200



REF

06437206 190

English

System information

For **cobas e** 411 analyzer: test number 1240 For MODULAR ANALYTICS E170, **cobas e** 601 and **cobas e** 602 analyzers: Application Code Number 195

Intended use

Immunoassay for the in vitro quantitative determination of free triiodothyronine in human serum and plasma.

The electrochemiluminescence immunoassay "ECLIA" is intended for use on Elecsys and cobas e immunoassay analyzers.

Summary

The thyroid hormones triiodothyronine (T3) and thyroxine (T4) are secreted into the bloodstream by the thyroid gland and play a vital role in regulating the body's metabolic rate, influencing the cardiovascular system, growth and bone metabolism, and are important for normal development of gonadal functions and nervous system.¹

T3 circulates in the bloodstream as an equilibrium mixture of free and serum bound hormone. Free T3 (fT3) is the unbound and biologically active form, which represents only 0.2-0.4 % of the total T3. The remaining T3 is inactive and bound to serum proteins, while the distribution of T3 between these binding proteins (thyroxine binding globulin, pre-albumin, albumin) is controversially discussed.^{2,3,4,5}

The determination of free T3 has the advantage of being independent of changes in the concentrations and binding properties of the binding proteins; additional determination of a binding parameter (T-uptake, TBG) is therefore unnecessary. Therefore free T3 is a useful tool in clinical routine diagnostics for the assessment of the thyroid status. Free T3 measurements support the differential diagnosis of thyroid disorders, are needed to distinguish different forms of hyperthyroidism, and to identify patients with T3 thyrotoxicosis.^{1,6,7}

A variety of methods are available for estimating the free thyroid hormone levels. The direct measurement of fT4 and fT3 via equilibrium dialysis or ultrafiltration is mainly used as a reference method for standardizing the immunological procedures generally used for routine diagnostic purposes.^{6,7}

In the Elecsys FT3 III assay a specific anti-T3 antibody labeled with a ruthenium complex $\!$ and to determine the free triiodothyronine concentration.

a) Tris(2,2'-bipyridyl)ruthenium(II)-complex (Ru(bpy)_3^{2+})

Test principle

Competition principle. Total duration of assay: 18 minutes.

- 1st incubation: 15 µL of sample and an anti-T3-specific antibody labeled with a ruthenium complex.
- 2nd incubation: After addition of biotinylated T3 and streptavidin-coated microparticles, the still-free binding sites of the labeled antibody become occupied, with formation of an antibody-hapten complex. The entire complex is bound to the solid phase via interaction of biotin and streptavidin.
- The reaction mixture is aspirated into the measuring cell where the microparticles are magnetically captured onto the surface of the electrode. Unbound substances are then removed with ProCell/ProCell M. Application of a voltage to the electrode then induces chemiluminescent emission which is measured by a photomultiplier.
- Results are determined via a calibration curve which is instrumentspecifically generated by 2-point calibration and a master curve provided via the reagent barcode or e-barcode.

Reagents - working solutions

The reagent rackpack is labeled as FT3 III.

SYSTEM

MODULAR ANALYTICS E170 cobas e 411 cobas e 601 cobas e 602

- M Streptavidin-coated microparticles (transparent cap), 1 bottle, 12 mL: Streptavidin-coated microparticles 0.72 mg/mL; preservative.
- R1 Anti-T3-Ab~Ru(bpy)₃²⁺ (gray cap), 1 bottle, 18 mL:

Monoclonal anti-T3-antibody (sheep) labeled with ruthenium complex 18 ng/mL; phosphate buffer 100 mmol/L, pH 7.0; preservative.

R2 T3~biotin (black cap), 1 bottle, 18 mL:

Biotinylated T3 2.4 ng/mL; phosphate buffer 100 mmol/L, pH 7.0; preservative.

Precautions and warnings

For in vitro diagnostic use.

Exercise the normal precautions required for handling all laboratory reagents.

Disposal of all waste material should be in accordance with local guidelines. Safety data sheet available for professional user on request.

Avoid foam formation in all reagents and sample types (specimens, calibrators and controls).

Reagent handling

The reagents in the kit have been assembled into a ready-for-use unit that cannot be separated.

All information required for correct operation is read in from the respective reagent barcodes.

Storage and stability

Store at 2-8 °C.

Do not freeze.

Store the Elecsys reagent kit **upright** in order to ensure complete availability of the microparticles during automatic mixing prior to use.

Stability:

unopened at 2-8 °C	up to the stated expiration date	
after opening at 2-8 °C	84 days (12 weeks)	
on the analyzers	42 days (6 weeks) onboard or	
	56 days (8 weeks) when stored alternatively in the refrigerator and on the analyzer, with the total time onboard on the analyzer not exceeding 120 hours	

Specimen collection and preparation

Only the specimens listed below were tested and found acceptable. Undiluted serum collected using standard sampling tubes or tubes containing separating gel.

Undiluted Li-heparin, K₂-EDTA and K₃-EDTA plasma.

Recovery with a total deviation $\le \pm 0.4$ pmol/L of initial value at concentrations < 2 pmol/L; recovery within ± 10 % of initial value at concentrations ≥ 2 pmol/L and slope 0.9-1.1 + intercept within $\le \pm 2x$ Limit of Blank + coefficient of correlation ≥ 0.95 .

Stable for 7 days at 2-8 $^{\circ}\text{C},$ 30 days at -20 $^{\circ}\text{C}.^{6}$ Freeze only once.

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary

cobas®

tubes (sample collection systems), follow the instructions of the tube manufacturer.

Centrifuge samples containing precipitates before performing the assay. Do not use heat-inactivated samples.

Do not use samples and controls stabilized with azide.

Ensure the samples, calibrators and controls are at 20-25 $^\circ\mathrm{C}$ prior to measurement.

Due to possible evaporation effects, samples, calibrators and controls on the analyzers should be analyzed/measured within 2 hours.

Materials provided

See "Reagents - working solutions" section for reagents.

Materials required (but not provided)

- REF 06437222190, FT3 III CalSet, for 4 x 1.0 mL
- REF 11731416190, PreciControl Universal, for 4 x 3.0 mL
- General laboratory equipment
- MODULAR ANALYTICS E170 or cobas e analyzer

Accessories for cobas e 411 analyzer:

- [REF] 11662988122, ProCell, 6 x 380 mL system buffer
- REF 11662970122, CleanCell, 6 x 380 mL measuring cell cleaning solution
- REF 11930346122, Elecsys SysWash, 1 x 500 mL washwater additive
- REF 11933159001, Adapter for SysClean
- REF 11706802001, AssayCup, 60 x 60 reaction cups
- REF 11706799001, AssayTip, 30 x 120 pipette tips
- REF 11800507001, Clean-Liner

Accessories for MODULAR ANALYTICS E170, **cobas e** 601 and **cobas e** 602 analyzers:

- REF 04880340190, ProCell M, 2 x 2 L system buffer
- REF 04880293190, CleanCell M, 2 x 2 L measuring cell cleaning solution
- REF 03023141001, PC/CC-Cups, 12 cups to prewarm ProCell M and CleanCell M before use
- [REF] 03005712190, ProbeWash M, 12 x 70 mL cleaning solution for run finalization and rinsing during reagent change
- REF 03004899190, PreClean M, 5 x 600 mL detection cleaning solution
- REF 12102137001, AssayTip/AssayCup, 48 magazines x 84 reaction
- cups or pipette tips, waste bags
- REF 03023150001, WasteLiner, waste bags
- REF 03027651001, SysClean Adapter M

Accessories for all analyzers:

International International International International International International International Internation
International Intern

Assay

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.

Resuspension of the microparticles takes place automatically prior to use. Read in the test-specific parameters via the reagent barcode. If in exceptional cases the barcode cannot be read, enter the 15-digit sequence of numbers (except for the **cobas e** 602 analyzer).

MODULAR ANALYTICS E170, cobas e 601 and cobas e 602 analyzers: PreClean M solution is necessary.

Bring the cooled reagents to approximately 20 °C and place on the reagent disk (20 °C) of the analyzer. Avoid foam formation. The system automatically regulates the temperature of the reagents and the opening/closing of the bottles.

Calibration

Traceability: This method has been standardized against the Elecsys FT3 assay ($[\mbox{REF}]$ 03051986190). The Elecsys FT3 assay ($[\mbox{REF}]$ 03051986190) is traceable to the Elecsys FT3 assay ($[\mbox{REF}]$ 11731386122) which was standardized using equilibrium dialysis. 5,8

Every Elecsys reagent set has a barcoded label containing specific information for calibration of the particular reagent lot. The predefined master curve is adapted to the analyzer using the relevant CalSet.

Calibration frequency: Calibration must be performed once per reagent lot using fresh reagent (i.e. not more than 24 hours since the reagent kit was registered on the analyzer).

Calibration interval may be extended based on acceptable verification of calibration by the laboratory.

Renewed calibration is recommended as follows:

- after 1 month (28 days) when using the same reagent lot
- after 7 days (when using the same reagent kit on the analyzer)
- as required: e.g. quality control findings outside the defined limits

Quality control

For quality control, use PreciControl Universal.

In addition, other suitable control material can be used.

Controls for the various concentration ranges should be run individually at least once every 24 hours when the test is in use, once per reagent kit, and following each calibration.

The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

If necessary, repeat the measurement of the samples concerned. Follow the applicable government regulations and local guidelines for quality control.

Calculation

The analyzer automatically calculates the analyte concentration of each sample (either in pmol/L, pg/mL or ng/dL).

Conversion factors:

pmol/L x 0.651 = pg/mL pg/mL x 1.536 = pmol/L pg/mL x 0.1 = ng/dL

Limitations - interference

The assay is unaffected by icterus (bilirubin < 1128 μ mol/L or < 66 mg/dL), hemolysis (Hb < 0.621 mmol/L or < 1.0 g/dL), lipemia (Intralipid < 2000 mg/dL), biotin (< 286 nmol/L or < 70 ng/mL), IgG < 7 g/dL, IgA < 1.6 g/dL and IgM < 1 g/dL.

Criterion: Recovery within ± 10 % of initial value.

Samples should not be taken from patients receiving therapy with high biotin doses (i.e. > 5 mg/day) until at least 8 hours following the last biotin administration.

No interference was observed from rheumatoid factors up to a concentration of 1200 $\mbox{IU/mL}.$

Any influence that might affect the binding behavior of the binding proteins can alter the result of the fT3 tests (e.g. drugs, NTIs (Non-Thyroid-Illness) or patients suffering from FDH (Familial Dysalbuminemic Hyperthyroxinemia)).^{9,10}

In vitro tests were performed on 17 commonly used pharmaceuticals. No interference with the assay was found.

The following special thyroid drugs were tested with concentrations shown in the table below. No interference with the assay was found. Criterion: Recovery within \pm 10 % of initial value.

Drug	Concentration (µg/mL)
lodid	0.200
Carbimazol	30
Thiamazol	80
Propylthiouracil	60
Perchlorat	2000
Propranolol	240
Amiodaron	200
Prednisolon	100

Drug	Concentration (µg/mL)
Hydrocortison	200
Fluocortolon	100
Octreotid	0.300

In in vitro studies the drugs Furosemide and Levothyroxine caused elevated fT3 findings at the daily therapeutic dosage level.

In rare cases, interference due to extremely high titers of antibodies to analyte-specific antibodies, streptavidin or ruthenium can occur. These effects are minimized by suitable test design.

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

Limits and ranges

Measuring range

0.4-50 pmol/L (defined by the Limit of Blank and the maximum of the master curve). Values below the Limit of Blank are reported as < 0.4 pmol/L. Values above the measuring range are reported as > 50 pmol/L.

Lower limits of measurement

Limit of Blank, Limit of Detection and Limit of Quantitation

Limit of Blank = 0.4 pmol/L

Limit of Detection = 0.6 pmol/L

Limit of Quantitation = 1.5 pmol/L

The Limit of Blank, Limit of Detection and Limit of Quantitation were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A requirements.

The Limit of Blank is the 95th percentile value from n \ge 60 measurements of analyte-free samples over several independent series. The Limit of Blank corresponds to the concentration below which analyte-free samples are found with a probability of 95 %.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low concentration samples. The Limit of Detection corresponds to the lowest analyte concentration which can be detected (value above the Limit of Blank with a probability of 95 %).

The Limit of Quantitation is defined as the lowest amount of analyte in a sample that can be accurately quantitated with a total allowable error of \leq 30 %.

Dilution

Samples for fT3 determinations cannot be diluted, as T3 in the blood is present in free and protein-bound forms which are in equilibrium. A change in the concentration of the binding proteins alters this equilibrium.

Expected values

Euthyroid: 3.1-6.8 pmol/L (2.0-4.4 pg/mL)

These values correspond to the 2.5^{th} and 97.5^{th} percentiles of results obtained from a total of 5366 healthy test subjects examined.

For detailed information about reference intervals in children, adolescents and pregnant women, refer to the brochure "Reference Intervals for Children and Adults", English: [REF] 04640292.

This booklet also contains results of a detailed study about influencing factors on thyroid parameters in a well characterized reference group of adults. Different inclusion and exclusion criteria were applied (e.g. sonographic results (thyroid volume and density) as well as criteria according to the guidelines of the National Academy of Clinical Biochemistry - NACB).

Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

Specific performance data

Representative performance data on the analyzers are given below. Results obtained in individual laboratories may differ.

Precision

Precision was determined using Elecsys reagents, samples and controls in a protocol (EP5-A2) of the CLSI (Clinical and Laboratory Standards Institute): 2 runs per day in duplicate each for 21 days (n = 84). The following results were obtained:

cobas e 411 analyzer					
		Repeatability		Interme precis	diate ion
Sample	Mean pmol/L	SD pmol/L	CV %	SD pmol/L	CV %
HS ^{b)} 1	1.53	0.045	3.0	0.125	8.2
HS 2	3.64	0.083	2.3	0.133	3.6
HS 3	6.43	0.129	2.0	0.199	3.1
HS 4	27.5	0.657	2.4	0.959	3.5
HS 5	46.8	0.495	1.1	0.884	1.9
PC U ^{c)} 1	5.92	0.077	1.3	0.150	2.5
PC U2	24.1	0.492	2.0	0.713	3.0

b) HS = human serum

c) PC U = PreciControl Universal

MODULAR ANALYTICS E170, cobas e 601 and cobas e 602 analyzers		
	Repeatability	Intermediate

		repeatability		precis	ion
Sample	Mean pmol/L	SD pmol/L	CV %	SD pmol/L	CV %
HS 1	1.33	0.086	6.5	0.096	7.2
HS 2	3.49	0.107	3.1	0.113	3.2
HS 3	6.24	0.121	1.9	0.134	2.1
HS 4	27.3	0.367	1.3	0.442	1.6
HS 5	46.0	0.640	1.4	0.855	1.9
PC U1	5.67	0.121	2.1	0.131	2.3
PC U2	23.4	0.349	1.5	0.455	1.9

Method comparison

A comparison of the Elecsys FT3 III assay (y) with the Elecsys FT3 assay (x) using clinical samples gave the following correlations: Number of samples measured: 155

Passing/Bablok ¹¹	Linear regression
y = 1.05x - 0.233	y = 1.04x - 0.107
r = 0.943	r = 0.998

The sample concentrations were between 0.777 and 47.8 pmol/L.

Analytical specificity

The following cross-reactivities were found, tested with fT3 concentrations of approximately 4.61 pmol/L (3.00 pg/mL) and 11.4 pmol/L (7.44 pg/mL):

1

Cross-reactant	Concentration tested pg/mL	Cross-reactivity %
L-T4	300000	0.009
D-T4	625000	0.005
rT3	1000000	0.0003
3-iodo-L-tyrosine	10000000	0.000
3,5-diiodo-L-tyrosine	10000000	0.000
3,3',5-triiodothyroacetic acid	6250	0.298
3,3',5,5'-tetraiodothyroacetic acid	1000000	0.0001

References

Kronenberg HM, Melmed S, Polonsky KS, et al. Williams Textbook of Endocrinology. Saunders Elsevier, Philadelphia, 12th edition, 2011, chapter 10, p. 301-311.

cobas®

cobas®

- 2 Robbins J, Rall JE. The interaction of thyroid hormones and protein in biological fluids. Recent Prog Horm Res 1957;13:161-208.
- 3 Oppenheimer JH. Role of plasma proteins in the binding, distribution and metabolism of the thyroid hormones. N Engl J Med 1968;278(21):1153-1162.
- 4 DeGroot LJ, Larsen PR, Hennemann G. Transport of thyroid hormone and cell uptake. The thyroid and its diseases. Wiley and Sons, New York, 1984:62-65.
- 5 Ekins RP. Measurement of free hormones in blood. Endocr Rev 1990;11(1):5-46.
- 6 Wu AHB. Tietz Clinical Guide To Laboratory Tests. Saunders Elsevier, Philadelphia, 4th edition, 2006, section II, p. 1076-1077.
- 7 Brent GA. Thyroid Function Testing. Springer, Berlin, 1st edition, 2010, chapter 5, p. 86-88.
- 8 Ekins RP, Ellis SM. The radioimmunoassay of free thyroid hormones in serum. In Robbins J, Braverman LE (eds). Thyroid research, Proceedings of the Seventh International Thyroid Conference, Boston. Amsterdam, Excerpta Medica 1975:597.
- 9 Wada N, Chiba H, Shimizu C, et al. A novel missense mutation in codon 218 of the albumin gene in a distinct phenotype of familial dysalbuminemic hyperthyroxinemia in a Japanese kindred. J Clin Endocrinol Metab 1997;82(10):3246-3250.
- Arevalo G. Prevalence of familial dysalbuminemic hyperthyroxinemia in serum samples received for thyroid testing. Clin Chem 1991;37(8):1430-1431.
- 11 Bablok W, Passing H, Bender R, et al. A general regression procedure for method transformation. Application of linear regression procedures for method comparison studies in clinical chemistry, Part III. J Clin Chem Clin Biochem 1988 Nov;26(11):783-790.

For further information, please refer to the appropriate operator's manual for the analyzer concerned, the respective application sheets, the product information and the Method Sheets of all necessary components (if available in your country).

A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Symbols

Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard (for USA: see https://usdiagnostics.roche.com for definition of symbols used):

CONTENT	Contents of kit
SYSTEM	Analyzers/Instruments on which reagents can be used
REAGENT	Reagent
CALIBRATOR	Calibrator
\longrightarrow	Volume after reconstitution or mixing
GTIN	Global Trade Item Number

COBAS, COBAS E, ELECSYS and PRECICONTROL are trademarks of Roche. INTRALIPID is a trademark of Fresenius Kabi AB.

All other product names and trademarks are the property of their respective owners Additions, deletions or changes are indicated by a change bar in the margin.

© 2018, Roche Diagnostics



Roche Diagnostics GmbH, Sandhofer Strasse 116, D-68305 Mannheim www.roche.com

