grade (Vial 2)	Y (depending on the assay)	20.0
Bovine Serum Albumin* (20 µg/µL)[1]	0.2	–
Template, such as DNA or positive sample eluates	5.0	–
Total volume	**20.0**	**20.0**

[1] Molecular biology-grade Bovine Serum Albumin* is recommended.

4. Seal each capillary with a stopper using the LightCycler® Capping Tool.

5. Place the centrifuge adapters (containing the capillaries) into a standard benchtop microcentrifuge.
   – Place the centrifuge adapters in a balanced arrangement within the centrifuge.
   – Centrifuge at 700 × *g* for 5 seconds (3,000 rpm in a standard benchtop microcentrifuge). Alternatively, use the LC Carousel Centrifuge for spinning the capillaries.

6. Place the capillaries in the following order in the sample carousel of the LightCycler® 2.0 Instrument:
   Carousel rotor position 1: Water
   Carousel rotor position 2: Monocolor PCR for Dye 1
   Carousel rotor position 3: Monocolor PCR for Dye 2

7. Cycle the samples as described above and edit the dominant channel in the “Analysis Type” - “Color Comp” accordingly.

Create Color Compensation objects

When the experiment is finished, click on the **Analysis** button and select **Color Compensation** (Other Methods) from the **Analysis** Menu. Save the experiment by clicking the **Save CC Object** button. Place the object in the “Special Data\CCC” folder under your user name.
After doing this, you can apply the specific “CC Object” you created to any dual-color hydrolysis probe experiment that is performed with the same dye combination.

Setup of the qPCR reaction for the LightCycler® 2.0 Instrument

This setup can also be used in a qPCR protocol for the LightCycler® 1.x Instrument.
Follow the procedure below to prepare at least ten 20 µL standard reactions:
Do not touch the surface of the LightCycler® Capillaries.

1. Thaw the solutions and, to ensure recovery of all the contents, briefly spin vials in a microcentrifuge before opening.
   – Mix carefully by pipetting up and down or vortex briefly.
   – Place samples on ice.

2. Prepare a 20x conc. solution of your primers and a 20x conc. solution of your probes.

3. In a 1.5 mL reaction tube, prepare the PCR Mix.
   – For best results, prepare at least 10 reactions in order to reduce pipetting errors. To prepare more reactions, multiply the amount in the "Volume 1 reaction" column below by the number of reactions to be run, plus at least one additional reaction.

Reagent	Volume 1 reaction [µL]	Volume 10 reactions [µL]	Final conc.
Water, PCR Grade (Vial 2)	8.8	88.0	–
qPCR Reaction Mix, 5x conc. (Vial 1)	4.0	40.0	1x
Primer Mix, 20x conc.(1)	1.0	10.0	1x
Probe Mix, 20x conc.	1.0	10.0	1x
Bovine Serum Albumin* (20 µg/µL)(2)	0.2	2.0	0.2 µg/µL
Total volume	**15.0**	**150.0**	

(1) Due to possible primer/primer interactions that occur during storage, it may be necessary to preheat the PCR primer mix for 1 minute at +95°C before pipetting the full mixture. This extra step will ensure optimum sensitivity.
(2) Molecular biology-grade Bovine Serum Albumin* is recommended.

4. Mix carefully by pipetting up and down or vortex briefly.

5. Prepare the reagents right before use; the working solution (everything combined except DNA template) is stable at +15 to +25°C up to 4 hours, and is therefore ideal for use in automated workflows.

6. Prepare sample concentration of the DNA.

7. Pipette 15 µL PCR Mix into a LightCycler® Capillary.
   – Add 5 µL of the DNA template.

8. Seal the LightCycler® Capillaries with a stopper using the LightCycler® Capping Tool.

9. If using the LC Carousel Centrifuge 2.0, proceed to Step 10.

10. Alternatively, place the capillaries in cooled adapters in a standard benchtop microcentrifuge in a balanced arrangement.
    – Centrifuge at 700 × *g* (3,000 rpm) for 5 seconds, and transfer the capillaries to the LightCycler® Sample Carousel.

11. Place the LightCycler® Sample Carousel in the LightCycler® Carousel-Based Instrument.
    – Start the program.

3. Results

Results on the LightCycler® PRO Instrument

The following results were obtained using the LightCycler® Multiplex DNA Master on the LightCycler® PRO Instrument.
A triplex reaction using a **gyrB**-specific assay (Cyan 500), **ipaH**-specific assay (HEX) and a **gyrB/dnaJ**-specific assay (Red 640). Target-specific plasmids served as templates.

All targets:

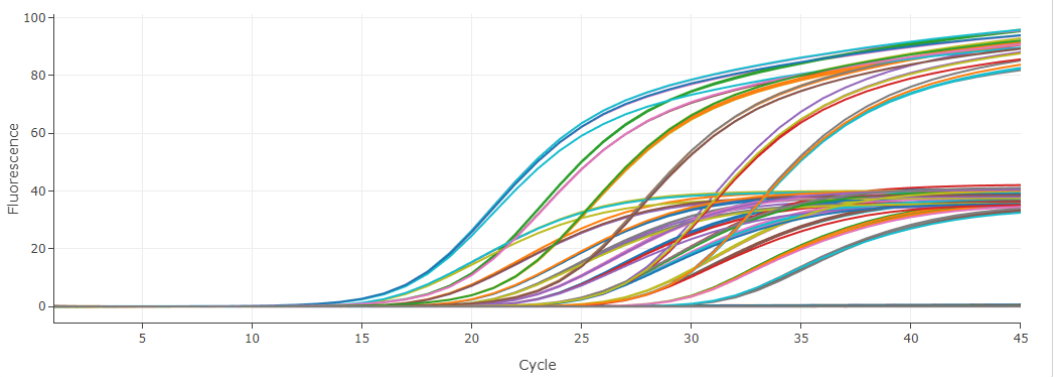

Fig. 1: Triple qPCR amplification reaction on the LightCycler® PRO Instrument within one picture.

The following three images show the individual amplification curves of the respective detection channels.

Cyan 500 channel (432 – 474)

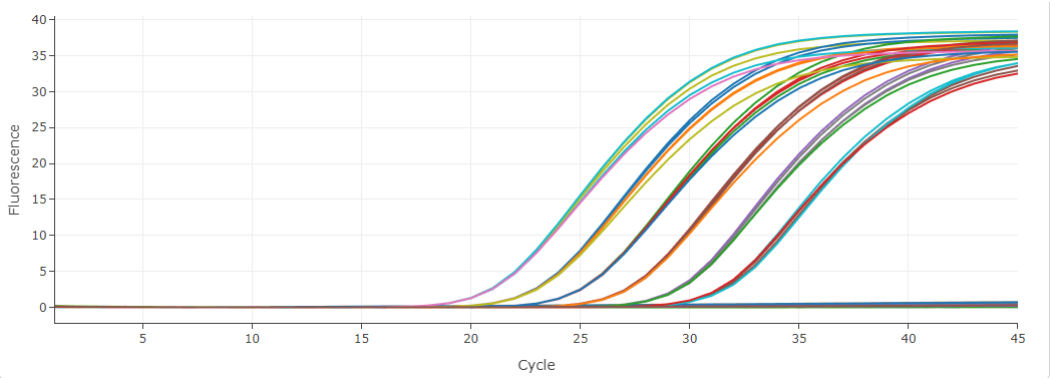

Fig. 2: The Cyan 500 channel shows the result for an amplification of a 133 bp fragment of the **gyrB** gene detected with a Cyan 500-labeled hydrolysis probe. Amplification curves shown were generated from a six-fold 1:4 dilution series [10,000,000 copies (far left), 2,500,000 copies, 625,000 copies,156,250 copies, 39,063 copies, and 9,766 (far right)] from a plasmid for ***Yersinia Enterocolitica***, 5 replicates per concentration. No template controls are shown as a flat line.
Triplex qPCR was performed in a reaction volume of 20 µL per well in a LightCycler® 480 Multiwell Plate 96.

HEX channel (541 – 565)

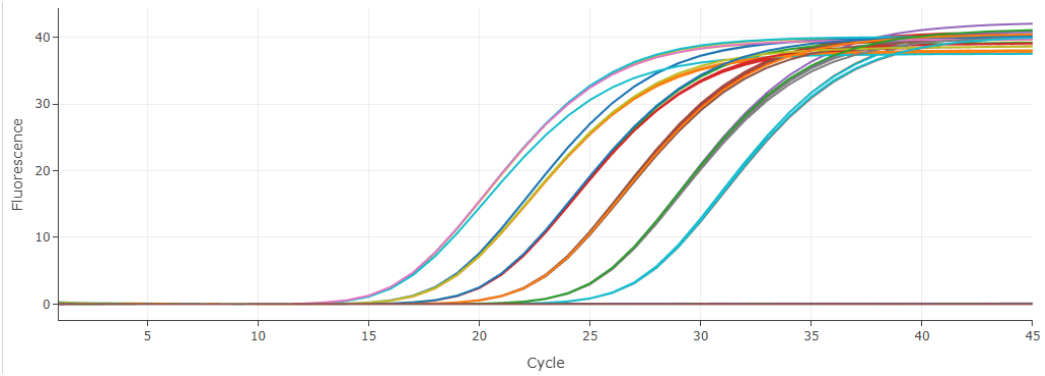

Fig. 3: The HEX channel shows the result for an amplification of a 111 bp fragment of the **ipaH** gene detected with a R6G-labeled hydrolysis probe. Amplification curves shown were generated from a six-fold 1:4 dilution series [10,000,000 copies (far left), 2,500,000 copies, 625,000 copies,156,250 copies, 39,063 copies, and 9,766 (far right)] from a plasmid for ***Shigella/EIEC***, 5 replicates per concentration. No template controls are shown as a flat line.
Triplex qPCR was performed in a reaction volume of 20 µL per well in a LightCycler® 480 Multiwell Plate 96.

Red 640 channel (621 – 636)

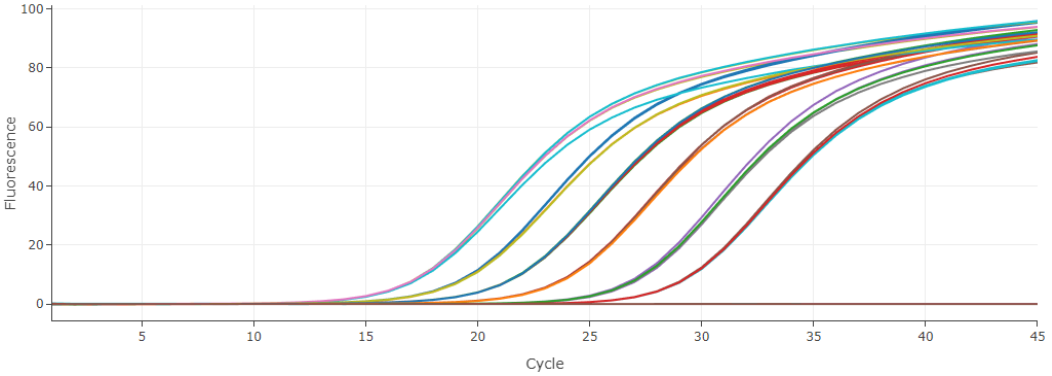

Fig. 4: The Red 640 channel shows the result for the dual target amplification approach of a 86 bp fragment of the **gyrB** gene and a 84 bp fragment of the **dnaJ** gene detected with a Red 640-labeled hydrolysis probe. Amplification curves shown were generated from a six-fold 1:4 dilution series [10,000,000 copies (far left), 2,500,000 copies, 625,000 copies,156,250 copies, 39,063 copies, and 9,766 (far right)] from a plasmid for ***Aeromonas***, 5 replicates per concentration. No template controls are shown as a flat line.
Triplex qPCR was performed in a reaction volume of 20 µL per well in a LightCycler® 480 Multiwell Plate 96.

Results on the LightCycler® 480 Instrument II

The following results were obtained using the LightCycler® Multiplex DNA Master on the LightCycler® 480 Instrument II. A duplex reaction using a **β-globin**-specific assay with BHQ-2 internally quenched probe (FAM) and a **β2M**-specific assay with UPL probe (Yellow 555). Human reference cDNA served as the template.

FAM Channel (465 - 510)

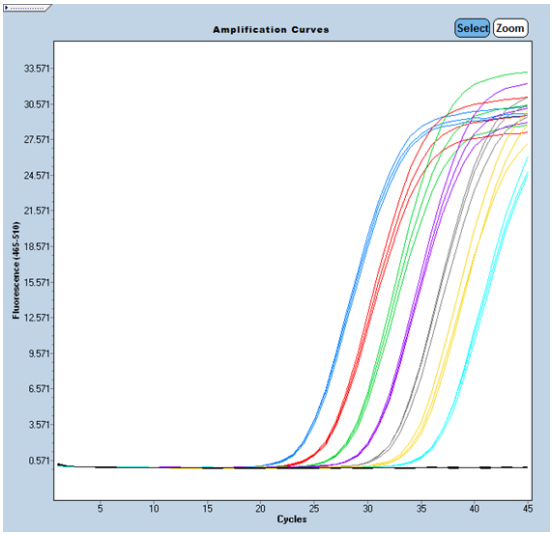

Fig. 5: The FAM channel shows the results for **β-globin**. Amplification curves shown were generated from a four-fold dilution series [10 ng (far left), 2.5 ng, 625 pg, 156 pg, 40 pg, 10 pg, and 2.4 pg (far right)] of Human Reference cDNA. No template controls are shown in black (flat line). Duplex qPCR was performed in a reaction volume of 20 µL per well in a LightCycler® 480 Multiwell Plate 96.

Yellow 555 Channel (533 - 580)

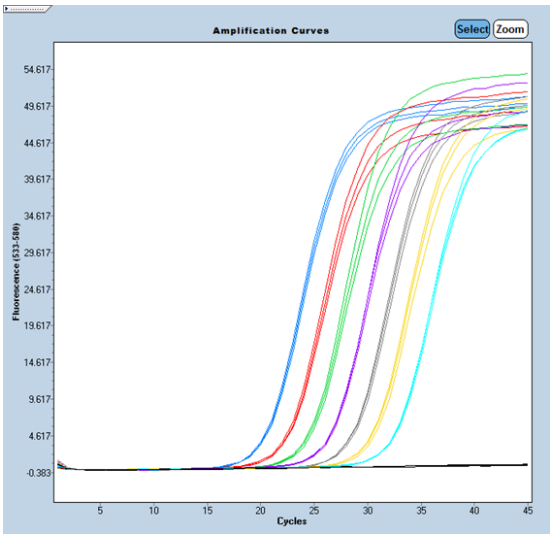

Fig. 6: The Yellow 555 channel shows the results for **β2M**. Amplification curves shown were generated from a four-fold dilution series [10 ng (far left), 2.5 ng, 625 pg, 156 pg, 39 pg, 10 pg, and 2.4 pg (far right)] of Human Reference cDNA. No template controls are shown in black (flat line). Duplex qPCR was performed in a reaction volume of 20 µL per well in a LightCycler® 480 Multiwell Plate 96.

4. Troubleshooting

Observation	Possible cause	Recommendation
Fluorescence intensity varies.	Some of the reagent is still in the upper part of the microwell or an air bubble is trapped in the microwell.	Repeat centrifugation, but allow sufficient centrifugation time so all reagent is at the bottom of the microwell and air bubbles are expelled.
	Skin oils or dirt are present on the lid or sealing foil.	Always wear gloves when handling the multiwell plate/8-tube strip.
Fluorescence intensity is very low.	Low concentration or deterioration of dyes in the reaction mixtures because dye was not stored properly.	Keep dye-labeled reagents protected from light.
		Store the reagents at −15 to −25°C and avoid repeated freezing and thawing.
	Poor PCR efficiency (reaction conditions not optimized).	Primer concentration should be in the range of 0.2 to 0.5 µM; probe concentration should be in the range of 0.2 to 0.5 µM and half of the primer concentration.
		Check annealing temperature of primers and probes.
		Check experimental protocol.
		Optimize annealing temperature in the reverse transcription step or in the PCR reaction.
		Always run a positive control along with your samples.
	Chosen integration time is too low.	Choose the appropriate detection format in combination with “dynamic” detection mode, or
		Increase integration time when using “manual” detection mode. *For details, see the LightCycler® 480 Instrument Operator’s Manual.*
	qPCR primers and probes are not optimized.	Check sequence and location of the hydrolysis probe on the PCR product.
		Check PCR product on an agarose gel.
	PCR has not been optimized.	Check primer design (quality).
		Check PCR product on an agarose gel.
	DNA is degraded during isolation or improper storage.	If possible, check DNA quality.
	Pipetting errors or omitted reagents.	Check for missing reagents.
		Check the pipetting procedure.
	Impure sample material inhibits reaction.	Dilute sample 1:10 and repeat the analysis.
		Repurify the nucleic acids to ensure removal of inhibitory agents.
Negative control sample gives a positive signal.	Contamination	Remake all critical reaction mixes. Be sure to use special pre-PCR setup working areas.

5. Additional Information on this Product

5.1. Quality Control

The LightCycler® Multiplex DNA Master is function tested using the LightCycler® System.

6. Supplementary Information

6.1. Conventions

To make information consistent and easier to read, the following text conventions and symbols are used in this document to highlight important information:

Text convention and symbols	
Information Note: Additional information about the current topic or procedure.	
Important Note: Information critical to the success of the current procedure or use of the product.	
① ② ③ etc.	Stages in a process that usually occur in the order listed.
❶ ❷ ❸ etc.	Steps in a procedure that must be performed in the order listed.
* (Asterisk)	The Asterisk denotes a product available from Roche Diagnostics.

6.2. Changes to previous version

Editorial changes.
Information about the LightCycler® PRO System has been added.
List of additional reagents and equipment has been updated.
Quality Control has been changed to LightCycler® System.

6.3. Ordering Information

Roche offers a large selection of reagents and systems for life science research. For a full overview of related products and manuals, please visit and bookmark our homepage lifescience.roche.com.

Product	Pack Size	Cat. No.
Accessories general (hardware)		
LightCycler® 480 Block Kit 384 Silver	1 block kit	05 015 197 001
LightCycler® 480 Block Kit 96 Silver	1 block kit	05 015 219 001
LightCycler® 8-Tube Strip Adapter Plate	1 piece, adapter plate The adapter plate can be used multiple times	06 612 598 001
Thermal Cycler Assembly 96	1 piece	09 742 565 001
Thermal Cycler Assembly 384	1 piece	09 742 581 001
Sealing Foil Applicator	1 piece	10 018 607 001
Consumables		
LightCycler® 480 Multiwell Plate 96, white	5 x 10 plates	04 729 692 001
LightCycler® 8-Tube Strips (white)	10 x 12 white strips and clear caps	06 612 601 001
LightCycler® Capillaries (20 µl)	5 × 96 capillaries, containing 5 boxes, each with 96 capillaries and stoppers	04 929 292 001
LightCycler® 480 Sealing Foil	50 foils	04 729 757 001
MagNA Pure 96 Internal Control Tube	150 tubes	06 374 905 001
LightCycler® 480 Multiwell Plate 384, white	5 x 10 plates	04 729 749 001
LightCycler® 480 Multiwell Plate 96	5 x 10 plates without sealing foils	05 220 319 001
LightCycler® 480 Multiwell Plate 384	5 x 10 plates without sealing foils	05 217 555 001
Instruments		
LightCycler® 96 Instrument	1 instrument	05 815 916 001
MagNA Pure 96 Instrument	1 instrument	06 541 089 001
LightCycler® 480 Instrument II	1 instrument, 96-well version	05 015 278 001
	1 instrument, 384-well version	05 015 243 001
LightCycler® PRO Instrument	1 instrument, 96-well version	09 541 713 001
	1 instrument, 384-well version	09 582 487 001
MagNA Pure 24 Instrument	Instrument with built-in control unit, software and accessories	07 290 519 001
Reagents, kits		
High Pure Viral Nucleic Acid Kit	1 kit, up to 100 isolations	11 858 874 001
MagNA Pure 96 DNA and Viral NA Small Volume Kit	For up to 3 x 192 isolations	06 543 588 001
Bovine Serum Albumin	20 mg, 1 ml	10 711 454 001
MagNA Pure 96 DNA and Viral NA Large Volume Kit	Kit for up to 3 x 96 isolations	06 374 891 001
MagNA Pure 24 Total NA Isolation Kit	Kit for up to 96 isolations (200 µL)	07 658 036 001

6.4. Trademarks

MAGNA PURE, APTATAQ and LIGHTCYCLER are trademarks of Roche.
All other product names and trademarks are the property of their respective owners.

6.5. License Disclaimer

For additional documentation such as certificates and safety data sheets, please visit:
documentation.roche.com.

6.6. Regulatory Disclaimer

For general laboratory use.

6.7. Safety Data Sheet

Please follow the instructions in the Safety Data Sheet (SDS).

6.8. Contact and Support

If you have questions or experience problems with this or any Roche product for Life Science, please contact our Technical Support staff. Our scientists are committed to providing rapid and effective help.
Please also contact us if you have suggestions for enhancing Roche product performance or using our products in new or specialized ways. Such customer information has repeatedly proven invaluable to the research community worldwide.
To ask questions, solve problems, suggest enhancements or report new applications,
please visit our **Online Technical Support** Site.

Visit **documentation.roche.com**, to download or request copies of the following Materials:
- Instructions for Use
- Safety Data Sheets
- Certificates of Analysis
- Information Material

To call, write, fax, or email us, visit **lifescience.roche.com** and select your home country to display country-specific contact information.

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